

Of mitochondrion and COVID-19.

ALFAROUK, Khalid Omer <<http://orcid.org/0000-0001-8656-6117>>, ALHOUFIE, Sari TS <<http://orcid.org/0000-0003-0253-7832>>, HIFNY, Abdelhameed, SCHWARTZ, Laurent, ALQAHTANI, Ali S, AHMED, Samrein <<http://orcid.org/0000-0001-9773-645X>>, ALQAHTANI, Ali M, ALQAHTANI, Saad S, MUDDATHIR, Abdel Khalig, ALI, Heyam, BASHIR, Adil HH, IBRAHIM, Muntaser E, GRECO, Maria Raffaella, CARDONE, Rosa A <<http://orcid.org/0000-0002-2011-9135>>, HARGUINDEY, Salvador and RESHKIN, Stephan Joel

Available from Sheffield Hallam University Research Archive (SHURA) at:

<http://shura.shu.ac.uk/33569/>

This document is the author deposited version. You are advised to consult the publisher's version if you wish to cite from it.

Published version

ALFAROUK, Khalid Omer, ALHOUFIE, Sari TS, HIFNY, Abdelhameed, SCHWARTZ, Laurent, ALQAHTANI, Ali S, AHMED, Samrein, ALQAHTANI, Ali M, ALQAHTANI, Saad S, MUDDATHIR, Abdel Khalig, ALI, Heyam, BASHIR, Adil HH, IBRAHIM, Muntaser E, GRECO, Maria Raffaella, CARDONE, Rosa A, HARGUINDEY, Salvador and RESHKIN, Stephan Joel (2021). Of mitochondrion and COVID-19. *Journal of enzyme inhibition and medicinal chemistry*, 36 (1), 1258-1267.

Copyright and re-use policy

See <http://shura.shu.ac.uk/information.html>



Of mitochondrion and COVID-19

Khalid Omer Alfarouk, Sari T. S. Alhoufie, Abdelhameed Hifny, Laurent Schwartz, Ali S. Alqahtani, Samrein B. M. Ahmed, Ali M. Alqahtani, Saad S. Alqahtani, Abdel Khalig Muddathir, Heyam Ali, Adil H. H. Bashir, Muntaser E. Ibrahim, Maria Raffaella Greco, Rosa A. Cardone, Salvador Harguindey & Stephan Joel Reshkin

To cite this article: Khalid Omer Alfarouk, Sari T. S. Alhoufie, Abdelhameed Hifny, Laurent Schwartz, Ali S. Alqahtani, Samrein B. M. Ahmed, Ali M. Alqahtani, Saad S. Alqahtani, Abdel Khalig Muddathir, Heyam Ali, Adil H. H. Bashir, Muntaser E. Ibrahim, Maria Raffaella Greco, Rosa A. Cardone, Salvador Harguindey & Stephan Joel Reshkin (2021) Of mitochondrion and COVID-19, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 36:1, 1258-1266, DOI: [10.1080/14756366.2021.1937144](https://doi.org/10.1080/14756366.2021.1937144)

To link to this article: <https://doi.org/10.1080/14756366.2021.1937144>



© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Published online: 09 Jun 2021.



[Submit your article to this journal](#)



Article views: 5510



[View related articles](#)



[View Crossmark data](#)



Citing articles: 10 [View citing articles](#)

Of mitochondrion and COVID-19

Khalid Omer Alfarouk^{a,b,c}, Sari T. S. Alhoufie^d, Abdelhameed Hifny^e, Laurent Schwartz^f, Ali S. Alqahtani^g, Samrein B. M. Ahmed^h, Ali M. Alqahtaniⁱ, Saad S. Alqahtani^j, Abdel Khalig Muddathir^k, Heyam Ali^k, Adil H. H. Bashir^l, Muntaser E. Ibrahim^l, Maria Raffaella Greco^m, Rosa A. Cardone^m, Salvador Harguindeyⁿ and Stephan Joel Reshkin^m

^aResearch Center, Zamzam University College, Khartoum, Sudan; ^bDepartment of Evolutionary Pharmacology and Tumor Metabolism, Hala Alfarouk Cancer Center, Khartoum, Sudan; ^cAl-Ghad International College for Applied Medical Sciences, Al-Madinah Al-Munwarah, Saudi Arabia; ^dMedical Laboratories Technology Department, College of Applied Medical Sciences, Taibah University, Al-Madinah Al-Munwarah, Saudi Arabia; ^eFaculty of Medicine, Al-Azhar University, Cairo, Egypt; ^fAssistance Publique, des Hôpitaux de Paris, Paris, France; ^gCollege of Applied Medical Sciences, Najran University, Najran, Saudi Arabia; ^hCollege of Medicine, University of Sharjah, Sharjah, UAE; ⁱDepartment of Pharmacology, College of Pharmacy, King Khalid University, Abha, Saudi Arabia; ^jPharmacy Practice Research Unit, Clinical Pharmacy Department, College of Pharmacy, Jazan University, Jazan, Saudi Arabia; ^kFaculty of Pharmacy, University of Khartoum, Khartoum, Sudan; ^lInstitute of Endemic Diseases, University of Khartoum, Khartoum, Sudan; ^mDepartment of Biosciences, Biotechnologies, and Biopharmaceutics, University of Bari, Bari, Italy; ⁿInstitute for Clinical Biology and Metabolism, Vitoria, Spain

ABSTRACT

COVID-19, a pandemic disease caused by a viral infection, is associated with a high mortality rate. Most of the signs and symptoms, e.g. cytokine storm, electrolytes imbalances, thromboembolism, etc., are related to mitochondrial dysfunction. Therefore, targeting mitochondrion will represent a more rational treatment of COVID-19. The current work outlines how COVID-19's signs and symptoms are related to the mitochondrion. Proper understanding of the underlying causes might enhance the opportunity to treat COVID-19.

ARTICLE HISTORY

Received 6 March 2021
Revised 20 April 2021
Accepted 20 May 2021

KEYWORDS

COVID-19; mitochondrion; inflammation; cytokine storm; treatment

Introduction

COVID-19 is a new emerging pulmonary infection caused by SARS-CoV-2. It is characterised by flu-like symptoms often followed by acute pulmonary inflammation. Multiple viruses are known to cause both inflammation and mitochondrial dysregulation (metabolic shifts). The influenza virus H1N1 targets the mitochondria of type II cells¹. Multiple other inflammatory viruses are known to induce metabolic changes, such as the cytomegalovirus (CMV)², the Epstein-Barr virus (EBV)³, or the hepatitis virus (HCV)⁴. These viruses interfere with cellular metabolism, increase glucose uptake, and decrease the mitochondrial energy yield resulting in intense glycolysis. In Caco-2 cells, infection with SARS-CoV-2 has been found to up-regulate carbon metabolism and decrease oxidative phosphorylation. I removed it because it is out of context and there is no reference- also no reference for the Caco-2 cells.

The mitochondrion is a doubled-membrane organelle, represents the backbone of the eukaryote cell metabolism^{5,6}. Mitochondrion is the cells' metabolic generator and plays a significant role in determining cellular proliferation⁷, cellular death pathways⁸ and also plays a crucial role in maintaining the redox state of the cell⁹.

