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Tart cherry juice has no acute effects on uric acid, vascular function and inflammation: a randomised crossover trial.

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Tart cherry juice has no acute effects on uric acid, vascular function and inflammation: a randomised crossover trial.

Abstract

Background

Hyperuricaemia increases the risk of gout, and cardiovascular disease, thus dietary modifications that reduce urate are of interest. Cherries have been reported to lower urate, but studies examining acute effects have mostly failed to include a control group, despite urate being known to exhibit diurnal fluctuations, typically falling throughout the day.

Aim

This study aimed to determine the acute effects of a single serving of tart cherry juice on uric acid metabolism and risk factors for cardiovascular disease relative to a control drink.

Methods

In an open-label, randomised, controlled, crossover design, 12 healthy adults (mean age 41.1 (± 11.1) y; mean body mass index 26.4 (± 4.3) kg/m²; 7 men and 5 women) consumed 250 mL tart cherry juice (containing 30 mL of concentrate) and 250 mL water (control) on separate occasions ≥ 7 days apart. Serum uric acid, central and brachial blood pressure, augmentation index, and pulse wave velocity were measured at baseline, 1, 2, 3, 5, and 24 hours, post-drink, serum c-reactive protein at baseline, 2 and 5 hours, and creatinine-adjusted urinary uric acid at 0-2, 2-4, and 4-5 hours.

Results

There were no statistically significant main effects of drink type or drink by time interactions (all outcomes $p > 0.05$). However, independent of drink type, serum uric acid ($p = 0.008$), urinary uric acid ($p < 0.001$), c-reactive protein ($p = 0.023$), and measures of blood pressure (all $p < 0.05$) changed with different temporal patterns throughout the day (main effects of time, $p < 0.05$).

Conclusion

These results indicate that diurnal fluctuations may partly explain the beneficial acute effects of cherry consumption on uric acid metabolism and inflammation previously reported in studies without a comparator control group.

Trial registry name and URL: ClinicalTrials.gov <https://clinicaltrials.gov/study/NCT04960527>

Trial Registration number: (NCT04960527)

Keywords: cherry juice; uric acid; inflammation; blood pressure; vascular function.

Introduction

Hyperuricaemia has been associated with elevated risk of gout, renal disease, cardiovascular disease (CVD) and metabolic dysfunction (Kuo et al., 2016; Terkeltaub et al., 2006). Several

46 dietary modifications have been proposed for the prevention of hyperuricaemia, including
47 restricting intakes of purine-rich and fructose-rich foods, limiting alcohol consumption,
48 remaining hydrated, and increasing cherry consumption (Collins et al., 2019; Schlesinger,
49 2005). The potential of cherries to prevent hyperuricaemia has been ascribed to their high
50 content of polyphenols, especially anthocyanins (Chaovanalikit and Wrolstad, 2004; Kelley et
51 al., 2018; Kirakosyan et al., 2009). Cherry consumption has been suggested to reduce serum
52 uric acid (sUA) by: (i) inhibiting hepatic xanthine oxidoreductase and/or (ii) increasing the
53 glomerular filtration of UA and inhibiting its tubular reabsorption, thereby increasing urinary
54 uric acid (UUA) excretion (Haidari et al., 2009; Jacob et al., 2003; Kirakosyan et al., 2018;
55 Zhang et al., 2012).

56 To our knowledge, three studies have reported that cherries decrease sUA in the hours after
57 consumption (Bell et al., 2014a; Hillman and Uhranowsky, 2021; Jacob et al., 2003), two of
58 which (Bell et al., 2014a; Jacob et al., 2003) also measured an increase in UUA excretion.
59 However, these studies had methodological limitations. Two studies (Bell et al., 2014a; Jacob
60 et al., 2003) had no control group. Whereas the third, which investigated the effect of one and
61 two daily servings of tart cherry (TC) in capsules and as juice, only contained placebo groups
62 for the once daily servings (Hillman and Uranowsky, 2021). Since sUA is known to exhibit a
63 diurnal rhythm, falling as the day progresses (Sennels et al., 2012), the failure to include control
64 groups complicates the interpretation of these studies. Thus, the primary aim of the present
65 study was to determine the acute effects of TC juice consumption on sUA and UUA excretion
66 relative to a control drink.

67 Hyperuricemia is a risk factor for CVD possibly because it promotes hypertension and
68 increases arterial stiffness (Borghi et al., 2022; An et al., 2024). Within vascular endothelial
69 cells, elevated UA promotes oxidative stress, inflammation, and depletes nitric oxide causing
70 endothelial dysfunction and vasoconstriction (Ndrepepa 2025). The consumption of TC might

be expected to reduce blood pressure (BP) and arterial stiffness by lowering urate or via the anti-inflammatory and antioxidant actions of its content of polyphenols. However, the results of human intervention studies investigating the effect of TC on BP and arterial stiffness have been mixed (Desai et al., 2021; Keane et al., 2016a; Kimble et al., 2021; Lynn et al., 2014). Thus, the secondary aim of this study was to determine the acute effects of TC juice on inflammation, blood pressure (BP), and arterial stiffness.

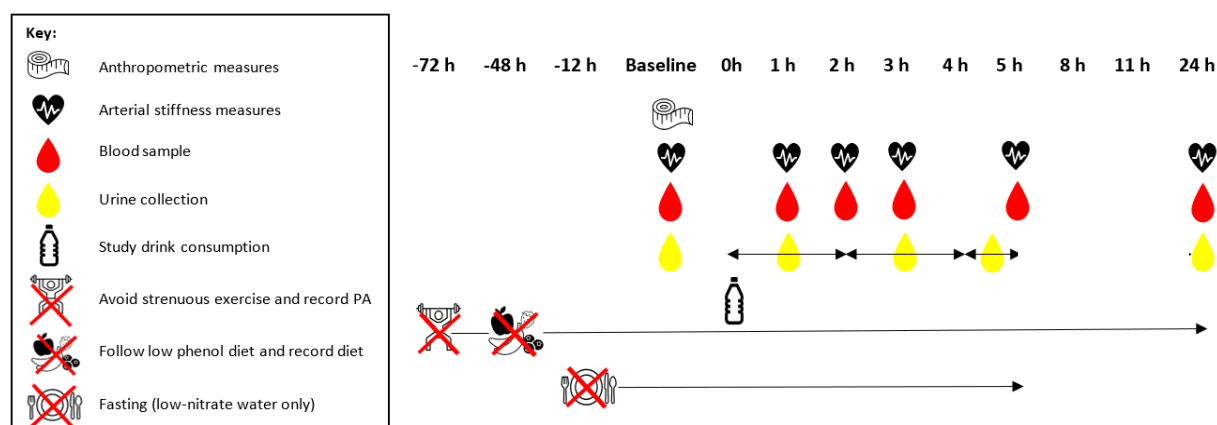
Methods

This study was reported in line with the CONSORT 2010 statement (Dwan et al., 2019).

Trial Design

The study was an open-label, 2-arm, randomised, placebo-controlled, crossover trial of thirteen healthy adults. Participants consumed 250 mL of TC juice (30 mL TC concentrate with 220 mL water) or 250 mL of water on two separate occasions, separated by a wash-out period of \geq 7 days. Blood, urine, and vascular measurements were collected at baseline and multiple time-points over 24 hours following each drink (**Fig 1**). Each participant attended each of their test sessions at the same time (between 9 and 10am).

