

Arsenic behavior across soil-water interfaces in paddy soils: coupling, decoupling and speciation

YUAN, Zhao-Feng, GUSTAVE, Williamson, BOYLE, John, SEKAR, Raju http://orcid.org/0000-0002-1182-9004, BRIDGE, Jonathan http://orcid.org/0000-0003-3717-519X, REN, Yuxiang, TANG, Xianjin, GUO, Bin and CHEN, Zheng

Available from Sheffield Hallam University Research Archive (SHURA) at: https://shura.shu.ac.uk/27526/

This document is the Accepted Version [AM]

Citation:

YUAN, Zhao-Feng, GUSTAVE, Williamson, BOYLE, John, SEKAR, Raju, BRIDGE, Jonathan, REN, Yuxiang, TANG, Xianjin, GUO, Bin and CHEN, Zheng (2020). Arsenic behavior across soil-water interfaces in paddy soils: coupling, decoupling and speciation. Chemosphere, p. 128713. [Article]

Copyright and re-use policy

See http://shura.shu.ac.uk/information.html

1 Title: Arsenic behavior across soil-water interfaces in paddy soils:

2 coupling, decoupling and speciation

- 3 Names of authors: Zhao-Feng Yuan 1, 2, Williamson Gustave 1, 2, 3, John Boyle 4, Raju Sekar 5,
- 4 Jonathan Bridge ⁶, Yuxiang Ren ¹, Xianjin Tang ⁷, Bin Guo ⁸ * and Zheng Chen ¹ *
- 5 Department of Health and Environmental Sciences, Xi'an Jiaotong-Liverpool University, 111 Ren'ai
- 6 Road, Suzhou, Jiangsu 215123, China.
- ⁷ Department of Environmental Science, University of Liverpool, Brownlow Hill, Liverpool L69 7ZX,
- 8 UK.
- 9 ³ Chemistry, Environmental & Life Sciences, University of The Bahamas, New Providence, Nassau,
- 10 The Bahamas.
- ⁴ Department of Geography & Planning, University of Liverpool, Roxby Building, Liverpool, L69 7ZT,
- 12 UK.
- ⁵ Department of Biological Sciences, Xi'an Jiaotong-Liverpool University, 111 Ren'ai Road, Suzhou,
- 14 Jiangsu 215123, China.
- 15 ⁶ Department of Natural and Built Environment, Sheffield Hallam University, Howard St, Sheffield S1
- 16 1WB, UK.

- 17 Institute of Soil and Water Resources and Environmental Science, Zhejiang Provincial Key
- Laboratory of Agricultural Resources and Environment, Zhejiang University, 866 Yuhangtang Road,
- 19 Hangzhou 310058, China.
- ⁸ Institute of Environment, Resource, Soil and Fertilizer, Zhejiang Academy of Agricultural Sciences,
- 21 Hangzhou, 310021, China.
- 23 * Corresponding author: Bin Guo (Email: ndgb@163.com); Zheng Chen (E-mail:
- 24 <u>ebiogeochem@outlook.com</u> or <u>Zheng.Chen@xjtlu.edu.cn</u>; Tel: +86-512-81880471; fax:
- 25 +86-512-88161899).

Abstract

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

The sharp redox gradient at soil-water interfaces (SWI) plays a key role in controlling arsenic (As) translocation and transformation in paddy soils. When Eh drops, As is released to porewater from solid iron (Fe) and manganese (Mn) minerals and reduced to arsenite. However, the coupling or decoupling processes operating within the redox gradient at the SWI in flooded paddy soils remain poorly constrained due to the lack of direct evidence. In this paper, we reported the mm-scale mapping of Fe, As and other associated elements across the redox gradient in the SWI of five different paddy soils. The results showed a strong positive linear relationship between dissolved Fe, Mn, As, and phosphorus (P) in 4 out of the 5 paddy soils, indicating the general coupling of these elements. However, decoupling of Fe, Mn and As was observed in one of the paddy soils. In this soil, distinct releasing profiles of Mn, As and Fe were observed, and the releasing order followed the redox ladder. Further investigation of As species showed the ratio of arsenite to total As dropped from 100% to 75.5% and then kept stable along depth of the soil profile, which indicates a dynamic equilibrium between arsenite oxidization and arsenate reduction. This study provides direct evidence of multi-elements' interaction along redox gradient of SWI in paddy soils.

43

Keywords: arsenic, iron, profile, paddy soil, coupling, decoupling

45

1. Introduction

46

Paddy fields globally represent artificial wetlands, supporting the growth of rice 47 (Bouman and Tuong, 2001). Paddy agriculture for the production of rice involves 48 intermittent flooding to favor the growth of rice and in some cases to reduce the 49 bioavailability of cadmium (Hu et al., 2015). During the flooding stage, 50 redox-sensitive elements in the soil, such as iron (Fe) and manganese (Mn), may 51 become mobile from the solid into the liquid phase (Xu et al., 2017; Zhang et al., 52 2018), and lead to dissolution of the sorbates (e.g. arsenic (As), and phosphorus (P)) 53 bound on the solid phase (act as sorbents) (Darland and Inskeep, 1997; Polizzotto et 54 al., 2008). Other redox-sensitive elements, such as sulfur (S), may become immobile 55 56 from the liquid to solid phase, and result in immobilization of dissolved elements through co-precipitation (e.g. FeS, As₂S₃) or adsorption (Burton et al., 2008). Among 57 the noted elements, the mobilization of As is of particular concern since rice plants 58 59 can efficiently uptake and accumulate As in their rice grains (Honma et al., 2016; Gustave et al., 2019a). This poses a threat to human health since long-term As 60 exposure is known to cause cancer and organ failure (Meharg, 2004; Roberts et al., 61 2010). Understanding the biogeochemical cycling of As in paddy soils under 62 heterogeneous and transient geochemical conditions during flooding and drainage is 63 therefore of high importance in predicting and thus mitigating its uptake into food 64 crops. 65