Many viral diseases disturb the mitochondrial physiology^{10–12}, e.g. Epstein-Barr virus (EBV) affects mitochondrial fission¹³, herpes simplex virus type 1 (HSV-1) and pseudorabies virus (PRV) affect calcium homeostasis¹⁴, and many viruses, e.g. influenza viruses,

Hepatitis B virus, support and/or encode proapoptotic proteins that lead to programmed cell death^{15–17}.

Since the occurrence of unidentified pneumonia patients in Wuhan hospitals in China in late 2019 and the labelling of the disease by the World Health Organisation (WHO) as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), the disease became a pandemic in less than three months, and as of the beginning of December 2020 the total confirmed cases of COVID-19 reached 65,257,767 worldwide according to a WHO update^{18–20}.

Despite the increased global incidence records of the COVID-19 cases, most of the infected patients showed either mild infection with no fever or signs of pneumonia or moderate infection with clinical manifestations like cough, sore throat, fever $\geq 38^\circ\text{C}$, fatigue, and shortness of breath²¹.

Severe infection with increased mortality rate occurs with pneumonia and respiratory failure. At the same time, other complications might present, such as acute respiratory distress syndrome (ARDS), microvascular thrombosis, coagulopathy, liver injury, acute kidney injury, acute cardiac failure and shock^{22–27}. Factors affecting the infection's severity are not fully understood; however, factors such as the state of the immune system, viral load, and underlying comorbid diseases might play a role in the severity of the infection^{28–30}.

In the current work, we present COVID-19 as a mitochondriopathy and demonstrate that many of the hallmarks of COVID-19 are driven by mitochondrial injury.

The role of mitochondria and cytokine storm

Hyperinflammation – e.g. cytokine storm – is a hallmark of COVID-19³¹. Such hyper-inflammation occurs due to a massive increase in Reactive Oxygen Species (ROS)^{32,33}. Increased ROS results in the release of tumour necrosis factor (TNF)- α and interleukin-1 β (IL-1 β)^{34,35}. The mitochondrion is a significant source of ROS in mammalian cells³⁶. Therefore, the mitochondrion lies within the cytokine storm's core³⁷.

The inflammasome is a cytosolic complex composed of multiple proteins of innate immunity to promote and activate the proinflammatory mediators such as IL-1 β , IL-18^{38–41}. One protein component is an intracellular pathogen sensor called nucleotide-binding oligomerization domain-like receptors, or NOD-like receptors (NLRs)⁴². NLRP3 is one NOD-like receptor (NLRs) family member that represents the backbone of the inflammasome. The role of NLRP3 in inflammation and the cytokine storm is crucial and complex. As a consequence of its activation, the cell reprograms its metabolic machinery into increased glycolysis with a subsequent reduction of the Krebs' cycle⁴³, i.e. induces mitochondrial atrophy. ROS also activates the NLRP3 where it is associated with mitochondrial cardiolipin⁴⁰ and might be correlated with mitochondrial ageing (which stimulates the inflammasome)⁴⁴.

SARS-COV-2 infection attacks the mitochondrion, especially the phosphorylation (OxPHOS) pathway, e.g. Complex-I⁴⁵, which results in abnormal ROS production supporting cellular diseases and ageing. SARS-CoV-2 might directly activate the NLRP3 inflammasome, with consequent flaring-up of the inflammation cascade⁴⁰. Hence, SARS-COV-2 alters mitochondrial physiology^{46,47}.

COVID-19 disrupts the possible mitochondrial role in iron homeostasis

Iron is an essential nutrient and its levels differ from one tissue to another and also depend on the tissues pathological state⁴⁸. Cellular iron homeostasis is a complexed process⁴⁹, but generally, it could be described as: the entrance of iron to the cell through: (i) endocytosis of transferrin receptor 1 (TfR1), or (ii) ferrous iron (Fe⁺²) transporters e.g. divalent metal transporter 1 (DMT1)⁵⁰ and Zinc transporters 8, 14 (ZIP8, ZIP14)^{51,52} with the assistance of the iron reductase enzyme Metalloreductase STEAP2⁵³, Duodenal cytochrome B (Dcytb)⁵², and Stromal cell-derived receptor 2 (SDR-2)⁵⁴. After being taken-up, the iron is stored in ferritin^{55–57} for different biochemical functions including the formation of ROS^{58,59} and managing transcription through regulating the iron-responsive element-binding proteins (IRP1, IRP2)^{60,61}. After that, iron export from the cell occurs via ferroportin-1 (also termed as solute carrier family 40 member 1 (SLC40A1) or iron-regulated transporter 1 (IREG1))⁶².

The role of mitochondria in iron homeostasis is one of the most challenging of recently addressed issues. Generally, ferritin is an intracellular protein that can act as an iron-buffering agent to re-equilibrate iron deficiency or iron overload⁶³. Ferritin is stored in the mitochondrion and imported from the cytoplasm via mitochondria ferrin carriers^{64,65}.

Disruption of mitoferrin leads to hyperferritinemia, accompanied by hyper-inflammation, an additional hallmark of COVID-19 severity^{64,66,67}. Severe iron overload leads to mitochondrial DNA damage that exacerbates the cellular oxidative stress⁶⁸.

For this reason, the iron-chelating agent, Deferoxamine, has been introduced in the management of COVID-19^{69,70}.

Lactate dehydrogenase in COVID-19

The lactate dehydrogenase (LDH) is an enzyme that catalyses a reversible biochemical reaction that converts pyruvate into lactate. After glucose entry, the hydrogen ions (proton, H⁺) level is rising, alters the cell's optimum pH to process its chemical pathways. After completing the Krebs' cycle, the cell yields in CO₂, energy in ATP, and hydrogen ions. The oxygen reacts with H⁺ to produce water. Therefore, oxygen in cellular respiration acts as a detoxifying agent (acting as a buffer)⁷¹. During transient hypoxia, some tissues, e.g. heart, brain, kidney, are prone to damage.

In contrast, other tissues are slightly adaptable by expressing the lactate dehydrogenase enzyme to shift the cellular metabolism to prevent the Krebs' cycle. Therefore, the glucose utilisation after its entry ends up by forming lactic acid and furthering extracellular acidity via Monocarboxylate Transporters (MCTs)^{72–74}. So, metabolic shifting to end in lactic acid will decrease the possible intracellular acidity and promote the extracellular acidity that exacerbates the cytokine storm as lactate is a signalling molecule that supports inflammation^{75,76}.

The conversion of pyruvate to lactate is associated with the conversion of NADH to NAD⁺. Increasing of NAD⁺ level inhibits not only mitochondrial metabolism but also supports the inflammation process^{77,78}.

LDH is correlated with COVID-19 and its severity⁷⁹ because the lactate synthesis is increased. The level of blood lactate is a prognostic factor for the intensity of the lung's inflammation and decreased survival⁸⁰.

Dysregulation of calcium homeostasis during COVID-19 affects mitochondrial biology

Calcium is a vital electrolyte that plays many critical roles in cellular physiology⁸¹. Calcium governs intracellular mitochondrial motility (mitochondrial dynamics)^{82,83}, manages mitophagy^{84–86}, controls ATP production⁸⁷, and impacts the role of the mitochondrion in the redox statue of the cell⁸⁸.

A reduced level of calcium is well-documented in covid-19 infection, and it is thought to have a role in its poor prognosis⁸⁹. Therefore, hypocalcaemia has a detrimental effect on the mitochondrion, promotes ROS formation, and activates the inflammatory cascade.

The role of the mitochondrion on coagulability

D-dimer

While the term D-dimer reflects the dimerisation process (two sub-units), it also seems to be an erroneous name suggested by one of the researchers that discovered it^{90,91}. All in all, D-dimer is fibrin fragments that are crosslinked with polypeptide bonds due to the degradation of fibrinogen via plasmin^{92,93}. Higher levels of D-dimer in the blood represent a severe sign of thromboembolism^{94–96} and recently has become an indicator of how COVID-19 patients develop thromboembolism and the disease severity^{97–99} since D-dimer level is markedly increased among critical patients and is a significant risk factor for mortality¹⁰⁰.

Oxidative stress is associated with thromboembolism¹⁰¹, in that ROS activates urokinase plasminogen activator (UPA)¹⁰², subsequently producing plasmin that hydrolyses fibrinogen into D-

dimer. The increased Plasmin, in turn, increases ROS¹⁰³, which produces an out-of-control positive feedback between ROS and plasmin. Furthermore, D-dimer expression also might increase the level of urokinase-type plasminogen activator (plasmin activator), and so it also enters a vicious cycle producing thromboembolism.

There is an inverse relationship between functional mitochondrial and urokinase plasminogen, such that upregulation of the UPA is an indicator of reduced mitochondrial function while, in contrast, downregulation of UPA restores mitochondrial function (e.g. activation of programmed cell death)¹⁰³.

Troponins

These are a group of proteins found in the heart and skeletal muscle that mediate calcium-dependent muscle contraction^{104,105}. An increased level of troponins in the blood is an indicator of necrosis rather than programmed cell death, i.e. mitochondrial injury or dysfunctionality due to hypoxia^{106–112}.

COVID-19 is associated with higher troponin levels¹¹³, which might correlate with mortality¹¹⁴. Indeed, higher troponin levels were confined to cardiac disorder and other diseases, such as sepsis or renal disease¹¹⁵, both of which were correlated with COVID-19^{112,116,117}. Also, during cardiac and muscle injury, troponin levels

are increased significantly in severe disease patients, leading to progression towards multiple organ failure (MOF) and death.

Targeting the mitochondrion to treat COVID-19

In 1956, Otto Warburg suggested that cancer occurs due to mitochondrial injury and, in this respect, it seems that COVID-19 could be looked at as an extrapolation of cancer¹¹⁸. At least it could be analysed through Warburg's lens and could stimulate the debate of whether mitochondriopathy is a direct cause of COVID-19 via SARS-COV-2 infection or just a symptom of COVID-19 in which, at least, mitochondrial injury might represent an early step of the SARS-COV-2 disease cascade. In this regard, the administration of pharmacological and non-pharmacological modulators of mitochondrial function¹¹⁹ could enhance patient recovery and improve patients' quality of life and might boost the vaccine's efficacy in the aged population (mitochondrial is a hub of ageing). An example of those agents includes:

1. NHE1 inhibitors:
 - In 2000, Reshkin et al. observed that the over-expression of NHE1 is the first event of carcinogenesis followed by alkaline increases in intracellular pH (alkaline pH)^{120,121};



Figure 1. How does Amiloride re-equilibrate the cytokine storm via boosting the anti-inflammatory cytokines and suppressing the proinflammatory cytokines.

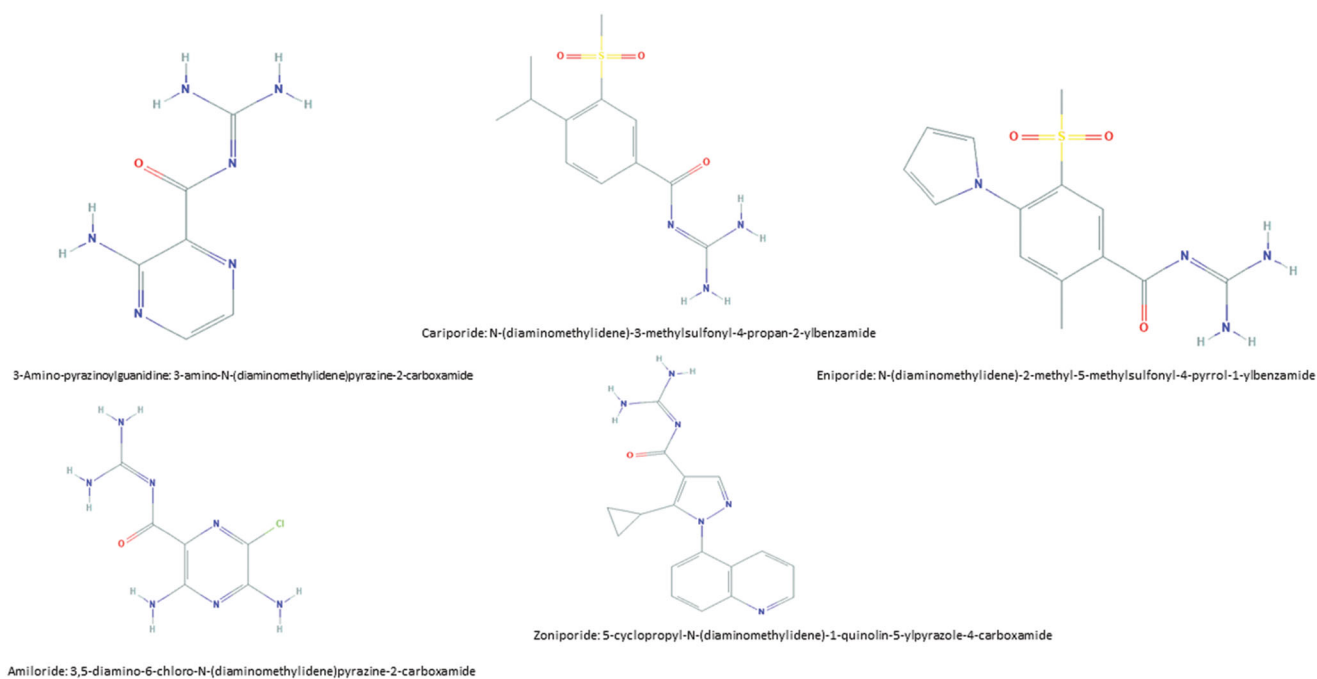


Figure 2. Different chemical formula of some of NHE1 inhibitors.

and alkaline pH_i results in mitochondrial atrophy. Therefore, NHE1 inhibition, and specifically mitochondrial NHE1, will boost the mitochondrial functionality¹²² and so decrease the effect of SARS-CoV-2.

- Amiloride is a potassium-sparing diuretic, and it is a well-known NHE-1 inhibitor. Amiloride perturbs SARS-CoV-2 biology¹²³, and early reports showed that Amiloride inhibited coronavirus replication¹²³
 - Amiloride also has potential as an anti-cytokine storm agent¹²⁴. One of the possible mechanisms of action that explains how Amiloride antagonises the cytokine storm via contrasting the effect of proinflammatory mediators (e.g. the NF-κB transcription factor), by boosting the expression of anti-inflammatory mediators such as Interleukin-10 (IL-10), and nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, alpha (IκBα)¹²⁴ (see Figure 1).
 - Significantly, Amiloride also suppresses the urokinase plasminogen activator (UPA), which might have a promising role in preventing thromboembolism^{125,126} and also prevents heart ischaemia¹²⁷ other NHE1 inhibitors include Cariporide, Eniporide, etc. (see Figure 2).
2. Fermented wheat germ extract:
 - a. Fermented wheat germ extract (FWGE) is a dietary supplement used to treat cancer and to slow ageing. The mode of action of FWGE is a mitochondrial restoration agent as it modulates the activity of the pyruvate dehydrogenase (PDH) complex to support the production of ATP from mitochondria¹²⁸. Also, FWGE inhibits LDH and reduces the NAD⁺ levels¹²⁸. Moreover, it shows promising action as an anti-cytokine storm drug¹²⁹⁻¹³¹.
 3. α-lipoic acid:
 - a. The history of α-lipoic dates to the 1950s (Figure 3) when German industry developed this drug. The first use of α-lipoic acid was for peripheral neuropathy due to diabetes¹³².
 - b. A preliminary Chinese study suggests the efficacy of α-Lipoic acid in the treatment of COVID-19¹³³, where α-lipoic acid might act in the same way as FWGE; combined with hydroxycitrate, it synergizes the effect as an acting buffer to correct pH_i to restore mitochondrial function^{134,135}.
 4. Methylene Blue
 - a. Methylene Blue is the oldest of synthetic drugs (Figure 4), even before aspirin. Heinrich Carro manufactured it in 1876 for the German firm BASF. Methylene blue is a simple molecule. The fusion of two benzene rings with one nitrogen and one sulphur atom leads to a tricyclic aromatic compound which has a complex pharmacology and multiple clinical indications. Its mechanism of action involves a stabilising effect on mitochondria. Also, Methylene blue inhibits the replication of SARS-CoV-2¹³⁶ and we reported a cohort of patients treated for cancer by Methylene Blue in cases without SARS-CoV-2¹³⁷.
 5. 2-deoxy-d-glucose (2DG)
 - a. The German scientist Otto Warburg discovered the Warburg effect in the 1920s¹³⁸. Warburg stated that cancer cells display increased glycolysis and lactic acid secretion and, opposite to normal cells, the presence of oxygen does not inhibit this fermentation. The advent of Positron Emission Tomography (PET) scan combined with radio-labelled fluorodeoxyglucose has revived interest in the Warburg effect as there is an increased uptake

of labelled glucose in the primary tumour and its distant metastases. The Warburg effect explains some of the cancer's hallmarks^{118,135} shift to aerobic glycolysis that has been reported to stimulate cell growth, evade tumour suppression, and resist cell death¹³⁹. Increased pressure resulting from unrelenting proliferation in the affected organ's limited space results in cells' extrusion in the vasculature and distant metastases. The release of lactic acid in the extracellular space is a consequence of the Warburg effect. Lactic acid promotes angiogenesis and immune cell modulation¹⁴⁰.

- b. Infection with SARS-CoV-2 in Caco-2 cells has been found to up-regulate glycolytic carbon metabolism and decrease oxidative phosphorylation. In line with this, treatment with the glycolysis inhibitor 2-deoxy-d-glucose (2DG) prevents replication of SARS-CoV-2 in these cells¹⁴¹ (Figure 5).
- c. The Warburg hypothesis was based on mitochondrial injury, but the debate is whether it is a cause of malignant transformation or just a consequence. Irrespective of which is correct, mitochondrial damage supports evo-



Figure 3. Chemical Structure of lipoic acid: 5-[(3R)-dithiolan-3-yl] pentanoic acid.



Figure 4. Chemical structure of methylene blue: [7-(dimethylamino) phenothiazin-3-ylidene]-dimethylazanum;chloride.

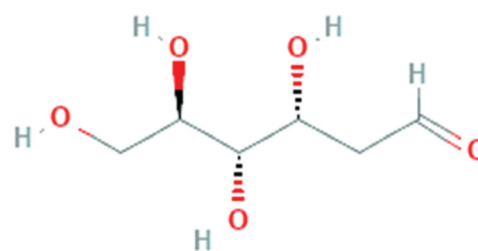


Figure 5. Chemical Structure of 2DG: (3R,4S,5R)-3,4,5,6-tetrahydroxyhexanal.

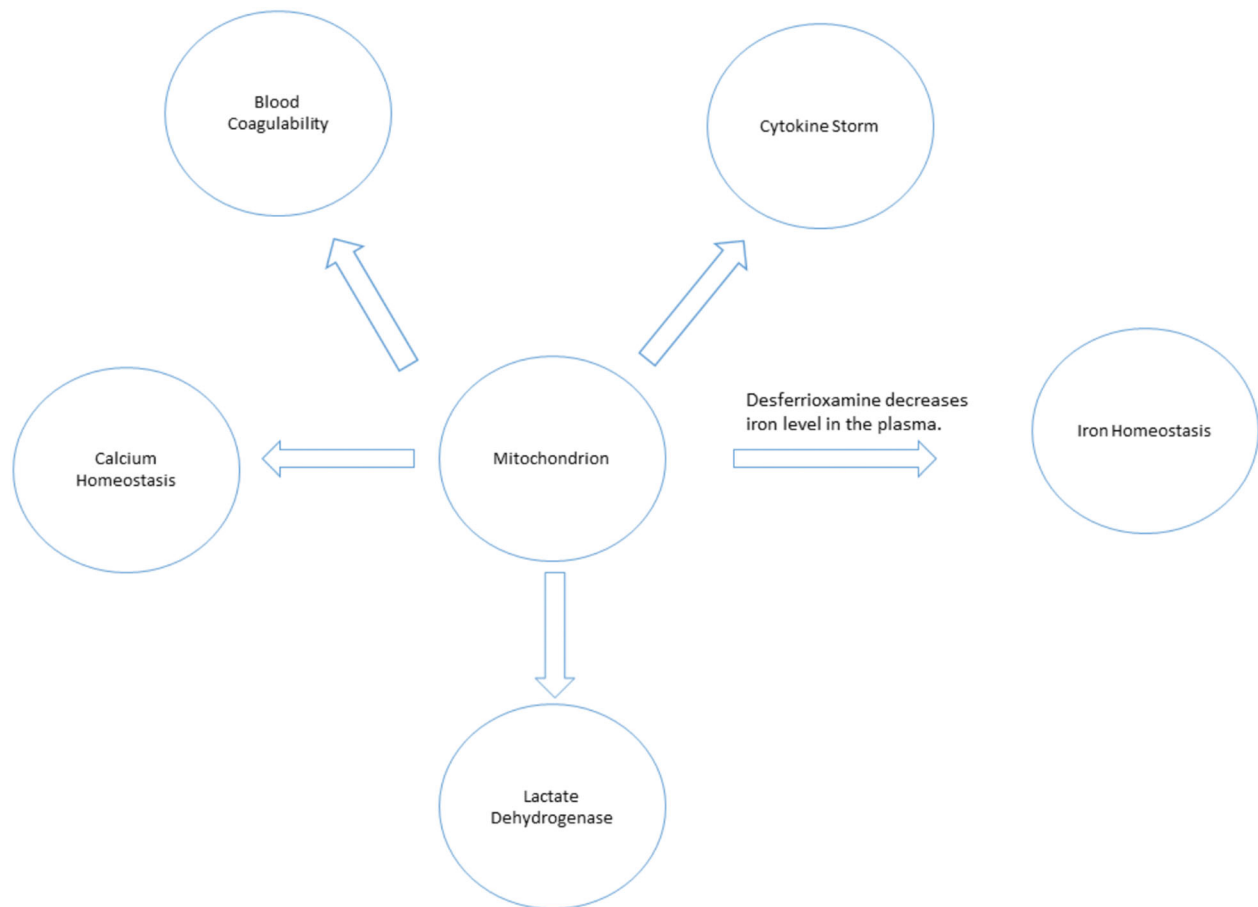


Figure 6. The mitochondrion lies within the core of COVID-19 cardinals.

lutionary tumour trajectory¹⁴². Parallel to this context, COVID-19 is associated with mitochondrial injury and such injury supports SARS-COV-2 pathogenicity and confers its evolutionary advantage. However, a significant concern is whether COVID-19 patients will develop cancer in the future due to such mitochondrial injury?

Recommendations and concluding remarks

COVID-19 has become a pandemic disease. The biology of the disease is exceptionally intricate, including many overlapping pathways. However, while the mitochondrion lies at the core of these pathways, its importance demands immediate attention and further investigation. A proper understanding of mitochondrial biology in COVID-19 pathogenesis will significantly enhance the strategy of fighting SARS-COV-2 (Figure 6). This paper has discussed and suggests a couple of pharmacological modulators that might represent potentially promising anti-COVID-19 treatments to block its progression and alleviate its aggressiveness.

Author contributions

KOA contributed to the conceptualisation, data curation, formal analysis, investigation, resources, software, writing (original draft). SJR contributed to the supervision, conceptualisation, data curation, formal analysis, investigation, resources, software, writing (review and editing). STA, AH, and LS contributed to the

conceptualisation, data curation, resources, writing (original draft). ASA, SBMA, AMA, and SSA contributed to methodology, resources, software. AKM, HA, AHHB, and MI contributed to the investigation, methodology, visualisation. SH, MR, and RAC contributed to investigation, methodology, and resources. SH also contributed to review and correct the final text.

Disclosure statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Davis I, Doolittle L, Guttridge D, et al. H1N1 influenza A virus infection of mice induces the warburg effect in ATII cells. *Am J Respir Crit Care Med* 2020;201:A3842.
2. Yu Y, Clippinger AJ, Alwine JC. Viral effects on metabolism: changes in glucose and glutamine utilization during human cytomegalovirus infection. *Trends Microbiol* 2011; 19:360–7.
3. Darekar S, Georgiou K, Yurchenko M, et al. Epstein-barr virus immortalization of human B-cells leads to stabilization of hypoxia-induced factor 1 alpha, congruent with the Warburg effect. *PLoS One* 2012;7:e42072.

4. Ripoli M, D'Aprile A, Quarato G, et al. Hepatitis C virus-linked mitochondrial dysfunction promotes hypoxia-inducible factor 1 α -mediated glycolytic adaptation. *J Virol* 2010; 84:647–60.
5. Gabaldón T, Huynen MA. From endosymbiont to host-controlled organelle: the hijacking of mitochondrial protein synthesis and metabolism. Subramaniam S, ed. *PLoS Comput Biol* 2007;3:e219.
6. Pittis AA, Gabaldón T. Late acquisition of mitochondria by a host with chimaeric prokaryotic ancestry. *Nature* 2016; 531:101–4.
7. Yan XJ, Yu X, Wang XP, et al. Mitochondria play an important role in the cell proliferation suppressing activity of berberine. *Sci Rep* 2017;7:41712.
8. Vakifahmetoglu-Norberg H, Ouchida AT, Norberg E. The role of mitochondria in metabolism and cell death. *Biochem Biophys Res Commun* 2017;482:426–31.
9. Handy DE, Loscalzo J. Redox regulation of mitochondrial function. *Antioxid Redox Signal* 2012;16:1323–67.
10. Reshi L, Wang H-V, Hong J-R. Modulation of mitochondria during viral infections. In: Taskin E, Guven C, eds. *Mitochondrial diseases*. London: InTech; 2018.
11. Khan M, Syed GH, Kim SJ, et al. Mitochondrial dynamics and viral infections: a close nexus. *Biochim Biophys Acta* 2015;1853:2822–33.
12. Tiku V, Tan MW, Dikic I. Mitochondrial functions in infection and immunity. *Trends Cell Biol* 2020;30:263–75.
13. Young LS, Rickinson AB. Epstein-Barr virus: 40 years on. *Nat Rev Cancer* 2004;4:757–68.
14. Kramer T, Enquist LW. Alpha herpesvirus infection disrupts mitochondrial transport in neurons. *Cell Host Microbe* 2012;11:504–14.
15. Takada S, Shirakata Y, Kaneniwa N, et al. Association of hepatitis B virus X protein with mitochondria causes mitochondrial aggregation at the nuclear periphery, leading to cell death. *Oncogene* 1999;18:6965–73.
16. Hennes T, Peterhans E, Stocker R. Alterations in antioxidant defences in lung and liver of mice infected with influenza A virus. *J General Virol* 1992;73:39–46.
17. Henkler F, Hoare J, Waseem N, et al. Intracellular localization of the hepatitis B virus HBx protein. *J General Virol* 2001;82:871–82.
18. Heinze G, Schemper M. A solution to the problem of separation in logistic regression. *Stat Med* 2002;21:2409–19.
19. Chen H, Guo J, Wang C, et al. Clinical characteristics and intrauterine vertical transmission potential of COVID-19 infection in nine pregnant women: a retrospective review of medical records. *Lancet* 2020;395:809–15.
20. Anon. WHO Coronavirus Disease (COVID-19) Dashboard | WHO Coronavirus Disease (COVID-19) Dashboard. <https://covid19.who.int/> [accessed 5 Dec 2020].
21. Guan W, Ni Z, Hu Y, et al. Clinical characteristics of coronavirus disease 2019 in China. *New Engl J Med* 2020;382:1708–20.
22. Sánchez-Recalde Á, Solano-López J, Miguelena-Hycka J, et al. COVID-19 and cardiogenic shock. Different cardiovascular presentations with high mortality. *Revista Española de Cardiología* 2020;73:669–72.
23. Du R-H, Liang L-R, Yang C-Q, et al. Predictors of mortality for patients with COVID-19 pneumonia caused by SARS-CoV-2: a prospective cohort study. *Eur Respir J* 2020;55:2000524.
24. Yang HJ, Zhang YM, Yang M, et al. Predictors of mortality for patients with COVID-19 pneumonia caused by SARS-CoV-2. *Eur Respir J* 2020;56:2002439.
25. Ciceri F, Ruggeri A, Lembo R, et al. Decreased in-hospital mortality in patients with COVID-19 pneumonia. *Pathogens Global Health* 2020;114:281–2.
26. Paek JH, Kim Y, Park WY, et al. Severe acute kidney injury in COVID-19 patients is associated with in-hospital mortality. Hirst JA, ed. *PLoS One* 2020;15:e0243528.
27. Malas MB, Naazie IN, Elsayed N, et al. Thromboembolism risk of COVID-19 is high and associated with a higher risk of mortality: a systematic review and meta-analysis. *EClinicalMedicine* 2020;29-30:100639.
28. Guo T, Fan Y, Chen M, et al. Cardiovascular implications of fatal outcomes of patients with coronavirus disease 2019 (COVID-19). *JAMA Cardiol* 2020;5:811–8.
29. Bhatraju PK, Ghassemieh BJ, Nichols M, et al. Covid-19 in Critically ill patients in the seattle region – case series. *New Engl J Med* 2020;382:2012–22.
30. Zheng S, Fan J, Yu F, et al. Viral load dynamics and disease severity in patients infected with SARS-CoV-2 in Zhejiang province, China, January. Retrospective cohort study. *BMJ* 2020;369:m1443.
31. Coperchini F, Chiovato L, Croce L, et al. The cytokine storm in COVID-19: an overview of the involvement of the chemokine/chemokine-receptor system. *Cytokine Growth Factor Rev* 2020;53:25–32.
32. Mittal M, Siddiqui MR, Tran K, et al. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal* 2014;20:1126–67.
33. Naha PC, Davoren M, Lyng FM, et al. Reactive oxygen species (ROS) induced cytokine production and cytotoxicity of PAMAM dendrimers in J774A.1 cells. *Toxicol Appl Pharmacol* 2010;246:91–9.
34. Yang D, Elner SG, Bian ZM, et al. Pro-inflammatory cytokines increase reactive oxygen species through mitochondria and NADPH oxidase in cultured RPE cells. *Exp Eye Res* 2007;85:462–72.
35. Kamata H, Honda SI, Maeda S, et al. Reactive oxygen species promote TNF α -induced death and sustained JNK activation by inhibiting MAP kinase phosphatases. *Cell* 2005; 120:649–61.
36. Murphy MP. How mitochondria produce reactive oxygen species. *Biochem J* 2009;417:1–13.
37. Saleh J, Peyssonnaud C, Singh KK, et al. Mitochondria and microbiota dysfunction in COVID-19 pathogenesis. *Mitochondrion* 2020;54:1–7.
38. Mariathasan S, Newton K, Monack DM, et al. Differential activation of the inflammasome by caspase-1 adaptors ASC and Ipaf. *Nature* 2004;430:213–8.
39. Broz P, Dixit VM. Inflammasomes: mechanism of assembly, regulation and signalling. *Nat Rev Immunol* 2016;16: 407–20.
40. Martinon F, Burns K, Tschopp J. The Inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL- β . *Mol Cell* 2002;10: 417–26.
41. Sui A, Chen X, Shen J, et al. Inhibiting the NLRP3 inflammasome with MCC950 ameliorates retinal neovascularization and leakage by reversing the IL-1 β /IL-18 activation pattern in an oxygen-induced ischemic retinopathy mouse model. *Cell Death Dis* 2020;11:901.

42. Shaw MH, Reimer T, Kim YG, et al. NOD-like receptors (NLRs): bona fide intracellular microbial sensors. *Curr Opin Immunol* 2008;20:377–82.
43. Swanson KV, Deng M, Ting JPY. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. *Nat Rev Immunol* 2019;19:477–89.
44. Kauppinen A. Mitochondria-associated inflammasome activation and its impact on aging and age-related diseases. In: Fulop T, Franceschi C, Hirokawa K, Pawelec G, eds. *Handbook of immunosenescence*. Cham: Springer International Publishing; 2018:1–20.
45. Ouyang L, Gong J. Mitochondrial-targeted ubiquinone: a potential treatment for COVID-19. *Med Hypotheses* 2020; 144:110161.
46. Singh KK, Chaubey G, Chen JY, et al. Decoding sars-cov-2 hijacking of host mitochondria in covid-19 pathogenesis. *Am J Physiol Cell Physiol* 2020;319:C258–C267.
47. Ganji R, Reddy PH. Impact of COVID-19 on mitochondrial-based immunity in aging and age-related diseases. *Front Aging Neurosci* 2020;12:614650.
48. Zhao N, Enns CA. Iron transport machinery of human cells, players and their interactions. In: Orlov S, ed. *Current topics in membranes*. Vol. 69. San Diego: Academic Press Inc.; 2012:67–93.
49. Collins JF, Anderson GJ. Molecular mechanisms of intestinal iron transport. In: Johnson LR, ed. *Physiology of the gastrointestinal tract*. Vol. 2. London: Elsevier Inc.; 2012: 1921–1947.
50. Garrick MD. Human iron transporters. *Genes Nutr* 2011;6: 45–54.
51. Liuzzi JP, Aydemir F, Nam H, et al. Zip14 (Slc39a14) mediates non-transferrin-bound iron uptake into cells. *Proc Natl Acad Sci USA* 2006;103:13612–7.
52. McKie AT, Barrow D, Latunde-Dada GO, et al. An iron-regulated ferric reductase associated with the absorption of dietary iron. *Science* 2001;291:1755–9.
53. Ji C, Kosman DJ. Molecular mechanisms of non-transferrin-bound and transferrin-bound iron uptake in primary hippocampal neurons. *J Neurochem* 2015;133:668–83.
54. Vargas JD, Herpers B, McKie AT, et al. Stromal cell-derived receptor 2 and cytochrome b561 are functional ferric reductases. *Biochim Biophys Acta* 2003;1651:116–23.
55. Mackenzie EL, Iwasaki K, Tsuji Y. Intracellular iron transport and storage: from molecular mechanisms to health implications. *Antioxid Redox Signal* 2008;10:997–1030.
56. Arosio P, Elia L, Poli M. Ferritin, cellular iron storage and regulation. *IUBMB Life* 2017;69:414–22.
57. De Domenico I, McVey Ward D, Kaplan J. Regulation of iron acquisition and storage: consequences for iron-linked disorders. *Nat Rev Mol Cell Biol* 2008;9:72–81.
58. Bystrom LM, Guzman ML, Rivella S. Iron and reactive oxygen species: friends or foes of cancer cells? *Antioxid Redox Signal* 2014;20:1917–24.
59. Dixon SJ, Stockwell BR. The role of iron and reactive oxygen species in cell death. *Nat Chem Biol* 2014;10:9–17.
60. Cairo G, Recalcati S. Iron-regulatory proteins: molecular biology and pathophysiological implications. *Exp Rev Mol Med* 2007;9:1–13.
61. Zhou ZD, Tan EK. Iron regulatory protein (IRP)-iron responsive element (IRE) signaling pathway in human neurodegenerative diseases. *Mol Neurodegener* 2017;12:1–12.
62. Yanatori I, Richardson DR, Imada K, et al. Iron export through the transporter ferroportin 1 is modulated by the iron chaperone PCBP2. *J Biol Chem* 2016;291:17303–18.
63. Daru J, Colman K, Stanworth SJ, et al. Serum ferritin as an indicator of iron status: what do we need to know? *Am J Clin Nutr* 2017;106:1634S–9S. Vol
64. Paradkar PN, Zumbrennen KB, Paw BH, et al. Regulation of mitochondrial iron import through differential turnover of mitoferrin 1 and mitoferrin 2. *Mol Cell Biol* 2009;29: 1007–16.
65. Shaw GC, Cope JJ, Li L, et al. Mitoferrin is essential for erythroid iron assimilation. *Nature* 2006;440:96–100.
66. Gao G, Chang YZ. Mitochondrial ferritin in the regulation of brain iron homeostasis and neurodegenerative diseases. *Front Pharmacol* 2014;5:19.
67. Chen C, Paw BH. Cellular and mitochondrial iron homeostasis in vertebrates. *Biochim Biophys Acta* 2012;1823: 1459–67.
68. Walter PB, Knutson MD, Paler-Martinez A, et al. Iron deficiency and iron excess damage mitochondria and mitochondrial DNA in rats. *Proc Natl Acad Sci USA* 2002;99: 2264–9.
69. Vlahakos VD, Marathias KP, Arkadopoulos N, et al. Hyperferritinemia in patients with COVID-19: an opportunity for iron chelation? *Artif Organs* 2021;45:163–7. aor.
70. Abobaker A. Can iron chelation as an adjunct treatment of COVID-19 improve the clinical outcome? *Eur J Clin Pharmacol* 2020;76:1619–20.
71. Alfarouk KO, Verduzco D, Rauch C, et al. Glycolysis, tumor metabolism, cancer growth and dissemination. A new pH-based etiopathogenic perspective and therapeutic approach to an old cancer question. *Oncoscience* 2014;1: 777–802.
72. Alfarouk KO. Tumor metabolism, cancer cell transporters, and microenvironmental resistance. *J Enzyme Inhibit Med Chem* 2016;31:859–8.
73. Hertz L, Diemel GA. Lactate transport and transporters: general principles and functional roles in brain cells. *J Neurosci Res* 2005;79:11–8.
74. Bosshart PD, Kalbermatter D, Bonetti S, et al. Mechanistic basis of L-lactate transport in the SLC16 solute carrier family. *Nature Commun* 2019;10:2649.
75. Haas R, Smith J, Rocher-Ros V, et al. Lactate regulates metabolic and proinflammatory circuits in control of T cell migration and effector functions. *PLoS Biol* 2015;13: e1002202.
76. Pucino V, Bombardieri M, Pitzalis C, et al. Lactate at the crossroads of metabolism, inflammation, and autoimmunity. *Eur J Immunol* 2017;47:14–21.
77. Gerner RR, Klepsch V, Macheiner S, et al. NAD metabolism fuels human and mouse intestinal inflammation. *Gut* 2018; 67:1813–23.
78. Adriouch S, Hubert S, Pechberty S, et al. NAD⁺ released during inflammation participates in T cell homeostasis by inducing ART2-mediated death of naive T cells in vivo. *J Immunol* 2007;179:186–94.
79. Henry BM, Aggarwal G, Wong J, et al. Lactate dehydrogenase levels predict coronavirus disease 2019 (COVID-19) severity and mortality: a pooled analysis. *Am J Emerg Med* 2020;38:1722–6.
80. Booth AL, Abels E, McCaffrey P. Development of a prognostic model for mortality in COVID-19 infection using machine learning. *Modern Pathol* 2021;34:522–31.

81. Romero-Garcia S, Prado-Garcia H. Mitochondrial calcium: transport and modulation of cellular processes in homeostasis and cancer (Review). *Int J Oncol* 2019;54:1155–67.
82. Paupe V, Prudent J. New insights into the role of mitochondrial calcium homeostasis in cell migration. *Biochem Biophys Res Commun* 2018;500:75–86.
83. Contreras L, Drago I, Zampese E, et al. Mitochondria: the calcium connection. *Biochim Biophys Acta* 2010;1797:607–18.
84. Gandhi S, Wood-Kaczmar A, Yao Z, et al. PINK1-associated Parkinson's disease is caused by neuronal vulnerability to calcium-induced cell death. *Mol Cell* 2009;33:627–38.
85. Dagda RK, Cherra SJ, Kulich SM, et al. Loss of PINK1 function promotes mitophagy through effects on oxidative stress and mitochondrial fission. *J Biol Chem* 2009;284:13843–55.
86. Gelmetti V, De Rosa P, Torosantucci L, et al. PINK1 and BECN1 relocate at mitochondria-associated membranes during mitophagy and promote ER-mitochondria tethering and autophagosome formation. *Autophagy* 2017;13:654–69.
87. Jouaville LS, Pinton P, Bastianutto C, et al. Regulation of mitochondrial ATP synthesis by calcium: evidence for a long-term metabolic priming. *Proc Natl Acad Sci USA* 1999;96:13807–12.
88. Kennedy ED, Rizzuto R, Theler JM, et al. Glucose-stimulated insulin secretion correlates with changes in mitochondrial and cytosolic Ca^{2+} in aequorin-expressing INS-1 cells. *J Clin Invest* 1996;98:2524–38.
89. Yang C, Ma X, Wu J, et al. Low serum calcium and phosphorus and their clinical performance in detecting COVID-19 patients. *J Med Virol* 2021;93:1639–51.
90. Gaffney PJ, Brasher M. Subunit structure of the plasmin-induced degradation products of crosslinked fibrin. *BBA* 1973;295:308–13.
91. Gaffney PD. dimer. History of the discovery, characterisation and utility of this and other fibrin fragments. *Fibrinol Proteol* 1993;7:2–8.
92. Haverkate F, Timan G. Protective effect of calcium in the plasmin degradation of fibrinogen and fibrin fragments D. *Thrombosis Res* 1977;10:803–12.
93. Favresse J, Lippi G, Roy P-M, et al. D-dimer: preanalytical, analytical, postanalytical variables, and clinical applications. *Crit Rev Clin Lab Sci* 2018;55:548–77.
94. Kelly J, Rudd A, Lewis RR, et al. Plasma D-dimers in the diagnosis of venous thromboembolism. *Arch Intern Med* 2002;162:747–56.
95. Pulivarthi S, Gurram MK. Effectiveness of D-dimer as a screening test for venous thromboembolism: an update. *North Am J Med Sci* 2014;6:491–9.
96. Matsuo H, Nakajima Y, Ogawa T, et al. Evaluation of D-dimer in screening deep vein thrombosis in hospitalized Japanese patients with acute medical diseases/Episodes. *Ann Vasc Dis* 2016;9:193–200.
97. Cho ES, McClelland PH, Cheng O, et al. Utility of D-dimer for diagnosis of deep vein thrombosis in coronavirus disease-19 infection. *J Vasc Surg* 2021;9:47–53.
98. Yu HH, Qin C, Chen M, et al. D-dimer level is associated with the severity of COVID-19. *Thrombosis Res* 2020;195:219–25.
99. Yao Y, Cao J, Wang Q, et al. D-dimer as a biomarker for disease severity and mortality in COVID-19 patients: a case control study. *J Intens Care* 2020;8:49.
100. Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 2020;395:1054–62.
101. Wang Q, Zennadi R. Oxidative stress and thrombosis during aging: the roles of oxidative stress in RBCs in venous thrombosis. *Int J Mol Sci* 2020;21:1–22.
102. Lee KH, Kim JR. Reactive oxygen species regulate the generation of urokinase plasminogen activator in human hepatoma cells via MAPK pathways after treatment with hepatocyte growth factor. *Exp Mol Med* 2009;41:180–8.
103. Tykhomyrov AA, Zhernosekov DD, Guzyk MM, et al. Plasminogen modulates formation of reactive oxygen species in human platelets. *Ukrain Biochem J* 2018;90:31–40.
104. Wijnker PJM, Sequeira V, Foster DB, et al. Length-dependent activation is modulated by cardiac troponin I bisphosphorylation at Ser23 and Ser24 but not by Thr143 phosphorylation. *Am J Physiol* 2014;306:H1171–H1181.
105. Sun YB, Lou F, Irving M. Calcium- and myosin-dependent changes in troponin structure during activation of heart muscle. *J Physiol* 2009;587:155–63.
106. Korff S, Katus HA, Giannitsis E. Differential diagnosis of elevated troponins. *Heart* 2006;92:987–93.
107. Skeik N, Patel DC. A review of troponins in ischemic heart disease and other conditions. *Int J Angiol* 2007;16:53–8.
108. Daubert MA, Jeremias A. The utility of troponin measurement to detect myocardial infarction: review of the current findings. *Vasc Health Risk Manage* 2010;6:691–9.
109. Okamoto R, Hirashiki A, Cheng XW, et al. Usefulness of serum cardiac troponins T and I to predict cardiac molecular changes and cardiac damage in patients with hypertrophic cardiomyopathy. *Int Heart J* 2013;54:202–6.
110. Pankuweit S, Richter A. Mitochondrial disorders with cardiac dysfunction: an under-reported aetiology with phenotypic heterogeneity. *Eur Heart J*. 36:2894–7.
111. Amgalan D, Pekson R, Kitsis RN. Troponin release following brief myocardial ischemia: apoptosis versus necrosis. *JACC* 2017;2:118–21.
112. Park KC, Gaze DC, Collinson PO, et al. Cardiac troponins: from myocardial infarction to chronic disease. *Cardiovasc Res* 2017;113:1708–18.
113. Tersalvi G, Vicenzi M, Calabretta D, et al. Elevated troponin in patients with coronavirus disease 2019: possible mechanisms. *J Cardiac Fail* 2020;26:470–5.
114. Lombardi CM, Carubelli V, Iorio A, et al. Association of troponin levels with mortality in Italian patients hospitalized with coronavirus disease 2019: results of a multicenter study. *JAMA Cardiol* 2020;5:1274–E7.
115. Mannu GS. The non-cardiac use and significance of cardiac troponins. *Scott Med J* 2014;59:172–8.
116. Collado S, Arenas MD, Barbosa F, et al. COVID-19 in grade 4–5 chronic kidney disease patients. *Kidney Blood Pressure Res* 2020;45:768–74.
117. Ajaimy M, Melamed ML. Covid-19 in patients with kidney disease. *Clin J Am Soc Nephrol* 2020;15:1087–9.
118. Alfarouk KO, Shayoub MEA, Muddathir AK, et al. Evolution of tumor metabolism might reflect carcinogenesis as a reverse evolution process (dismantling of multicellularity). *Cancers* 2011;3:3002–17.
119. Andreux PA, Houtkooper RH, Auwerx J. Pharmacological approaches to restore mitochondrial function. *Nat Rev Drug Discovery* 2013;12:465–83.

120. Reshkin SJ, Bellizzi A, Caldeira S, et al. Na⁺/H⁺ exchanger-dependent intracellular alkalinization is an early event in malignant transformation and plays an essential role in the development of subsequent transformation-associated phenotypes. *FASEB J* 2000;14:2185–97.
121. Cardone RAR, Alfarouk KOK, Elliott RLR, et al. The role of sodium hydrogen exchanger 1 in dysregulation of proton dynamics and reprogramming of cancer metabolism as a sequela. *Int J Mol Sci* 2019;20:3694.
122. Alvarez BV, Villa-Abrille MC. Mitochondrial NHE1: a newly identified target to prevent heart disease. *Front Physiol* 2013;4:152.
123. Wilson L, Gage P, Ewart G. Hexamethylene amiloride blocks E protein ion channels and inhibits coronavirus replication. *Virology* 2006;353:294–306.
124. Haddad JJ, Land SC. Amiloride blockades lipopolysaccharide-induced proinflammatory cytokine biosynthesis in an I κ B- α /NF- κ B-dependent mechanism: evidence for the amplification of an antiinflammatory pathway in the alveolar epithelium. *Am J Respir Cell Mol Biol* 2002;26:114–26.
125. Jankun J, Skrzypczak-Jankun E. Molecular basis of specific inhibition of urokinase plasminogen activator by amiloride. *Cancer Biochem Biophys* 1999;17:109–23.
126. Vassalli JD, Belin D. Amiloride selectively inhibits the urokinase-type plasminogen activator. *FEBS Lett* 1987;214:187–91.
127. Javadov S, Choi A, Rajapurohitam V, et al. NHE-1 inhibition-induced cardioprotection against ischaemia/reperfusion is associated with attenuation of the mitochondrial permeability transition. *Cardiovasc Res* 2007;77:416–24.
128. Bencze G, Bencze S, Rivera KD, et al. Mito-oncology agent: fermented extract suppresses the Warburg effect, restores oxidative mitochondrial activity, and inhibits in vivo tumor growth. *Sci Rep* 2020;10:14174.
129. Mueller T, Voigt W. Fermented wheat germ extract – nutritional supplement or anticancer drug? *Nutr J* 2011;10:89.
130. Sukkar SG, Cella F, Rovera GM, et al. A multicentric prospective open trial on the quality of life and oxidative stress in patients affected by advanced head and neck cancer treated with a new benzoquinone-rich product derived from fermented wheat germ (Aveamar). *Mediterranean J Nutr Metab* 2008;1:37–42.
131. Jeong H-Y, Choi Y-S, Lee J-K, et al. Anti-inflammatory activity of citric acid-treated wheat germ extract in lipopolysaccharide-stimulated macrophages. *Nutrients* 2017;9:730.
132. Mijnhout GS, Kollen BJ, Alkhalaf A, et al. Alpha lipoic acid for symptomatic peripheral neuropathy in patients with diabetes: a meta-analysis of randomized controlled trials. *Int J Endocrinol* 2012;2012:2012.
133. Zhong M, Sun A, Xiao T, et al. A randomized, single-blind, group sequential, active-controlled study to evaluate the clinical efficacy and safety of α -Lipoic acid for critically ill patients with coronavirus disease 2019 (COVID-19). *medRxiv* 2020:2020.04.15.20066266.
134. Schwartz L, Seyfried T, Alfarouk KO, et al. Out of Warburg effect: an effective cancer treatment targeting the tumor specific metabolism and dysregulated pH. *Semin Cancer Biol* 2017;43:134–8.
135. Schwartz L, Supuran CT, Alfarouk KO. The Warburg effect and the hallmarks of cancer. *Anti-Cancer Agents Med Chem* 2017;17:164–70.
136. Gendrot M, Andreani J, Dufloy I, et al. Methylene blue inhibits replication of SARS-CoV-2 in vitro. *Int J Antimicrob Agents* 2020;56:106202.
137. Henry M, Summa M, Patrick L, et al. A cohort of cancer patients with no reported cases of SARS-CoV-2 infection: the possible preventive role of Methylene Blue. *Substantia* 2020;4:888.
138. Warburg O. On the origin of cancer cells. *Science* 1956;123:309–14.
139. Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* 2009;324:1029–1033.
140. Dhup S, Dadhich RK, Porporato PE, et al. Multiple biological activities of lactic acid in cancer: influences on tumor growth, angiogenesis and metastasis. *Curr Pharm Des* 2012;18:1319–30.
141. Ayres JS. A metabolic handbook for the COVID-19 pandemic. *Nat Metab* 2020;2:572–585.
142. Alfarouk KO, Muddathir AK, Shayoub MEA. Tumor acidity as evolutionary spite. *Cancers* 2011;3:408–414.