Fig. 1 Study protocol. PA; physical activity



The study opened recruitment in July 2021 and closed at the end of February 2022. It was approved by Sheffield Hallam University (SHU) ethics committee (ER9199256) and registered at ClinicalTrials.gov (NCT04960527) before recruitment commenced. The study was conducted in accordance with the 1964 Declaration of Helsinki and its later amendments.

Participants and settings

A total of 13 healthy, non-smoking, adult volunteers were recruited through word-of-mouth. Inclusion criteria were, aged between 18 and 85 years, and no history of, gout, type 1 or type 2 diabetes, gastrointestinal disorders, CVD, or kidney disease. Interested individuals were provided with a participant information sheet containing further details of the study. Potential participants also received a verbal explanation of the study and were screened for inclusion criteria. Written informed consent was gained from all participants. Measurements were made at the Nutrition Research Laboratory of SHU, Sheffield, United Kingdom, United Kingdom.

Dietary Interventions

During the active intervention arm of the study, participants consumed 250 mL of TC juice, consisting of 30 mL Montmorency TC concentrate (CherryActive®, ActiveEdge™, Hanworth, UK) and 220 mL low-nitrate water (Buxton®, UK). Analysis of the TC concentrate in our

nutrition research laboratory revealed that each serving contained a mean phenol content of 408 (SD 5.4) mg gallic acid equivalents (Folin-Ciocalteu method; Singleton and Rossi, 1965) and an anthocyanin content of 3.8 (SD 0.3) mg cyanidin-3-glucoside equivalents (pH differential method; Lee et al., 2005). During the control arm, participants consumed 250 mL of low-nitrate water (Buxton®, UK). A low-nitrate water was selected for the control drink to minimise vascular effects (Hobbs et al., 2013) and avoid bioactive compounds such as polyphenols. The two study arms were separated by a wash-out period of ≥ 7 days. The wash-out duration was based on the known pharmacokinetics of cherry polyphenols (Keane et al., 2016b) and likely transient nature of any effects on the outcome measures. An investigator not involved in data collection generated a block randomised allocation sequence using www.random.org (block size 4) and assigned participants to their sequence of interventions. The use of a water control meant it was impossible to conceal this sequence from participants or the researcher collecting data.

Participants were provided with a dietary advice sheet containing meal recommendations to help them follow a low-polyphenolic diet, including avoiding fruits, vegetables, wholegrains, and nuts, for 48 hours prior to each test day. The evening prior to each test day participants were provided with a low-phenol spaghetti carbonara ready-meal (Sainsbury's PLC, UK), low-phenol dessert (Bonne Maman®, Somerset, UK), and low-nitrate water (Buxton®, UK) to consume. Participants attended the laboratory following an overnight fast of ≥ 10 hours, although low-nitrate water was permitted.

Participants remained fasted during the first 5 hours post drink consumption; however, 500 mL low-nitrate water was provided during this time. Participants were advised to drink when thirsty but avoid consuming large volumes at a single time-point to minimise possible effects on vascular function (Callegaro et al., 2007). A low-phenol lunch of sandwiches made from white bread and ham, ready salted crisps, and a plain Greek yoghurt were provided

immediately following the 5-hour measurements. Participants were also provided with low-phenol snacks, a low-phenol macaroni cheese ready-meal, and low-nitrate water to consume over the rest of the day. Participants returned to the laboratory following another overnight fast of ≥ 10 hours for their 24-hour measurements.

Participants were asked to record their dietary intake throughout the first arm of the study and instructed to replicate this during the second arm. Participants were also asked to avoid strenuous exercise from 72 hours before each test day until after their 24-hour measurements.

Outcomes

The primary outcome measure was between-treatment difference in the change in sUA from baseline to 24 hours post-drink. Secondary outcome measures were between-treatment differences in the change in the inflammatory marker, serum CRP from baseline to 5 hours post-consumption, and changes in UUA excretion and vascular function (resting brachial and central BP and arterial stiffness) from baseline to 24 hours post-consumption. Non-efficacy outcomes included physical activity (PA) and dietary intake measures, for example consumption of high-phenolic foods.

Anthropometry

Height (to 0.1 cm) and mass (to 0.1 kg) of participants were measured during their first visit to the laboratory and used to calculate body mass index (BMI) (mass (kg)/height (m)²).

Arterial Stiffness and Blood Pressure (BP)

A Vicorder® device (SMT Medical, Germany) was used to measure brachial and central BP, carotid-femoral PWV, and augmentation index (AIx). Participants were familiarised with the Vicorder® prior to their first experimental session to reduce the effects of anxiety on BP and

other vascular measures (Franklin et al., 2013). Familiarisation consisted of practice measurements with carotid, arm, and femoral cuffs, so participants could become accustomed to the sensation of each cuff inflating. BP, PWV, and AIx were measured at baseline and 1, 2-, 3-, 5-, and 24-hours post-drink consumption. Following the Vicorder® instructions, brachial DBP values were also used as central DBP values. Three replicate measures with 1-minute intervals were taken at each time-point. Participants rested in a supine position for 15 minutes before the measurements and remained still throughout.

Collection and Processing of Blood Samples

Blood was collected at baseline, 1, 2, 3, 5, and 24 hours, post-consumption. Samples were centrifuged at 2500 x g for 15 minutes at 18 °C to separate serum (Hermle Z 36 HK, HERMLE Labortechnik GmbH, Germany), which was stored at -80°C until analysis.

Serum C-reactive Protein (CRP) and Uric Acid (sUA)

CRP was measured in serum collected at baseline, 2, and 5 hours using a CRP Quantikine enzyme-linked immunosorbent assay kit (R&D systems, Abingdon, UK). The intra-assay CV was 5.5%. sUA was determined in serum collected at baseline, 1, 2, 3, 5, and 24 hours using a UA (Amplex® Red, Invitrogen™, UK) assay kit. The intra-assay CV was 3.9%. Both analytes were measured on a microplate reader (BioTek synergy HT, Winooski, USA).

Urine collection and analysis

Spot urine samples were collected at baseline and 24 hours post-drink consumption. Urine was also collected between 0-2, 2-4, and 4-5 hours. Samples were centrifuged twice at 2800 x g for 15 minutes to remove unwanted cells and material (Hermle Z 36 HK, HERMLE Labortechnik GmbH, Germany) and stored at -80 °C until analysis. Urine samples were analysed for UUA (Amplex® Red, Invitrogen™, UK) and creatinine (ELISA; R&D systems, Abingdon, UK)

concentrations, using a microplate reader (BioTek synergy HT, Winooski, USA). The intra-assay CV was 2.0% for UUA and 2.1% for urinary creatinine. UUA (μmol) was corrected for creatinine concentration (μMol) to provide a UUA to urinary creatinine excretion ratio.

Assessment of Diet and Physical Activity (PA)

From 48 hours prior to baseline until 24-hour post-consumption, participants completed a food diary. Participants recorded PA in a diary from 72 hours prior to their two main laboratory sessions until their 24-hour post-consumption measurements.

Statistical Methods

The primary outcome was change in sUA concentration. Change in UUA, CRP, BP, and arterial stiffness were secondary outcomes. The effect of treatment (TC versus water) on all outcomes was analysed as the percentage change from baseline using two-way repeated measures analyses of variance (ANOVA) with Bonferroni post-hoc tests. Partial Eta-Squared (η^2) effect sizes for ANOVA were classified as small (0.01 – 0.059), moderate (0.06-0.137), and large (≥ 0.138) (Pallant, 2010). Further exploratory analyses investigating between-sex differences on the effect of cherry consumption on sUA and UUA were undertaken by adding sex as a between-subjects factor in two-way repeated measures ANOVAs. Baseline data is presented as mean (SD) or median and interquartile range (IQR), as appropriate. Results are reported as mean % and SD for continuous data. All analyses were conducted using IBM SPSS Statistics v24. The critical value for statistical significance was set at $p < 0.05$.

A sample size of thirteen was determined sufficient to detect a decrease in sUA of 15 $\mu\text{mol/L}$ with 80% power at a significance level of 0.05, using data from White et al. (2018).

Results

Participant Characteristics

Thirteen participants started the study however one dropped out following completion of the control treatment. Of the 12 participants (7 male/5 female) who completed the study, mean age was 41.1 (\pm 11.1; range 27 to 60) years, and average BMI 26.4 (\pm 4.3; range 20.1 to 35.1) kg/m². Baseline clinical data did not differ between TC juice and control drink visits ($p > 0.05$ for all), **Table 1**.

Table 1 Baseline clinical data of participants prior to the provision of 250 mL tart cherry juice and 250 mL water. Data are presented as mean \pm SD or median (IQR)

	Tart cherry juice	Water (control)
Brachial systolic blood pressure, mmHg	126.2 \pm 11.0	123.6 \pm 8.4
Brachial diastolic blood pressure, mmHg	65.8 \pm 6.1	64.9 \pm 6.4
Central systolic blood pressure, mmHg	120.3 \pm 11.8	118.7 \pm 8.6
Pulse wave velocity, m/s	8.2 (2.6)	7.3 (2.0)
Augmentation index, % ^a	14.8 \pm 7.3	18.2 \pm 9.8
Serum uric acid, μ mol/L	155.4 \pm 59.2	168.3 \pm 57.1
Urinary urate:urinary creatinine, mmol/mmol	0.4 \pm 0.1	0.5 \pm 0.2
C-reactive protein, mg/L	0.4 (0.7)	0.6 (1.8)

$n = 12$ for both treatment arms except for ^a where $n = 10$.

Dietary Adherence and Avoidance of High-Intensity Physical Activity

Evaluation of participants' diet and PA diaries indicated that participants complied with the low-phenol diet and physical activity guidance.

213 *Serum Uric Acid (sUA)*

214 There was a large-sized main effect of time on sUA following consumption of the drinks ($F_{5,55}$
215 $= 3.529$, $p = 0.008$, $\eta^2 = 0.243$) with a mean 10.4% reduction in sUA between 1 hour and 5
216 hours post-consumption ($p = 0.034$) and a mean 8.5% increase from 5 hours to 24 hours post-
217 consumption ($p = 0.022$) (**Fig 2a**, Table 2). However, no drink type ($F_{1,11} = 2.061$, $p = 0.179$,
218 $\eta^2 = 0.158$) or drink by time interaction ($F_{5,55} = 1.222$, $p = 0.311$, $\eta^2 = 0.100$) effects were
219 found. Furthermore, there were no between-sex differences in response ($F_{5,55} = 1.151$, $p =$
220 0.347 , $\eta^2 = 0.103$).

221 *Urinary Uric Acid (UUA)*

222 As shown in **Fig 2b** (and Table 2), creatinine-adjusted UUA fluctuated significantly over time
223 and this main effect was large ($F_{4,44} = 11.656$, $p < 0.001$, $\eta^2 = 0.514$). The greatest increase in
224 UUA above baseline (0 hours) was observed at 2-4 hours (41.0 %; $p = 0.001$), followed by 0-
225 2 hours (32.2 %; $p < 0.001$). UUA was significantly lower at 24 hours than at 0-2 hours ($p <$
226 0.001) and 2-4 hours ($p = 0.005$). There were no statistically significant main effects of drink
227 type ($F_{1,11} = 0.015$, $p = 0.906$, $\eta^2 = 0.001$) or drink by time interaction ($F_{4,44} = 1.084$, $p = 0.358$,
228 $\eta^2 = 0.090$). There were also no between-sex differences in response ($F_{4,44} = 1.397$, $p = 0.263$,
229 $\eta^2 = 0.123$).

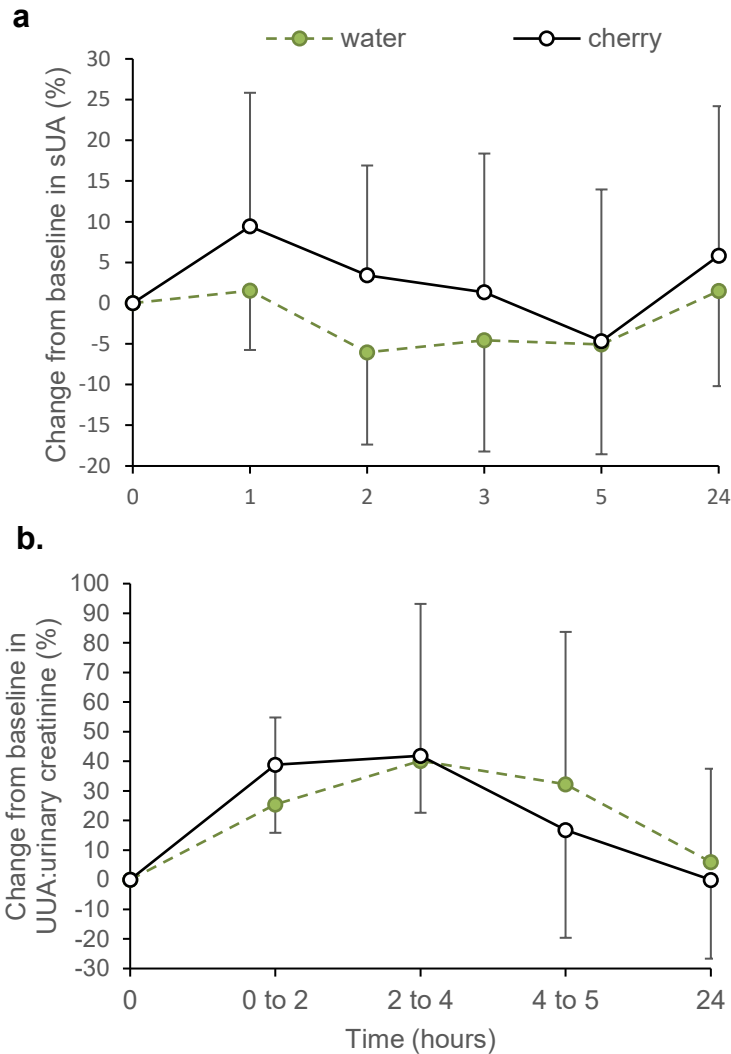


Fig. 2 Effect of tart cherry juice and water on percentage change from baseline values in **a)** serum uric acid (sUA) concentration and **b)** urinary uric acid (UUA) to urinary creatinine ratio. Data are presented as mean \pm SD, $n = 12$ for both outcomes.

C-reactive Protein

There was a large-sized main effect of time for change in CRP from baseline ($F_{2,22} = 4.488$, $p = 0.023$, $\eta^2 = 0.290$), with a statistically significant 7.4 % reduction between 2 hours and 5 hours ($p = 0.020$) (**Fig 3** and Table 2). Despite this, CRP at 5 hours was not significantly different from baseline ($p = 0.202$) and no main effect of drink type ($F_{1,11} = 0.434$, $p = 0.524$, $\eta^2 = 0.038$) or drink by time interaction ($F_{2,22} = 0.644$, $p = 0.525$, $\eta^2 = 0.055$) were detected.

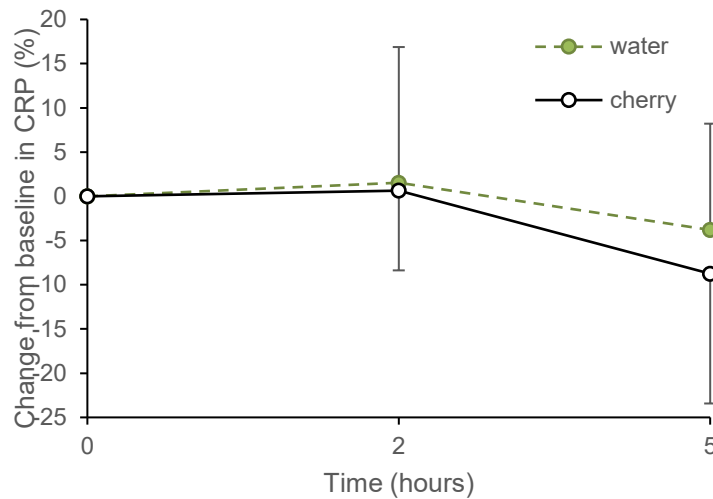


Fig. 3 Effect of tart cherry juice and water on percentage change in c-reactive protein (CRP) concentration from baseline values. Data are presented as mean \pm SD, $n = 12$.

Blood Pressure

Brachial Systolic BP (SBP)

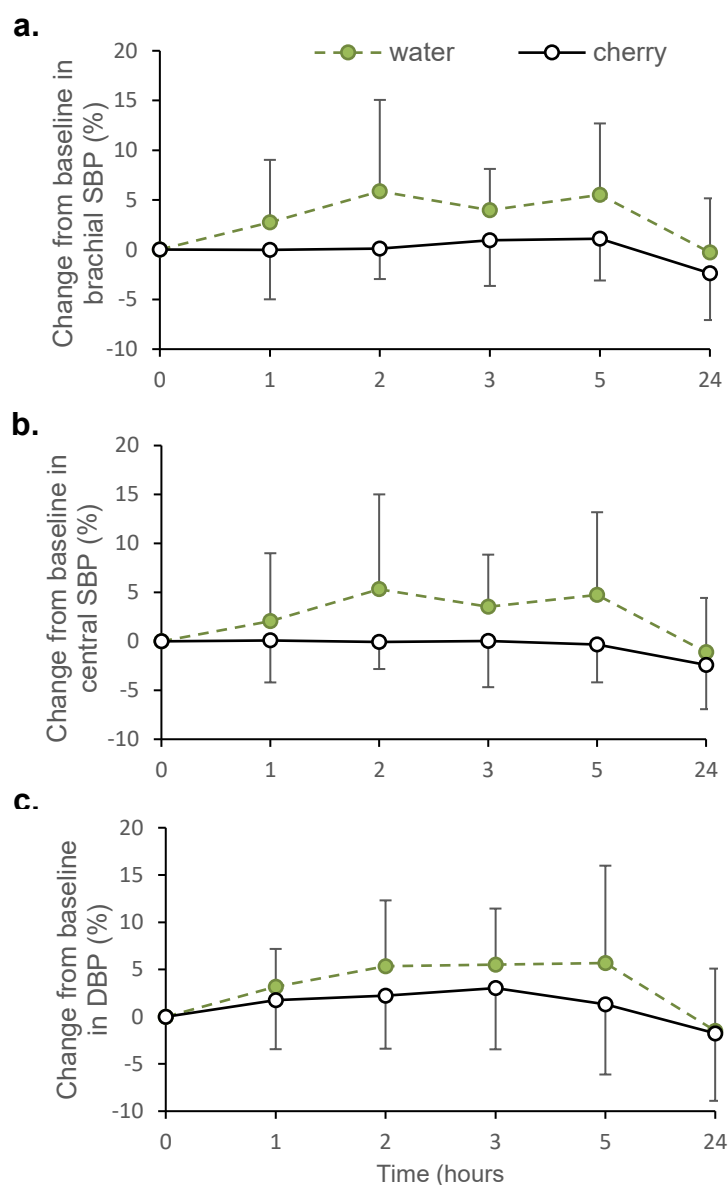
A large main effect of time ($F_{5,55} = 5.360$, $p < 0.001$, $\eta^2 = 0.328$) was detected for brachial SBP (**Fig 4a**, and Table 2), with a mean reduction of 4.6 % between 5 hours and 24 hours ($p < 0.001$). There was a non-significant large main effect of drink ($F_{1,11} = 3.654$, $p = 0.082$, $\eta^2 = 0.249$); estimated marginal mean for brachial SBP was 3.0 % (95% CI -6.5, 0.5) lower in the water arm than in the TC juice arm. No drink by time interaction effect ($F_{5,55} = 1.459$, $p = 0.218$, $\eta^2 = 0.117$) was observed.

Central Systolic BP (SBP)

A large main effect of time ($F_{5,55} = 3.403$, $p = 0.009$, $\eta^2 = 0.236$) was detected for central SBP, with a 4.0 % reduction observed between 5 hours and 24 hours ($p = 0.014$) (**Fig 4b**, Table 2). There was no drink by time interaction effect ($F_{5,55} = 1.866$, $p = 0.154$, $\eta^2 = 0.145$) or main effect of drink type ($F_{1,11} = 3.234$, $p = 0.100$, $\eta^2 = 0.227$).

254 *Brachial and Central Diastolic BP (DBP)*

255 Brachial DBP values were used as central DBP values, in accordance with the Vicorder®
 256 instructions. There was a large significant main effect of time ($F_{5,55} = 5.908$, $p < 0.001$, $\eta p^2 =$
 257 0.349) for DBP (**Fig 4c**, Table 2). On average, DBP fell by 5.4 % between 2 hours and 24 hours
 258 ($p = 0.027$) and by 5.9 % between 3 hours and 24 hours ($p = 0.001$). There was no drink by
 259 time interaction effect ($F_{5,55} = 0.718$, $p = 0.612$, $\eta p^2 = 0.061$) or main effect of drink type ($F_{1,11}$
 260 $= 1.782$, $p = 0.209$, $\eta p^2 = 0.139$).



261 **Fig. 4** Effect of tart cherry juice and water on percentage change from baseline values in **a)**
 262 brachial systolic blood pressure (SBP), **b)** central systolic blood pressure (SBP), and **c)** brachial

263 and central diastolic blood pressure (DBP). Data are presented as mean (\pm SD); $n = 12$ for all
264 outcomes.

265 *Arterial Stiffness*

266 *Pulse Wave Velocity (PWV)*

267 Carotid-femoral PWV fluctuated over the measurement period (Table 2), however, these
268 fluctuations were not statistically significant (time: $F_{5,50} = 0.493$, $p = 0.667$, $\eta p^2 = 0.047$). There
269 were no main effects of drink type ($F_{1,10} = 1.948$, $p = 0.193$, $\eta p^2 = 0.163$) or drink by time
270 interaction ($F_{5,50} = 1.257$, $p = 0.297$, $\eta p^2 = 0.112$), ($n = 12$).

271 *Augmentation Index (AIx)*

272 AIx was measured in ten of the twelve participants. For two participants, AIx could not be
273 consistently measured due to low pulse amplitude so these participants were excluded from
274 analysis. There were no main effects of time ($F_{5,45} = 1.819$, $p = 0.204$, $\eta p^2 = 0.168$) or drink
275 type ($F_{1,9} = 2.688$, $p = 0.136$, $\eta p^2 = 0.230$), and no drink by time interaction ($F_{5,45} = 1.085$, $p =$
276 0.344 , $\eta p^2 = 0.108$) for AIx (Table 2).

277

278 **Harms**

279 No adverse effects were reported.

280 **Table 2** Acute effects of 250 ml tart cherry juice versus water (control) on vascular function, inflammation and urate¹.

Outcome	Study drink	Time (hours)					
		Baseline	1	2	3	5	24
Brachial systolic blood pressure, mmHg*	Cherry	126.2 ± 11.0	126.1 ± 12.4	126.2 ± 9.8	127.2 ± 10.8	127.4 ± 10.0	122.9 ± 9.0
	Water (control)	123.6 ± 8.4	126.9 ± 11.3	130.6 ± 12.2	128.3 ± 8.2	130.2 ± 9.7	123.0 ± 7.9
Brachial and central diastolic blood pressure, mmHg*	Cherry	65.8 ± 6.1	66.9 ± 5.8	67.1 ± 5.1	67.6 ± 5.4	66.6 ± 6.8	64.4 ± 4.5
	Water (control)	64.9 ± 6.4	67.0 ± 6.9	68.3 ± 6.9	68.3 ± 6.9	68.3 ± 6.0	63.7 ± 4.2
Central systolic blood pressure, mmHg*	Cherry	120.3 ± 11.8	120.4 ± 12.7	120.1 ± 10.8	120.2 ± 12.1	119.7 ± 10.0	117.1 ± 8.9
	Water (control)	118.7 ± 8.6	121.2 ± 12.8	124.9 ± 13.7	122.8 ± 10.1	124.1 ± 10.9	117.2 ± 7.9
Pulse wave velocity, m/s,	Cherry	7.9 ± 1.4	7.2 ± 1.2	7.3 ± 1.7	7.8 ± 1.4	7.4 ± 1.4	7.2 ± 1.4
	Water (control)	8.1 ± 2.8	7.6 ± 1.2	7.4 ± 1.4	7.2 ± 1.1	7.4 ± 0.8	7.3 ± 1.4
Augmentation index, %	Cherry	14.8 ± 7.3	14.9 ± 7.4	14.7 ± 7.5	14.5 ± 8.9	14.3 ± 8.0	16.1 ± 6.7
	Water (control)	18.2 ± 9.8	15.5 ± 7.7	15.2 ± 8.2	15.2 ± 9.0	15.2 ± 8.7	16.8 ± 7.7

Serum uric acid, μmol/L*	Cherry	155.4 ± 59.2	170.2 ± 70.3	163.3 ± 77.6	160.2 ± 76.4	149.9 ± 74.0	165.8 ± 78.9
	Water (control)	168.3 ± 57.1	171.0 ± 60.3	159.3 ± 59.6	164.0 ± 69.3	163.8 ± 71.8	169.7 ± 61.4
C-reactive protein, mg/L,*	Cherry	0.69 ± 0.61		0.68 ± 0.60		0.61 ± 0.54	
	Water (control)	1.16 ± 1.45		1.13 ± 1.43		1.04 ± 1.21	
		<div>0-22-44-524</div>					
Urinary urate:urinary creatinine, mmol/mmol *	Cherry	0.42 ± 0.13		0.57 ± 0.15	0.58 ± 0.14	0.47 ± 0.15	0.41 ± 0.12
	Water (control)	0.46 ± 0.16		0.55 ± 0.17	0.61 ± 0.23	0.58 ± 0.19	0.46 ± 0.21

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296 ¹All values are means ± SD; *n* = 12 for all outcomes except augmentation index were *n* = 10. There were no significant main effects of drink type or drink x
297 time interactions. * indicates a significant main effect of time (*p* < 0.05)

Discussion

The primary aim of the present study was to investigate the effect of TC juice on sUA. In contrast to other studies, we did not find evidence that TC juice reduces sUA for up to 24 hours post-consumption. We also found no evidence that TC juice acutely increases UUA excretion or lowers inflammation, BP, or arterial stiffness.

The failure of TC juice to reduce sUA and increase UUA excretion contrasts with previous studies of sweet and tart cherries. Jacob et al. (2003) reported that 280 g of sweet cherries and Bell et al. (2014a) reported that 30 mL and 60 mL of TC concentrate (diluted with 100 mL of H₂O) lowered sUA and increased UUA excretion post-consumption in healthy adults. This contrast may be partly explained by the failure of Jacob et al. (2003) and Bell et al. (2014a) to include a control group, because sUA has previously been reported to fall from morning onwards (Sennels et al., 2012), a phenomenon observed in our participants after consumption of both placebo and TC drinks. The maximal decrease observed in sUA of 10% and increase in UUA excretion of 41% was comparable to that reported by Jacob et al. (2003) (sUA -14% & UUA +69%) in women with similar baseline sUA to our participants, but much lower than reported by Bell et al. (2014a) (sUA -36% and UUA +250%), but their participants had much greater baseline sUA (approximately 480 μ mol/L) than our participants (162 μ mol/L), despite being described as healthy young adults. Notwithstanding the difficulty of interpreting the results of Bell et al. (2014a) because of the lack of a control arm, it is possible that the UA lowering effect of TC juice may partly depend on baseline sUA. Hillman & Uhranowsky (2021) reported that one and two daily servings of TC in powdered form (480 mg per capsule) and two daily servings of TC juice reduced sUA over a 48-hour period, whereas one daily serving of TC juice (30 mL of concentrate diluted to 240 mL with H₂O) seemed ineffective leading to a small increase in sUA at 8 hours post-consumption. The lack of benefit of a single daily serving of TC juice containing 30 mL of concentrate is broadly consistent with

our findings. Interpretation of the results of Hillman & Uhranowsky (2021) is complicated by the inclusion of apple juice in their TC drinks, because it is known to increase sUA (White et al., 2018), and lack of clarity whether reported treatment effects are in comparison to a placebo (and if so, which of the two placebos in their study) or within arm baseline values. Our findings challenge the results of previous acute studies reporting beneficial effects of cherries on urate metabolism and highlight the need for future studies to include a control group. However, our participants were healthy and there is a need to confirm whether TC acutely alters urate metabolism in individuals with elevated sUA such as those suffering from gout.

Processed TC products (Ou et al., 2012), whole TC extracts (Seeram et al., 2001), or anthocyanins found in TC, namely cyanidin-3-glucosylrutinoside and cyanidin-3-rutinoside (Wang et al., 1999) have been shown to exert anti-inflammatory effects *in vitro* (Virgen Gen et al., 2020), reduce exercise-induced inflammation (Bell et al., 2014b, 2015, 2016, Dimitriou et al., 2015; Howatson et al., 2009; Levers et al., 2016), and lower serum CRP for up to 5 hours after consumption in an uncontrolled study of purportedly healthy young adults with raised baseline CRP (Bell et al., 2014a). In contrast, we failed to observe a significant difference between TC and water with CRP falling between 2 and 5 hours after the consumption of both drinks. This finding demonstrates the difficulty of interpreting results from uncontrolled studies. This is further illustrated by studies with longer intervention periods. For example, in healthy adults with normal CRP at baseline (Lynn et al., 2014) and obese adults with raised CRP at baseline (Martin et al., 2018) TC failed to lower CRP, relative to control groups, whereas an uncontrolled study reported that sweet cherry consumption lowered serum CRP after 14 and 28 days (Kelley et al., 2006).

The consumption of cherries has been proposed to reduce BP by altering the synthesis and activity of vasodilators and vasoconstrictors (Kelley et al., 2018). However, we observed no effect of TC juice on brachial or central BP or measures of arterial stiffness. The lack of

modulation of BP contrasts with two studies that reported that TC reduced SBP for up to 3 hours post consumption in men with early hypertension (Keane et al., 2016a) and middle-aged adults with moderately raised SBP (Keane et al., 2016c). The disagreement with our study may be explained by the lower baseline BP of our participants. In a review of factors influencing the effects of dietary anthocyanins on the regulation of BP, elevated baseline BP was highlighted as a major determinant of whether anthocyanins exerted hypotensive effects (Vendrame and Klimis-Zacas, 2019). The quantity of TC juice would also be expected to be important. Keane et al. (2016a) and Keane et al. (2016c) administered 60 mL of TC concentrate whereas we used 30 mL of concentrate. However, 30 mL is the typical suggested serving size for TC concentrate and therefore may be the amount commonly drunk by consumers. In agreement with our study, Desai et al. (2021) observed no acute effect of a single 30 mL serving of TC concentrate on SBP in individuals with metabolic syndrome, but they did report that 24 h ambulatory BP was reduced at the end of a 7-day intervention period. Thus, it is possible that a longer duration of intake is needed for 30 mL servings of TC concentrate to lower BP, although Lynn et al. (2014) failed to find an effect of 30 mL/d of TC concentrate consumed for 4 weeks by normotensive adults when BP was measured at laboratory visits.

TC might be expected to reduce arterial stiffness via urate lowering or through putative anti-oxidant and anti-inflammatory effects. However, we found no significant differences in PWV or AIx between TC and water over the 24-hour measurement period. This is in line with Keane et al. (2016a) who reported no effect of 60 mL of TC concentrate on PWV or AIx over an 8-hour measurement period. The lack of an acute effect of TC juice on PWV and AIx is consistent with studies of other polyphenol rich fruits (Del Bo et al., 2014; Richter et al., 2017; Rodriguez-Mateos et al., 2013, 2016), indicating that neither measure of arterial stiffness is particularly amenable to rapid modulation by polyphenol rich fruits.

This study has several limitations. First, our participants were apparently healthy, and TC might only benefit individuals with elevated sUA and markers of cardiovascular risk. Second, it is possible that TC juice may have changed some outcome markers outside the time-period we took measurements. Third, the TC concentrate we used may not have supplied sufficient bioactive compounds to exert an effect. Whilst our analyses of the TC concentrate revealed that a serving supplied a dose of total phenols within the range shown in other TC interventions to exert physiological effects (Keane et al. 2016a; Connolly et al. 2006), its content of intact anthocyanins was relatively low (Martin & Coles 2010; Bell et al. 2014a). This could partly explain our null findings if intact anthocyanins are the primary compounds driving the biological actions of TC. Fourth, the study was not blinded, but this might have been expected to increase the likelihood of finding a treatment effect for an outcome such as BP which is particularly susceptible to the placebo effect (Howard et al., 2016). Fifth, although each participant arrived at the same time for both of their study visits we could not control their wake time, which could have introduced variability into our measurements given that many have been reported to exhibit diurnal patterns (Shimizu et al. 2023; Hernandez et al. 2024;). Sixth, the final sample size was one less than the pre-determined sample size, because one participant dropped out. It is unlikely however that one more participant completing the study would have meaningfully changed the outcome for sUA, because the difference between TC and control was not close to statistically significant. The sample size was determined to detect a change in sUA so the study may have been underpowered to detect changes in other outcomes. Also, we did not power the study to investigate between sex differences in the response of urate metabolism to TC so the results of these analyses should be interpreted cautiously.

This study has several strengths. First, unlike some studies reporting a UA lowering effect of cherries (Bell et al., 2014a; Jacob et al., 2003), there was a control group. Second, diet

was controlled during the study by giving participants standardised meals low in polyphenols the evening before and during the 24-hour measurement periods and by providing clear guidance on consuming a diet low in polyphenols for the duration of the study. Third, the low-nitrate water control drink was devoid of factors that could influence uric acid metabolism and vascular function such as fructose and nitrate (Hobbs et al., 2013).

In conclusion, the present study found no evidence that a single serving of TC benefits urate metabolism, inflammation, or markers of vascular function in healthy adults compared with a control drink of water. However, following consumption of both TC and water, changes in urate metabolism, inflammation and BP occurred over the 24-hour measurement period likely reflecting diurnal fluctuations. Our findings need to be considered when interpreting the results of previous uncontrolled studies that have reported beneficial acute effects of TC juice or sweet cherries in healthy adults. Future controlled studies are needed to determine whether TC consumption exerts beneficial acute effects in individuals with hyperuricaemia or gout.

Ethical Statements

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Availability of data: The data that support the findings of this study are available from the corresponding author, AL, upon reasonable request.

Conflict of interest: AL has previously been a recipient of a research grant from the Cherry Marketing Institute, Michigan, USA. The authors report no further conflicts of interest.

Consent: Written informed consent was obtained from all study participants.

Ethical approval: The study was approved by the ethics committee of Sheffield Hallam University, UK.

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422 **References**

- 423 An, L., Wang, Y., Liu, L. et al. (2024) High serum uric acid is a risk factor for arterial
424 stiffness in a Chinese hypertensive population: a cohort study. *Hypertension*
425 *Research* 47: 1512–1522 <https://doi.org/10.1038/s41440-024-01591->
- 426 Bell PG, Gaze DC, Davison GW et al. (2014a) Montmorency tart cherry (*Prunus*
427 *cerasus* L.) concentrate lowers uric acid, independent of plasma cyanidin-3-O-
428 glucosiderutinoside. *Journal of Functional Foods* 11: 82–90.
429 <https://doi.org/10.1016/j.jff.2014.09.004>
- 430 Bell PG, Walshe IH, Davison GW et al. (2014b) Montmorency cherries reduce the
431 oxidative stress and inflammatory responses to repeated days high-intensity stochastic
432 cycling. *Nutrients* 6(2): 829–843. <https://doi.org/10.3390/nu6020829>
- 433 Bell PG, Walshe IH, Davison GW et al. (2015) Recovery facilitation with
434 Montmorency cherries following high-intensity, metabolically challenging exercise.
435 *Applied Physiology, Nutrition, and Metabolism* 40(4): 414–423.
436 <https://doi.org/http://dx.doi.org/10.1139/apnm-2014-0244>
- 437 Bell PG, Stevenson E et al. (2016) The effects of montmorency tart cherry concentrate
438 supplementation on recovery following prolonged, intermittent exercise. *Nutrients*
439 8(441): 1–11. <https://doi.org/10.3390/nu8070441>
- 440 Borghi C, Agnoletti D, Cicero AFG et al. (2022) Uric Acid and Hypertension: a Review
441 of Evidence and Future Perspectives for the Management of Cardiovascular Risk.
442 *Hypertension*. 79(9):1927-1936.
443 <https://doi.org/10.1161/HYPERTENSIONAHA.122.17956>.

Callegaro CC, Moraes RS, Negrão CE et al. (2007) Acute water ingestion increases arterial blood pressure in hypertensive and normotensive subjects. *Journal of Human Hypertension* 21(7): 564–570. <https://doi.org/10.1038/sj.jhh.1002188>

Chaovanalikit A and Wrolstad RE (2004) Total anthocyanins and total phenolics of fresh and processed cherries and their antioxidant properties. *Journal of Food Science* 69(1): 67–72. <https://doi.org/10.1111/j.1365-2621.2004.tb17858.x>

Collins MW, Saag KG and Singh JA (2019) Is there a role for cherries in the management of gout? *Therapeutic Advances in Musculoskeletal Disease* 11: 1–16. <https://doi.org/10.1177/1759720X19847018>

Connolly, DAJ, McHugh MP and Padilla-Zakour OI (2006). Efficacy of a tart cherry juice blend in preventing the symptoms of muscle damage. *British journal of sports medicine*, 40(8), 679-683.

Del Bo C, Porrini M, Fracassetti D et al. (2014) A single serving of blueberry (*V. corymbosum*) modulates peripheral arterial dysfunction induced by acute cigarette smoking in young volunteers: A randomized-controlled trial. *Food and Function* 5(12): 3107-3116. <https://doi.org/10.1039/C4FO00570H>

Desai T, Roberts M and Bottoms L (2021) Effects of short-term continuous Montmorency tart cherry juice supplementation in participants with metabolic syndrome. *European Journal of Nutrition* 60(3): 1587–1603. <https://doi.org/10.1007/s00394-020-02355-5>

Dimitriou L, Hill JA, Jehnali A et al. (2015) Influence of a Montmorency cherry juice blend on indices of exercise-induced stress and upper respiratory tract symptoms following marathon running - A pilot investigation. *Journal of the International Society of Sports Nutrition* 12(1): 1–7. <https://doi.org/10.1186/s12970-015-0085-8>

Dwan K, Li T, Altman DG and Elbourne D (2019) CONSORT 2010 statement: Extension to randomised crossover trials. *British Medical Journal* 366(14378): 1-16. <https://doi.org/10.1136/bmj.l4378>

Franklin SS, Thijs L, Hansen TW et al. (2013) White-coat hypertension: new insights from recent studies. *Hypertension* 62(6): 982-987. <https://doi.org/10.1161/HYPERTENSIONAHA.113.01275>

Haidari F, Mohammad Shahi M et al. (2009) Inhibitory effects of tart cherry (*Prunus cerasus*) juice on xanthine oxidoreductase activity and its hypouricemic and antioxidant effects on rats. *Malaysian Journal of Nutrition* 15(1): 53–64.

Hernández LM, Byrne ML, Taylor MK. (2024) Salivary C-reactive protein exhibits a diurnal pattern and relates to biobehavioral health in military men. *Brain Behaviour and Immunity*. 2024 Nov;122:465-470. <https://doi.org/10.1016/j.bbi.2024.08.040>.

Hillman AR and Uhranowsky K (2021) Acute ingestion of Montmorency tart cherry reduces serum uric acid but has no impact on high sensitivity c-reactive protein or oxidative capacity. *Plant Foods for Human Nutrition* 76: 83–89. <https://doi.org/10.1007/s11130-021-00879-7>

Hobbs DA, George TW and Lovegrove JA (2013) The effects of dietary nitrate on blood pressure and endothelial function: A review of human intervention studies. *Nutrition Research Reviews* 26(2): 210–222. <https://doi.org/10.1017/S0954422413000188>

Howard JP, Shun-Shin MJ, Hartley A et al. (2016) Quantifying the 3 biases that lead to unintentional overestimation of the blood pressure-lowering effect of renal denervation. *Circulation: Cardiovascular Quality and Outcomes* 9(1): 14-22. <https://doi.org/10.1161/CIRCOUTCOMES.115.002533>

Howatson G, McHugh MP, Hill JA et al. (2009) Influence of tart cherry juice on indices of recovery following marathon running. *Scandinavian Journal of Medicine and Science in Sports* 20(6): 843–852. <https://doi.org/10.1111/j.1600-0838.2009.01005.x>

Jacob R, Spinozzi G, Simon V et al. (2003) Consumption of cherries lowers plasma urate in healthy women. *Journal of Nutrition* 133: 1826–1829. <https://doi.org/10.1093/jn/133.6.1826>

Keane KM, George TW, Constantinou CL et al. (2016a) Effects of Montmorency tart cherry (*Prunus Cerasus L.*) consumption on vascular function in males with early hypertension. *American Journal of Clinical Nutrition* 103(6): 1531–1539. <https://doi.org/10.3945/ajcn.115.123869>.

Keane KM, Bell PG, Lodge JK et al. (2016b) Phytochemical uptake following human consumption of Montmorency tart cherry (*L. Prunus cerasus*) and influence of phenolic acids on vascular smooth muscle cells in vitro. *European Journal of Nutrition* 55(4): 1695–1705. <https://doi.org/10.1007/s00394-015-0988-9>

Keane KM, Haskell-Ramsay CF et al. (2016c) Montmorency tart cherries (*Prunus cerasus L.*) modulate vascular function acutely, in the absence of improvement in cognitive performance. *British Journal of Nutrition* 116(11): 1935–1944. <https://doi.org/10.1017/S0007114516004177>

Kelley DS, Rasooly R, Jacob RA et al. (2006) Consumption of Bing sweet cherries lowers circulating concentrations of inflammation markers in healthy men and women. *Journal of Nutrition* 136(4): 981–986. <https://doi.org/10.1093/jn/136.4.981>

Kelley D, Adkins Y and Laugero K (2018) A review of the health benefits of cherries. *Nutrients* 10(3): 368-390. <https://doi.org/10.3390/nu10030368>

Kimble R, Keane KM, Lodge JK, et al. (2021) The Influence of Tart Cherry (*Prunus cerasus*, cv Montmorency) Concentrate Supplementation for 3 Months on

Cardiometabolic Risk Factors in Middle-Aged Adults: A Randomised, Placebo-
Controlled Trial. *Nutrients* 13(5):1417. doi: 10.3390/nu13051417.

Kirakosyan A, Seymour EM, Llanes DEU et al. (2009) Chemical profile and
antioxidant capacities of tart cherry products. *Food Chemistry* 115(1): 20–25.
<https://doi.org/10.1016/j.foodchem.2008.11.042>

Kirakosyan A, Gutierrez E, Ramos Solano B et al. (2018) The inhibitory potential of
Montmorency tart cherry on key enzymes relevant to type 2 diabetes and cardiovascular
disease. *Food Chemistry* 252: 142–146.
<https://doi.org/10.1016/j.foodchem.2018.01.084>

Kuo CF, Grainge MJ, Mallen C et al. (2016) Comorbidities in patients with gout prior
to and following diagnosis: case-control study. *Annals of the Rheumatic Diseases* 75(1):
210–217. <https://doi.org/10.1136/annrheumdis-2014-206410>

Lee J, Durst RW and Wrolstad RE (2005) Determination of total monomeric
anthocyanin pigment content of fruit juices, beverages, natural colorants, and wines by
the pH differential method: Collaborative study. *The Journal of AOAC International*
88(5): 1269–1278. <https://doi.org/10.1093/jaoac/88.5.1269>

Lee SG, Vance TM, Nam TG et al. (2016) Evaluation of pH differential and HPLC
methods expressed as cyanidin-3-glucoside equivalent for measuring the total
anthocyanin contents of berries. *Journal of Food Measurement and Characterization*
10: 562-568. <https://doi.org/10.1007/s11694-016-9337-9>.

Levers K, Dalton R, Galvan E et al. (2016) Effects of powdered Montmorency tart
cherry supplementation on acute endurance exercise performance in aerobically trained
individuals. *Journal of the International Society of Sports Nutrition* 13(22): 1–23.
<https://doi.org/10.1186/s12970-016-0133-z>

Lynn A, Mathew S, Moore CT et al. (2014) Effect of a tart cherry juice supplement on arterial stiffness and inflammation in healthy adults: A randomised controlled trial. *Plant Foods for Human Nutrition* 69(2): 122–127. <https://doi.org/10.1007/s11130-014-0409-x>

Martin KR & Coles 2010

Martin KR, Burrell L and Bopp J (2018) Authentic tart cherry juice reduces markers of inflammation in overweight and obese subjects: a randomized, crossover pilot study. *Food and Function* 9(10): 5290–5300. <https://doi.org/10.1039/c8fo01492b>

Ndrepepa G. (2025) Uric acid and cardiovascular disease—recent evidence on the association and underlying mechanisms. *Journal of Laboratory and Precision Medicine* 10 (8): DOI: 10.21037/jlpm-25-8.

Ou B, Bosak KN, Brickner PR et al. (2012) Processed tart cherry products - Comparative phytochemical content, in vitro antioxidant capacity and in vitro anti-inflammatory activity. *Journal of Food Science* 77(5): 105–112. <https://doi.org/10.1111/j.1750-3841.2012.02681.x>

Pallant J (2010) Statistical techniques to compare groups. In: Pallant J (ed) *SPSS Survival Manual*, 4th edn, McGraw-Hill, pp. 210.

Richter CK, Skulas-Ray AC, Gaugler TL et al. (2017) Incorporating freeze-dried strawberry powder into a high-fat meal does not alter postprandial vascular function or blood markers of cardiovascular disease risk: A randomized controlled trial. *American Journal of Clinical Nutrition* 105(2): 313-322. <https://doi.org/10.3945/ajcn.116.141804>

Rodriguez-Mateos A, Feliciano RP, Boeres A et al. (2016) Cranberry (poly) phenol metabolites correlate with improvements in vascular function: A double-blind,

randomized, controlled, dose-response, crossover study. *Molecular Nutrition and Food Research* 60(10): 2130-2140. <https://doi.org/10.1002/mnfr.201600250>

Rodriguez-Mateos A, Rendeiro C, Bergillos-Meca T et al. (2013) Intake and time dependence of blueberry flavonoid-induced improvements in vascular function: a randomized, controlled, double-blind, crossover intervention study with mechanistic insights into biological activity. *American Journal of Clinical Nutrition* 98(5): 1179-1191. <https://doi.org/10.3945/ajcn.113.066639>

Schlesinger N (2005) Dietary factors and hyperuricaemia. *Current Pharmaceutical Design* 11(32):4 133–4138. <https://doi.org/10.2174/138161205774913273>

Schumacher HR, Pullman-Mooar S, Gupta SR et al. (2013) Randomized double-blind crossover study of the efficacy of a tart cherry juice blend in treatment of osteoarthritis (OA) of the knee. *Osteoarthritis and Cartilage* 21(8): 1035-1041. <https://doi.org/10.1016/j.joca.2013.05.009>

Seeram NP, Momin RA et al. (2001) Cyclooxygenase inhibitory and antioxidant cyanidin glycosides in cherries and berries. *Phytomedicine* 8(5): 362–369. <https://doi.org/10.1078/0944-7113-00053>

Sennels HP, Jørgensen HL et al. (2012) Rhythmic 24-hour variations of frequently used clinical biochemical parameters in healthy young males - The Bispebjerg study of diurnal variations. *Scandinavian Journal of Clinical and Laboratory Investigation* 72(4): 287–295. <https://doi.org/10.3109/00365513.2012.662281>

Shimizu M, Naito R, Sato A, et al. (2023). Diurnal Variations in Serum Uric Acid, Xanthine, and Xanthine Oxidoreductase Activity in Male Patients with Coronary Artery Disease. *Nutrients*, 15(20), 4480. <https://doi.org/10.3390/nu15204480>

Singleton VL and Rossi JA (1965) Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents. *American Journal of Enology and Viticulture* 16(3): 144–158. <http://www.ajevonline.org/content/16/3/144.abstract>

Terkeltaub R, Bushinsky DA and Becker MA (2006) Recent developments in our understanding of the renal basis of hyperuricemia and the development of novel antihyperuricemic therapeutics. *Arthritis Research and Therapy* 8(1): 1–9. <https://doi.org/10.1186/ar1909>

Vendrame S and Klimis-Zacas D (2019) Potential factors influencing the effects of anthocyanins on blood pressure regulation in humans: a review. *Nutrients* 11(6): 1431–1450. doi:10.3390/nu11061431

Virgen Gen JJ, Guzmán-Gerónimo RI, Martínez-Flores K et al. (2020) Cherry extracts attenuate inflammation and oxidative stress triggered by monosodium urate crystals in THP-1 cells. *Journal of Food Biochemistry* 44(10): 1–9. <https://doi.org/10.1111/jfbc.13403>

Wang H, Nair MG, Strasburg GM et al. (1999) Antioxidant and antiinflammatory activities of anthocyanins and their aglycon, cyanidin, from tart cherries. *Journal of Natural Products* 62(2): 294–296. <https://doi.org/10.1021/np980501m>

White SJ, Carran EL, Reynolds AN et al. (2018) The effects of apples and apple juice on acute plasma uric acid concentration: a randomized controlled trial. *American Journal of Clinical Nutrition* 107(2): 165–172. <https://doi.org/10.1093/ajcn/nqx059>

Zhang Y, Neogi T, Chen C et al. (2012) Cherry consumption and decreased risk of recurrent gout attacks. *Arthritis and Rheumatology* 64(12): 4004–4011. <https://doi.org/10.1002/art.34677>