Arsenic behavior in saturated soils is closely coupled to that of Fe oxides. This occurs because Fe oxides provide the main adsorption sites for As (Zobrist et al., 2000; Chen et al., 2006; Tufano et al., 2008). When paddy soils are flooded, Fe oxides are reduced and dissolved by dissimilatory Fe reducing bacteria, leading to the simultaneous release of adsorbed As into porewater (Takahashi et al., 2004). The term "coupling of Fe and As" was used to describe this phenomenon, in which the release of Fe and As into the soil porewater shows a close positive correlation (Weber et al., 2010). In contrast, the term "decoupling of Fe and As" was defined to refer to the lack of a linear correlation between dissolved Fe and As concentrations (Weber et al., 2010).

Coupling of Fe and As has been well documented in soils, yet the decoupling process remains not well resolved (Weber et al., 2010; Bennett et al., 2012). To clarify the mechanisms regulating the potential decoupling of As and Fe, simplified water-Fe minerals-bacteria systems were employed (Tufano et al., 2008; Tufano and Scott, 2008; Weber et al., 2010), which indicated that coupling-decoupling is thermodynamically controlled. For example, Zobrist et al. identified *sulfurospirillum barnesii* was able to reduce adsorbed arsenate [As(V)] to arsenite [As(III)] without requiring dissolution of the host Fe oxides (Zobrist et al., 2000), which could be caused by an energy trade-off between reduction of adsorbed As(V) and host Fe minerals by microorganisms (Campbell et al., 2006) (Islam et al., 2004). However, to date this has not been confirmed in soils.

In saturated soils, it has been a challenge to distinguish the potential decoupling between Fe and As. Homogeneous saturated soils were frequently used to investigate the time-dependent behaviors of Fe and As (Masscheleyn et al., 1991; White et al., 2007; Weber et al., 2010; Das et al., 2016; Honma et al., 2016; Xu et al., 2017), yet the potential decoupling of As and Fe was only demonstrated in few deep aquifers (White et al., 2007; Das et al., 2016). In most cases, the time-series investigation is unable to capture the time points and/or hotspots that decoupling of Fe and As occurs. The unsuccessful identification of the decoupling process is not surprising, because the Fe reduction and As releasing usually happen in sequence with little temporal offset (Zhang et al., 2018). The dynamic change of Fe and As makes it hard to catch the appropriate time point, even if the decoupling of Fe and As occurs.

When flooded, the bulk soil becomes reduced owing to the abundant organic matters (Frenzel et al., 1992). At the same time, a stable redox gradient is formed between the O₂-rich surface water and subsurface soil, which could significantly influence the spatial distribution of redox-sensitive elements (Mucci et al., 2000; Widerlund and Davison, 2007; Arsic et al., 2018). Hence, the narrow but stable soil water interface (SWI) is an ideal place to study the coupling/decoupling processes of Fe and As along depth (Bennett et al., 2012; Gorny et al., 2015). Although As(V) reduction theoretically happened at higher redox potential (Eh) compared to Fe(III) (Borch et al., 2010), few studies have detected the spatial separation of Fe(II) and As species in SWI of natural soils. This could be attributed to the lack of a suitable

method to simultaneously measure fine-scale Fe, As and other associated elements across SWI (Gorny et al., 2015).

We recently developed a high-throughput method to simultaneously measure concentrations of multi-element (e.g. Fe, Mn, As, P and S) and their species across SWI (Yuan et al., 2019; Yuan et al., 2021). The SWI profiler used in our method shows a big advantage in non-destructively and repeatedly sampling of porewater at high-resolution (HR) (mm level) across SWI (Yuan et al., 2019), compared with traditional HR samplers (e.g. DET, peeper). The analytical technique in our method makes it possible to extract all interested parameters from volume-limited (~ 100 μL) samples collected by HR samplers (Yuan et al., 2021). In this study, the method was applied to reveal the coupling/decoupling process of Fe and As, as well as other associated elements, including Mn, S and P. Their profiles across the SWI in five paddy soils were mapped to 1) illustrate multi-element and Eh profiles in typical saturated paddy soils; 2) study the coupling/decoupling behaviors among the redox-sensitive elements and 3) reveal the As speciation process across the SWI.

2. Materials and methods

2.1 Reagents, materials and solutions. Analytical grade reagents were purchased from Aladdin Chemical Reagent Co., Ltd. (Shanghai, China), unless otherwise stated. Calibration standards, including As species, Fe, Mn, P, and S, were supplied by

Guobiao (Beijing) Testing & Certification Co., Ltd (Beijing, China). All solutions were prepared with ultrapure water (18.2 M Ω cm, Millipore Corp., Bedford, USA) deoxygenated by bubbling pure N_2 overnight.

Paddy soils with different levels of As (25.6 - 146 mg·kg⁻¹, Table S1) were collected from Wenshan (WS, 23°45'N, 105°26'E), Bijie (BI, 26°39'N, 105°47'E), Wuxue (WX, 29°59'N, 115°38'E), Shaoguan (SG, 25°6'N, 113°38'E) and Ganzhou (GZ, 25°30'N, 114°36'E), China. The plow layer soil (0 - 20 cm) was sampled and directly transported to the laboratory. Soils were wet mixed and sieved through a 1.0 mm diameter sieve. The wet sieved soils were mixed evenly and each soil type was added to three black plastic pots (inner diameter × height = 13 cm × 21 cm), with a soil depth of ~15 cm (total 15 pots). Table S1 shows the selected soil properties. The soils were flooded with ultrapure water and allowed to stabilize for 30 d in dark conditions, with a constant room temperature (22 °C) controlled by an air-conditioner.

2.2 Deployment and sampling of SWI profiler. The SWI profiler is a recently developed HR porewater sampler (Yuan et al., 2019). It has a sampling depth of 60 mm, with a spatial resolution of 1.7 mm. The SWI profiler was provided by Tidu Environment Inc. (Suzhou, China). Fifteen SWI profilers were inserted, one per pot, into the 15 pots comprising three replicates for each of WS, BI, WX, SG, and GZ paddy soils, such that the uppermost 10 mm of the profile length was in overlying water and the lower 50 mm buried in saturated soil. After 30 d of incubation,

porewater was sampled by SWI profiler. Eh profiles were simultaneously measured using a custom-made platinum micro-electrode with an Ag/AgCl reference electrode.

Before sampling, O₂-free ultrapure water was pumped into SWI profiler as the carrier solution, with an injection pump (TYD01, Lei Fu, China). After loading of the carrier solution, small ions and molecules can passively diffuse from porewater into SWI profiler driving by the concentration gradient. When the diffusion process reached equilibrium (after 24 h), the sample was pumped out by the injection pump (Fig. S1). The sample acidified with O₂-free HCl in the first sampling event was used for total elements analysis (Gustave et al., 2019b). In the second sampling event (24 h after the first one), the sample was preserved with O₂-free EDTA to complex metals (e.g. Fe), which would facilitate As species analysis under alkali conditions (Gallagher et al., 2001; Yuan et al., 2021). The collected samples were preserved at 4 °C fridge before downstream instrumental analysis.

2.3 Total elements analysis. Total Fe, Mn, As, P, and S obtained from SWI profiler samples were measured by inductively coupled plasma-mass spectrometry (ICP-MS, NexION 350X, PerkinElmer, Inc., Shelton, CT USA). The sample in 0.6 mL centrifuge tube (~ 100 μL) was introduced into ICP-MS by a PFA-200 Microflow Nebulizer for elemental analysis. The counts of ⁵⁷Fe⁺, ⁵⁵Mn⁺, ⁴⁷PO⁺, ⁴⁸SO⁺ and ⁹¹AsO⁺ were recorded in dynamic reaction cell (DRC) or extended dynamic range (EDR) mode (Yuan et al., 2021). Spiked standards were measured after every 30 samples as a quality control measure.

The ICP-MS was optimized to the following conditions: DRC (O ₂ , gas flow: 1.0
mL·min ⁻¹); data only analysis; RF power: 1600W; plasma gas flow rate: 15 L·min ⁻¹ ;
auxiliary gas flow: 1.2 L·min-1; nebulized gas flow: 0.90 L·min-1; nickel sampling and
skimmer cones were used. A voltage parameter in EDR mode, called "rejection
parameter a" (Rpa), was optimized to 0.01 for Fe, Mn and S to enable simultaneous
measurement of majors (e.g. Fe, Mn, S) and traces (e.g. As) by ICP-MS, following the
procedure described in Yuan et al. (2021).
2.4 Speciation of As in soil porewater. For As speciation analysis, the $\sim 100~\mu L$
sample was manually loaded into a 25 μL sample loop of ion chromatography (IC,
Dionex ICS-1100, Thermo Scientific, USA) (Yuan et al., 2021). The IC system
consisted of an anion-exchange column (IonPac AS23, 250mm×4mm, Dionex).
Twenty mM NH_4HCO_3 (pH = 10) were used as the mobile phase (Suzuki et al., 2009;
Yuan et al., 2021). We expanded the comparison of As speciation in this study to
digitized As species published in previous reports with the Engauge Digitizing
software (version 11.3) used in a previous paddy soil study (Yuan et al., 2016).
2.5 Statistical analysis. Data were analyzed and plotted using R software (version
3.5.0). Standard errors were used to show the variance, and linear regression analysis

3. Results and discussion

was used to identify the coupling and decoupling process of different elements.

3.1 Vertical changes of Eh and elements across the SWI. The Eh (vs. Ag/AgCl

reference electrode) and multi-element were measured and depicted in Fig. 1.

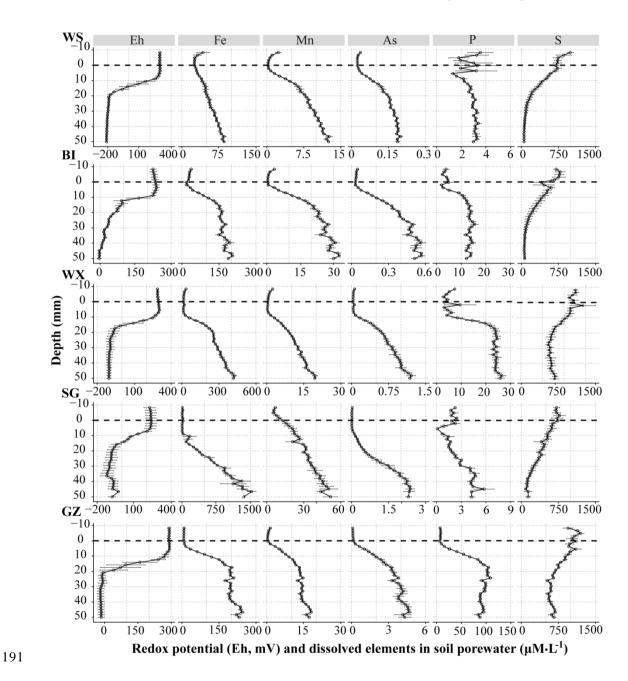


Figure 1 Redox potential (Eh, mV, vs. Ag/AgCl reference electrode) and aqueous iron (Fe), manganese (Mn), arsenic (As), phosphorus (P), sulfur (S) (μ M·L⁻¹) across the soil-water interface. WS, BI, WX, SG and GZ represent soils collected from Wenshan, Bijie, Wuxue, Shaoguan and Ganzhou paddies respectively. The black dotted line at depth = 0 represents the soil-water interface. The error bar is standard error (n = 3). Note the different axis scales used.

The Eh changed rapidly from highly oxidizing (~300 mV) to reducing conditions along the depth of the SWI. However, the reducing levels varied among the different paddy soils. Highly reducing condition (~-150 mV) was observed in WS/WX/SG, yet a relatively moderate one (~-10 mV) in BI/GZ. This difference in Eh might be caused by different physico-chemical properties of soils. Manganese and Fe are important factors for regulating soil redox (Brannon et al., 1984; Xu et al., 2017). Here, we found high Mn (6.32 g·Kg⁻¹, WS) or Fe (> 70 g·Kg⁻¹, WX/SG) alone was unable to buffer the decrease of soil redox (Table S1). Buffering of soil redox was only achieved with both high Fe and Mn (> 70 and 1.10 g·Kg⁻¹) in BI/GZ. Although Mn is believed to be the key for buffering soil redox (Xu et al., 2017), the formation of Fe oxides coating with the nucleating agent is essential for nucleation and growth of Mn oxides (Burns and Burns, 1975). Hence, Fe may also get involved in retarding soil redox by affecting the reactivity of Mn.

Following Eh measurement, vertical profiles of Fe, Mn, As, P and S were mapped by SWI profiler (Fig. 1). Dissolved Fe, Mn and As were almost undetectable under oxidizing conditions, however their concentrations increased to 85.2 - 1359, 12.5 - 42.0 and 0.187 - 4.31 μM·L⁻¹ respectively in reducing soils. In terms of the release trend of those elements, they all showed a subsurface increase trend, which agree well with previous reports of their typically vertical distributions along SWI (Di et al., 2012; Wu et al., 2016a; Ma et al., 2017; Arsic et al., 2018). A similar trend was also found on P. Similar to As (Xiu et al., 2016), P cycling is tightly tied to that of Fe

oxides in soils (Ding et al., 2016), due to Fe oxides provide the main adsorption sites for P and As. The reductive mobilization of Fe, Mn, P and As was expected to negatively correlate with Eh and S, nonetheless this was not always true for all the paddy soils. For instance, the most As was mobilized in GZ which had a relatively high Eh and S concentrations (\sim -15.0 mV, \sim 635 μ M·L⁻¹ S, Fig. 1GZ), while the least As was released in WS soil with an extremely low Eh and S (\sim -200 mV, \sim 54.8 μM·L⁻¹ S, Fig. 1WS). Similar observations to these results have been frequently reported when using different soil samples (Bogdan and Schenk, 2008; Xu et al., 2017). This is due to the complex process of As mobilization in soils. Although Fe oxides are considered as a major agent controlling As mobilization (Xu et al., 2017), it is also influenced by organic matter content, and formation of secondary minerals along flooding (Bogdan and Schenk, 2008). Rich soil organic matter enhance As release by accelerating biotic reduction of Fe oxides (Bogdan and Schenk, 2008). The dissolved As could be re-immobilized by forming secondary minerals (like FeS, Fe₂S, As₂S₃, siderite, vivianite etc.) (Bogdan and Schenk, 2008). Owing to the complex biogeochemical processes involved in As mobilization, future studies should consider multiple associated factors when assessing As mobilization.

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

Total dissolved S profiles have opposite trends as Fe, Mn, As and P, which decreased rapidly from as high as 1000 μ M·L⁻¹ in oxidizing conditions to a concentration of $\sim 300~\mu$ M·L⁻¹ in reducing soil conditions. High dissolved sulfide might trigger the formation of thioarsenate/thioarsenite in soil porewater (Wang et al.,

2020b). However, due to thioarsenate/thioarsenite was strongly suppressed by high dissolved Fe in reducing soils (Wang et al., 2020b), thioarsenate/thioarsenite was not detected in the oxic-anoxic transition zone in this study.

The decrease of S is presumably caused by sulfate reduction (Pester et al., 2012), during which S(VI) is transformed to insoluble sulfide minerals by sulfate reducing bacteria (Wu et al., 2016a; Wu et al., 2016b). In soils investigated, up to 85% S was immobilized in WS/BI/SG, yet only 40% in WX/GZ. This difference of S fate might be caused by the activity of S-reducing/oxidizing bacteria (Pester et al., 2012). Higher activity of S(VI)-reducing bacteria tended to transform more soluble S(VI) to S(-II) minerals (like FeS, Fe₂S etc.) (Pester et al., 2012). By contrast, higher activity of S-oxidizing bacteria can sustain a larger S(VI) pool in saturated soils (Pester et al., 2012).

3.2 Coupling of As with Fe and Mn across SWI. Arsenic showed a significant linear relationship with Fe and Mn (Fig. 2 and Table S2). Excellent positive correlations were yielded in 4 out of the 5 soils (WS, BI, WX and GZ, $R^2 \ge 0.848$, p < 0.01), showing the tight coupling of As with Fe and Mn in typically natural soils and sediments (Anawar et al., 2004; Xu et al., 2017; Arsic et al., 2018). Among the 4 soils, a steeper slope of the relationship between As and Fe (~ 0.0202 , respectively) was found in GZ soil than those in other soils (~ 0.00154) (Fig. 2A). The slopes may depend on the content of poorly crystalline Fe oxides, which have a high affinity for As (Tufano et al., 2008), and are subject to bioreduction by dissimilatory Fe reducing

bacteria (Zachara et al., 2002). Low content of poorly crystalline Fe oxides may retard As mobilization in the solid phase, even in soils rich in Fe (Khan et al., 2010).

Although close linear relationships of As and Mn were also found in WS, BI, WX and GZ soils except for SG soil (Fig. 2B), the role of Mn minerals may differ from Fe minerals. In the 4 soils, the Mn profiles are almost identical to Fe profiles, suggesting the reduction of Mn and Fe was controlled by the same group of microbial organisms (Myers and Nealson, 1988). However, the correlation of As and Fe/Mn in SG soil was not as well as those in other soils (Fig. 2). Furthermore, this result showed, unlike Fe reduction, Mn reduction doesn't follow with As release in the top layer of SG soil with low concentrations of both elements. We speculated the apparent decoupling of As with Mn may be caused by two reasons, 1) Fe oxides are still abundant when Mn minerals start to dissolute, which bind with the As desorbed from Mn minerals (Mitsunobu et al., 2020); 2) Mn minerals catalyze the oxidation of As(III) to As(V) (Chen et al., 2006), which is less mobile.

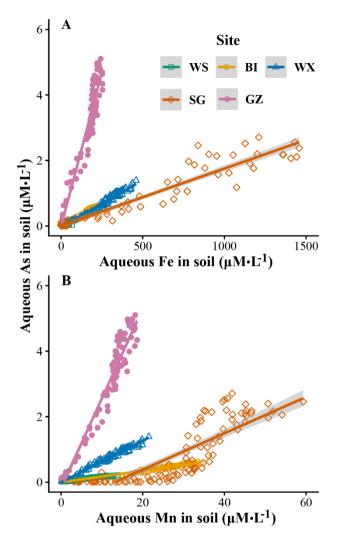


Figure 2 The correlation between vertical aqueous As with Fe (A) and Mn (B) $(\mu M \cdot L^{-1})$ in paddy soils. Soils were collected from Wenshan (WS), Bijie (BI), Wuxue (WX), Shaoguan (SG) and Ganzhou (GZ) paddies.

In most cases, the cascade of Mn, As and Fe releasing to porewater along SWI is hard to spatially distinguish because their releasing zones are often overlapped (Dočekalová et al., 2002; Gao et al., 2006; Bennett et al., 2012; Arsic et al., 2018). Thus, the SG soil, which is high in Fe and Mn, is an excellent environmental sample to study the decoupling with the dominant coupling process. Regression residual analysis revealed the decoupling process happened in the oxic-anoxic transition zone

(Table S2). Much higher residuals were obtained in that zone of SG (residuals \geq 0.265) than in other soils with a similar regression slope (including WS, BI and WX, residuals \leq 0.0417).

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

3.3 Speciation of As across SWI. To reveal the arsenic speciation along Eh gradient across SWI, we measured fine-scale As species profiles across the SWI of SG soil (Fig. 3) and the As species at 3 depth in the other 4 soils (Table S3). Surprisingly, no methyl arsenic was detected in all the samples, only As(III) and As(V) were found. Although it is known that the methylation process of As is controlled by a family of As(III) S-adenosylmethionine methyltransferases enzymes designated ArsM in microbes or AS3MT in higher eukaryotes (Ajees and Rosen, 2015; Zhao et al., 2019), the occurrence of methyl As is hard to predict in field samples. Chen et al. found that the methyl As may only exist at a narrow Eh range because, when the Eh drops below the Eh range, the methylotrophic methanogens capable of demethylating methyl As outcompete those microbes possessing As methylating ability (Chen et al., 2019), resulting in the disappearance of methyl As. According to this result, Eh only is not sufficient to induce significant methyl As synthesis in the flooded soil system, as there is no methylated As detected along the Eh gradient in all the soil samples. We speculated that the availability of organic substrate and the functional microbial community were the limiting factors in our case, more investigations are needed to identify the micro-hotspot of methyl As in paddy soils.

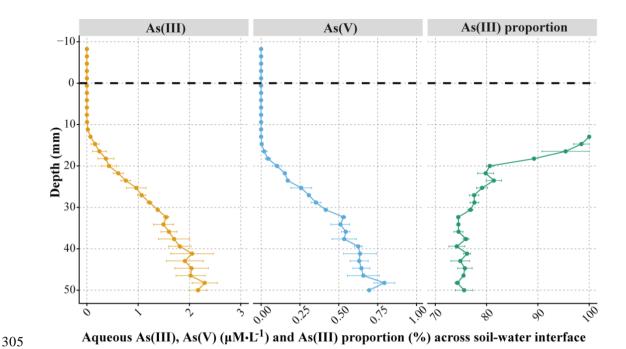


Figure 3 Profile of As species (μ M·L⁻¹) and arsenite (As(III)) proportion across the soil-water interface in Shaoguan (SG) paddy. Two As species, including As(III) and arsenate (As(V)), were detected in soil porewater.

In SG soil, the concentrations of As(III) and As(V) remained almost undetectable in the oxic zone but increased rapidly from 0.0070 (9.4 mm below SWI) and 0.0091 (15 mm below SWI) μM·L-1 to as high as 2.2 and 0.75 μM·L-1 in reducing soils, respectively. A clear As(III) release spatially prior to As(V) is observed along SWI (Fig. 3). The similar phenomenon was only reported in marine sediments by using traditional soil slicing (Chaillou et al., 2003). Bennett et al. detected As(III) release was prior to that of ferrous Fe in the oxic-anoxic transition zone (Bennett et al., 2012). Those pieces of evidence suggested a decoupling of As(III) and Fe(II), and strong coupling of As(V) and Fe(II) in flooded soils and sediments.

Generally, three main processes were proposed to explain As releasing from solid Fe/Mn oxides to liquid phase. First, desorption of As from solid phase due to

repartitioning of adsorbed As between solid and liquid phase (Fig. 4A&E) (Zobrist et al., 2000; Williams et al., 2011). Second, the Fe(III) in the complex of Fe/Mn-As is reduced to Fe(II) following As release (Fig. 4B&F) (Xu et al., 2011; Gustave et al., 2018). Third, As(V) binding on Fe oxides is reduced to As(III), thus desorbed (Fig. 4C) (Zhang et al., 2018). According to the redox ladder (Borch et al., 2010), the third process is thermodynamically favored, which has been proven with pure minerals (Tufano et al., 2008; Tufano and Scott, 2008). To the best of our knowledge, the phenomenon has not been observed in saturated soils, because the latter two processes were considered to occur simultaneously (Weber et al., 2010). The data presented in Fig. 3 might be the first evidence from saturated soils supporting that the reduction of As(V) happens on solid minerals instead of in solution.

Based on the As(III) and As (V) profiles, the proportion of As(III) in total As was calculated and depicted in Fig. 3. The proportion change along SWI could be divided into three stages: 1) a rapid decrease from 100 to 80.6% in the first 10 - 20 mm topsoil; 2) a slow decrease from 80.6 to 75.5% in 20 - 30 mm soils and 3) a stable value (75.0%) in deep soils (30 - 50 mm). In 30 - 50 mm below SWI (Fig. 3), the stable values of As concentration (As(III) and As(V): 2.40 and 0.498 μM·L⁻¹ respectively) and As species in total As (75% As(III), 25% As(V)) indicate a dynamic equilibrium of aqueous As(III) oxidation *vs.* As(V) reduction and their immobilization *vs.* mobilization in deep soils. The immobilization process is most likely stimulated by the formation of secondary sulfide S(-II) minerals (e.g. FeS) (Fig. 4), since those

minerals can provide additional adsorption sites for dissolved As and other elements like P (Zhang and Selim, 2008; Wang et al., 2020a). The similar stable profile of As was frequently reported in anoxic freshwater sediments (Bennett et al., 2012; Di et al., 2012; Arsic et al., 2018). By contrast, a decrease of As along depth was captured in some marine sediments rich in S (Bennett et al., 2012). Under those conditions, the immobilization of As may outcompete the mobilization and lead to As decrease in anoxic soils when S(VI) reduction predominates.

It is interesting to note that, according to the Eh-pH diagram, all the inorganic As in Eh < 0 mV (vs. standard hydrogen electrode) should be As(III) under circumneutral conditions (Akter et al., 2005). However, a considerable amount of As(V) (~ 25.0%) was observed in SG soil, and the proportion was independent of soil depth in the zone (Fig. 3). The persistent As(V) existence in reducing soils may represent a thermodynamic equilibration of four biogeochemical processes: 1) abiotic (by Mn oxides) or microbial oxidation of As(III) to As(V) (Fig. 4D) (Liu, 2006; Suda and Makino, 2016; Tong et al., 2019); 2) desorption of As(V) from the solid phase due to competition of adsorption sites by analogues (e.g. carbonate, bicarbonate, phosphate, dissolved organic matter) (Fig. 4E) (Zobrist et al., 2000; Grafe et al., 2001; Violante and Pigna, 2002; Anawar et al., 2004); 3) release of the adsorbed As(V) due to microbial reduction and solubilization of the host Fe oxides (Fig. 4F) (Tufano et al., 2008; Zhang et al., 2018); and 4) readily As(V) supply from humic acids (Fig. 4) (Anawar et al., 2003; Chaillou et al., 2003). To verify whether there is a

thermodynamic equilibration of As(III) and As(V) in soil porewater, we collected the As species data from publications using typically natural soils/sediments (Table S4). The results showed dissolved As under anoxic was usually dominated by As(III), but significant As(V) (~ 20.0%) was observed in most cases (Chaillou et al., 2003; Zheng et al., 2003; Roberts et al., 2010; Somenahally et al., 2011; Hu et al., 2015; Shakoor et al., 2015; Kumar et al., 2016; Bondu et al., 2017; Xu et al., 2017; Arsic et al., 2018; Kazi et al., 2018; Lock et al., 2018; Wang et al., 2019).

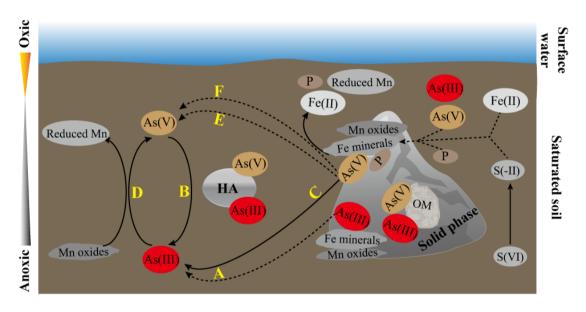


Figure 4 Diagram (nonspatial) depicting the processes controlling the behavior of multi-element in saturated soils. A-C) pathways contribute to As(III) enrichment in soil porewater; D-F) pathways contribute to As(V) enrichment in soil porewater. OM is solid organic matter, and HA is humic acids.

4. Conclusion

This study investigated vertical changes of Eh, Fe, Mn, As, P, S as well as As species across SWI in paddy soils. High-resolution (mm) mapping of total aqueous Fe, Mn,

As, P and S by SWI profiler visibly showed one-dimensional coupling and decoupling of As with Fe and Mn in different soils. Profiling of As species further identified As(III) excess was the main cause of the decoupling of As with Fe and Mn in the oxic-anoxic transition zone. Future studies, combining high-resolution mapping of multi-element with microbial community, are essential to improve the understanding of As behaviors in soils, sediments and other aquatic environments.

Conflict of Interest

The authors declare no conflict of interest.

Acknowledgements

This work was supported by the National Science Foundation of China (41571305, 41977320), Key Programme Special Fund of XJTLU (KSF-A-20). We want to thank the kind help of Fuyuan Liu for designing 3D printing model. We are grateful for the assistance of Yi-Li Cheng, Xiao Zhou, Xiao-Yan Zhang, and Liang-Ping Long with chemical analysis.

References

- 395 Ajees, A.A., Rosen, B.P., 2015. As(III) S-adenosylmethionine methyltransferases and other arsenic
- 396 binding proteins. Geomicrobiol. J. 32, 570-576.
- 397 Akter, K.F., Owens, G., Davey, D.E., Naidu, R., 2005. Arsenic speciation and toxicity in biological
- 398 systems. Reviews of environmental contamination and toxicology. Springer, pp. 97-149.
- Anawar, H.M., Akai, J., Komaki, K., Terao, H., Yoshioka, T., Ishizuka, T., Safiullah, S., Kato, K., 2003.
- 400 Geochemical occurrence of arsenic in groundwater of Bangladesh: sources and mobilization processes.
- 401 J. Geochem. Explor. 77, 109-131.
- 402 Anawar, H.M., Akai, J., Sakugawa, H., 2004. Mobilization of arsenic from subsurface sediments by
- effect of bicarbonate ions in groundwater. Chemosphere 54, 753-762.
- 404 Arsic, M., Teasdale, P.R., Welsh, D.T., Johnston, S.G., Burton, E.D., Hockmann, K., Bennett, W.W.,
- 405 2018. Diffusive gradients in thin films (DGT) reveals antimony and arsenic mobility differs in a
- 406 contaminated wetland sediment during an oxic-anoxic transition. Environ. Sci. Technol. 52, 1118-1127.
- Bennett, W.W., Teasdale, P.R., Panther, J.G., Welsh, D.T., Zhao, H., Jolley, D.F., 2012. Investigating
- 408 arsenic speciation and mobilization in sediments with DGT and DET: a mesocosm evaluation of
- oxic-anoxic transitions. Environ. Sci. Technol. 46, 3981-3989.
- Bogdan, K., Schenk, M.K., 2008. Arsenic in rice (Oryza sativa L.) related to dynamics of arsenic and
- 411 silicic acid in paddy soils. Environ. Sci. Technol. 42, 7885-7890.
- Bondu, R., Cloutier, V., Rosa, E., Benzaazoua, M., 2017. Mobility and speciation of geogenic arsenic in
- bedrock groundwater from the Canadian Shield in western Quebec, Canada. Sci. Total Environ. 574,
- 414 509-519.

- Borch, T., Kretzschmar, R., Kappler, A., Cappellen, P.V., Gindervogel, M., Voegelin, A., Campbell, K.,
- 416 2010. Biogeochemical redox processes and their impact on contaminant dynamics. Environ. Sci.
- 417 Technol. 44, 15-23.
- Bouman, B., Tuong, T.P., 2001. Field water management to save water and increase its productivity in
- irrigated lowland rice. Agr. Water Manage. 49, 11-30.
- 420 Brannon, J., Gunnison, D., Smart, R., Chen, R., 1984. Effects of added organic matter on iron and
- manganese redox systems in sediment. Geomicrobiol. J. 3, 319-341.
- Bums, R.G., Bums, V.M., 1975. Mechanism for nucleation and growth of manganese nodules. Nature
- 423 255, 130-131.
- Burton, E.D., Bush, R.T., Sullivan, L.A., Johnston, S.G., Hocking, R.K., 2008. Mobility of arsenic and
- 425 selected metals during re-flooding of iron- and organic-rich acid-sulfate soil. Chem. Geol. 253, 64-73.
- 426 Campbell, K.M., Malasam, D., Saltikov, C.W., Newman, D.K., Hering, J.G., 2006. Simultaneous
- microbial reduction of iron (III) and arsenic (V) in suspensions of hydrous ferric oxide. Environ. Sci.
- 428 Technol. 40, 5950-5955.
- Chaillou, G., Schäfer, J., Anschutz, P., Lavaux, G., Blanc, G., 2003. The behaviour of arsenic in muddy
- 430 sediments of the Bay of Biscay (France). Geochim. Cosmochim. Ac. 67, 2993-3003.
- Chen, C., Li, L., Huang, K., Zhang, J., Xie, W.Y., Lu, Y., Dong, X., Zhao, F.J., 2019. Sulfate-reducing
- 432 bacteria and methanogens are involved in arsenic methylation and demethylation in paddy soils. ISME
- 433 J. 13, 2523-2535.
- 434 Chen, Z., Kim, K.W., Zhu, Y.G., McLaren, R., Liu, F., He, J.Z., 2006. Adsorption (AsIII, V) and
- oxidation (AsIII) of arsenic by pedogenic Fe–Mn nodules. Geoderma 136, 566-572.

- Darland, J.E., Inskeep, W.P., 1997. Effects of pH and phosphate competition on the transport of
- 437 arsenate. J. Environ. Qual. 26, 1133-1139.
- 438 Das, S., Liu, C.C., Jean, J.S., Lee, C.C., Yang, H.J., 2016. Effects of microbially induced
- 439 transformations and shift in bacterial community on arsenic mobility in arsenic-rich deep aquifer
- 440 sediments. J. Hazard. Mater. 310, 11-19.
- Di, X., Wei, W., Shiming, D., Qin, S., Chaosheng, Z., 2012. A high-resolution dialysis technique for
- 442 rapid determination of dissolved reactive phosphate and ferrous iron in pore water of sediments. Sci.
- 443 Total Environ. 421-422, 245-252.
- Ding, S., Wang, Y., Wang, D., Li, Y.Y., Gong, M., Zhang, C., 2016. In situ, high-resolution evidence for
- iron-coupled mobilization of phosphorus in sediments. Sci. Rep. 6, 24341.
- Dočekalová, H., Clarisse, O., Salomon, S., Wartel, M., 2002. Use of constrained DET probe for a
- high-resolution determination of metals and anions distribution in the sediment pore water. Talanta 57,
- 448 145-155.
- 449 Frenzel, P., Rothfuss, F., Conrad, R., 1992. Oxygen profiles and methane turnover in a flooded rice
- 450 microcosm. Biol. Fert. Soils 14, 84-89.
- 451 Gallagher, P.A., Schwegel, C.A., Wei, X., Creed, J.T., 2001. Speciation and preservation of inorganic
- arsenic in drinking water sources using EDTA with IC separation and ICP-MS detection. J. Environ.
- 453 Monitor. 3, 371-376.
- Gao, Y., Leermakers, M., Gabelle, C., Divis, P., Billon, G., Ouddane, B., Fischer, J.C., Wartel, M.,
- 455 Baeyens, W., 2006. High-resolution profiles of trace metals in the pore waters of riverine sediment
- assessed by DET and DGT. Sci. Total Environ. 362, 266-277.

- 457 Gorny, J., Billon, G., Lesven, L., Dumoulin, D., Madé, B., Noiriel, C., 2015. Arsenic behavior in river
- 458 sediments under redox gradient: a review. Sci. Total Environ. 505, 423-434.
- Grafe, M., Eick, M., Grossl, P., 2001. Adsorption of arsenate (V) and arsenite (III) on goethite in the
- 460 presence and absence of dissolved organic carbon. Soil Sci. Soc. Am. J. 65, 1680-1687.
- 461 Gustave, W., Yuan, Z.F., Ren, Y.X., Sekar, R., Chen, Z., 2019a. Arsenic alleviation in rice by using
- paddy soil microbial fuel cells. Plant Soil 441, 111-127.
- Gustave, W., Yuan, Z.F., Sekar, R., Ren, Y.X., Chang, H.C., Liu, J.Y., Chen, Z., 2018. The change in
- biotic and abiotic soil components influenced by paddy soil microbial fuel cells loaded with various
- resistances. J. Soil. Sediment. 19, 106-115.
- Gustave, W., Yuan, Z.F., Sekar, R., Ren, Y.X., Liu, J.Y., Zhang, J., Chen, Z., 2019b. Soil organic matter
- 467 amount determines the behavior of iron and arsenic in paddy soil with microbial fuel cells.
- 468 Chemosphere 237, 124459.
- Honma, T., Ohba, H., Kaneko-Kadokura, A., Makino, T., Nakamura, K., Katou, H., 2016. Optimal soil
- 470 Eh, pH, and water management for simultaneously minimizing arsenic and cadmium concentrations in
- rice grains. Environ. Sci. Technol. 50, 4178-4185.
- Hu, P., Ouyang, Y., Wu, L., Shen, L., Luo, Y., Christie, P., 2015. Effects of water management on
- arsenic and cadmium speciation and accumulation in an upland rice cultivar. J. Environ. Sci. 27,
- 474 225-231.
- Islam, F.S., Gault, A.G., Boothman, C., Polya, D.A., Chamock, J.M., Chatterjee, D., Lloyd, J.R., 2004.
- 476 Role of metal-reducing bacteria in arsenic release from Bengal delta sediments. Nature 430, 68-71.
- 477 Kazi, T.G., Brahman, K.D., Baig, J.A., Afridi, H.I., 2018. A new efficient indigenous material for

- simultaneous removal of fluoride and inorganic arsenic species from groundwater. J. Hazard. Mater.
- 479 357, 159-167.
- Khan, M.A., Stroud, J.L., Zhu, Y.G., McGrath, S.P., Zhao, F.J., 2010. Arsenic bioavailability to rice is
- elevated in Bangladeshi paddy soils. Environ. Sci. Technol. 44, 8515-8521.
- 482 Kumar, M., Ramanathan, A., Rahman, M.M., Naidu, R., 2016. Concentrations of inorganic arsenic in
- 483 groundwater, agricultural soils and subsurface sediments from the middle Gangetic plain of Bihar,
- 484 India. Sci. Total Environ. 573, 1103-1114.
- Liu, F., 2006. Arsenite oxidation by three types of manganese oxides. J. Environ. Sci. 18, 292-298.
- Lock, A., Wallschläger, D., Belzile, N., Spiers, G., Gueguen, C., 2018. Rates and processes affecting As
- 487 speciation and mobility in lake sediments during aging. J. Environ. Sci. 66, 338-347.
- 488 Ma, W.W., Zhu, M.X., Yang, G.P., Li, T., 2017. In situ, high-resolution DGT measurements of
- 489 dissolved sulfide, iron and phosphorus in sediments of the East China Sea: Insights into phosphorus
- 490 mobilization and microbial iron reduction. Mar. Pollut. Bull. 124, 400-410.
- 491 Masscheleyn, P.H., Delaune, R.D., Patrick Jr, W.H., 1991. Effect of redox potential and pH on arsenic
- speciation and solubility in a contaminated soil. Environ. