

Editorial: Structural and functional venomics as a powerful approach for drug discovery and development

ABDEL-RAHMAN, Mohamed A., CAO, Zhijian, ABD EL-AZIZ, Tarek Mohamed, STRONG, Peter and SABATIER, Jean-Marc

Available from Sheffield Hallam University Research Archive (SHURA) at:

<https://shura.shu.ac.uk/33625/>

This document is the Published Version [VoR]

Citation:

ABDEL-RAHMAN, Mohamed A., CAO, Zhijian, ABD EL-AZIZ, Tarek Mohamed, STRONG, Peter and SABATIER, Jean-Marc (2024). Editorial: Structural and functional venomics as a powerful approach for drug discovery and development. *Frontiers in Pharmacology*, 15: 1405681. [Article]

Copyright and re-use policy

See <http://shura.shu.ac.uk/information.html>



OPEN ACCESS

EDITED AND REVIEWED BY
Diana Conte Camerino,
University of Bari Aldo Moro, Italy

*CORRESPONDENCE
Mohamed A. Abdel-Rahman,
✉ mohamed_hassanain@science.suez.edu.eg

RECEIVED 23 March 2024

ACCEPTED 28 March 2024

PUBLISHED 10 April 2024

CITATION

Abdel-Rahman MA, Cao Z, Abd El-Aziz TM, Strong PN and Sabatier J-M (2024), Editorial: Structural and functional venomomics as a powerful approach for drug discovery and development.
Front. Pharmacol. 15:1405681.
doi: 10.3389/fphar.2024.1405681

COPYRIGHT

© 2024 Abdel-Rahman, Cao, Abd El-Aziz, Strong and Sabatier. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Structural and functional venomomics as a powerful approach for drug discovery and development

Mohamed A. Abdel-Rahman^{1*}, Zhijian Cao²,
Tarek Mohamed Abd El-Aziz^{3,4}, Peter N. Strong⁵ and
Jean-Marc Sabatier⁶

¹Zoology Department, Faculty of Science, Suez Canal University, Ismailia, Egypt, ²National “111” Center for Cellular Regulation and Molecular Pharmaceutics, Key Laboratory of Fermentation Engineering (Ministry of Education), School of Life and Health Sciences, Hubei University of Technology, Wuhan, China, ³Department of Internal Medicine, Cardiovascular Division, Washington University School of Medicine, St. Louis, MO, United States, ⁴Zoology Department, Faculty of Science, Minia University, El-Minia, Egypt, ⁵Biomolecular Sciences Research Centre, Sheffield Hallam University, Sheffield, United Kingdom, ⁶Aix-Marseille University, CNRS, INP, Marseille, France

KEYWORDS

animal venoms, venom peptide, venom gland transcriptome, toxinome, venomomics, antivenomics, drug discovery

Editorial on the Research Topic

Structural and functional venomomics as a powerful approach for drug discovery and development

The diversity of terrestrial and marine venomous creatures is extremely high with more than 200,000 species spread throughout the Animal Kingdom, including vertebrate animals (reptiles and fishes) and invertebrate animals (marine cone snails, octopi, leeches, myriapods, insects and arachnids). Venomous animals have a specialized tissue or organ called the “venom gland” in which their venoms are produced/stored and introduced into the prey by a specialized delivery apparatus such as a fang, stinger, teeth, or harpoon. Animal venoms are very complicated and often deadly cocktails, which contain unique mixtures of toxins (mainly peptides and proteins) targeting vital systems of the victim or prey. For their high degree of specificity and potency, venom peptides have been used as good templates to develop various venom-derived drugs such as Captopril, Ziconotide, and Exenatide for treating hypertension, chronic pain, and type-2 diabetes, respectively. Also, the scorpion venom peptide chlorotoxin (ClTx) is a useful tumor imaging agent and radiolabelled ClTx has undergone clinical trials for targeted glioma radiotherapy.

Generally, the application of cutting-edge “OMICS” technologies in venom research (venomics to include proteomics, transcriptomics and high-throughput functional assays) is extremely advantageous for several reasons: (1) gaining a better understanding of venom composition and evolution, (2) revealing molecular mechanisms involved in venom toxicity and developing effective antivenoms, (3) understanding biological processes taking place in venom glands and (4) discovery of novel diagnostic and pharmacological agents that can be used in research and

medicine. Accordingly, this Research Topic mainly focuses on the new structural and functional findings (with biological and pharmacological significance) obtained from the venom of terrestrial and marine venomous animals with the aid of venomomics technology. Our Research Topic of Frontiers in Pharmacology includes five original research manuscripts discussing the significance of venomomics approaches in the structural and functional characterization of various toxins (α -Elapitoxin-Nn2a and α -Elapitoxin-Nn3a; apamin; omega-lycotoxin-Gsp2671e; μ -Conotoxin KIIIA and α D-conotoxins VxXXB) derived from snake (*Naja naja*), bee (*Apis mellifera*), spider (*Lycosa praegrandis*), and marine *Conus* (*Conus kinoshitai* and *Conus vexillum*) venoms, respectively.

In the first article described by [Huynh et al.](#), two postsynaptic neurotoxins, α -Elapitoxin-Nn2a and α -Elapitoxin-Nn3a, were isolated and functionally characterized from the venom of Indian Cobra (*Naja naja*) using a combination of proteomic and pharmacologic approaches. Structural analyses revealed that α -Elapitoxin-Nn2a has 62 residues (7,020 Da with four disulfide bridges) and belongs to short-chain neurotoxins. On the other hand, α -Elapitoxin-Nn3a has 71 residues (7,807.5 Da with five disulfide bridges) and belongs to long-chain neurotoxins. Using chick biventer cervicis nerve-muscle preparation, both toxins induced concentration-dependent inhibition of indirect twitches and reduction in contractile responses to exogenous nicotinic agonists (acetylcholine and carbachol). More importantly, the *in vitro* addition of Indian polyvalent antivenom can reverse the neurotoxicity induced by the long-chain neurotoxin Nn3a, but not the short-chain neurotoxin Nn2a.

The second article ([Kuzmenkov et al.](#)) of this Research Topic discusses the contradiction in the structural and electrophysiological data of an important neurotoxin derived from insect venom. In the 1960s, a small peptide (18 amino acid residues, 2026.7 Da) called apamin was purified from the honeybee *A. mellifera* venom. When injected into mice, it was shown to induce muscle spasms, jerks, and convulsions. Apamin selectively acts on small conductance Ca^{2+} -activated potassium channels (KCa^{2+}). However, there is a controversy regarding the publications discussing structural and pharmacological data of apamin. Through extensive structural and electrophysiological profiling of this toxin against several molecular targets (five Ca^{2+} -activated K^{+} Channels (KCa), one inwardly rectifying K^{+} channels (Kir), 15 Voltage-gated K^{+} channels (Kv), 10 Voltage-gated Na^{+} channels (Nav), three Acid-sensing ion channels (ASIC), one transient receptor potential channels (TRP), three glutamate receptors (GluR) and three nicotinic acetylcholine receptors (nAChR)), the contradicting claims have been precisely reconciled in the [Kuzmenkov et al.](#) article. The NMR spectroscopy data showed that apamin forms a β -turn (Asn2–Ala5) and a short α -helix (Ala9–Gln16) and that there are no electrostatic, π -cation, or stacking interactions that might sustain the observed apamin structure. The findings show that apamin selectively inhibits KCa^{2+} channels ($\text{KCa}^{2.1}$, $\text{KCa}^{2.2}$, and $\text{KCa}^{2.3}$) at nanomolar or sub-nanomolar concentrations (IC_{50} values were 4.1 nM, 87.7 pM, and 2.3 nM, respectively), with negligible impact on other molecular

targets. Accordingly, the structural and functional data presented in this work support the utilization of apamin as a KCa^{2+} -selective pharmacological tool and a good model for developing new drugs.

The third article in this Research Topic examined how omega-lycotoxin-Gsp2671e (as a P/Q-type VGCC modulator) from *L. praegrandis* spider venom improves memory in glutamate-induced excitotoxicity rats ([Keimasi et al.](#)). Various venomomics and behavioral analyses were used in this investigation including venom extraction, gel filtration chromatography, capillary electrophoresis, mass spectrometry, acute cytotoxicity (LD_{50}), Morris Water Maze task, and Novel Object Recognition test (to assess long and short-term memory). In hyper-stimulated NMDA (N-Methyl-D-aspartate receptors) rats, OLG1e administration resulted in increased expression of synaptophysin (SYN), synaptosomal-associated protein-25 kDa (SNAP-25), and synaptotagmin 1 (SYT1), as well as the restoration of upper levels of field excitatory postsynaptic potentials (fEPSP) in comparison to the non-treated NMDA group. The results of this study can be used as a foundation for future analyses of how OLG1e affects cognitive functions like pain sensitivity, associative learning, sensorimotor and locomotor ability.

The fourth ([Kimball et al.](#)) and fifth ([Ho et al.](#)) articles of this Research Topic discuss the electrophysiological activity of two important conotoxins (μ -conotoxin KIIIA and α D-VxXXB, respectively) derived from the venoms of marine *Conus*. Kimball and colleagues used a combination of *in silico* (Rosetta Computational Modeling and Rosetta Dock) and experimental approaches to investigate the molecular interaction between μ -conotoxin KIIIA (KIIIA) and human Nav1.7 channel (hNav1.7). The obtained results through this integrative approach can be potentially useful for developing specific peptide-based therapeutics targeting Nav1.7 which plays a crucial role in pain signaling. Moving to the second conotoxin α D-VxXXB ([Ho et al.](#)) which is derived from the venom of *C. vexillum*. Through a non-competitive (allosteric) mechanism, α D-conotoxins (11 kDa homodimers) potentially block nicotinic acetylcholine receptors (nAChRs). Ho and colleagues described the allosteric binding mechanism between the extracellular ligand-binding domain of nAChRs and the granulin-like C-terminal (CTD) of VxXXB. According to mutational and docking studies, the CTDs of α D-VxXXB bind cooperatively at two unique allosteric sites on nAChRs. Thus, the information that Ho and colleagues have acquired opens up a new direction in the development of novel subtype-specific allosteric nAChR antagonists.

Finally, the research findings presented through this Research Topic clearly demonstrate the usefulness of venomomics approaches in venom separation, identification, and characterization of various venomous animals. Different venom peptides have been characterized and can be considered good templates for developing effective candidate drugs targeting various diseases. We are confident that this Research Topic of articles will motivate several toxinologists and clinicians globally to do further research in the field of natural toxins derived from venomous animals using the cutting-edge technologies of venomomics.

Author contributions

MA-R: Conceptualization, Methodology, Supervision, Writing–original draft, Writing–review and editing. ZC: Conceptualization, Validation, Writing–review and editing. TAE-A: Conceptualization, Validation, Writing–review and editing. PS: Conceptualization, Validation, Writing–review and editing. J-MS: Conceptualization, Validation, Writing–review and editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. MA-R receives funding to support his venomomics research from the Science, Technology & Innovation Funding Authority (STDF, Egypt; ID 45890) and the Academy of Scientific Research and Technology (ASRT, Egypt).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.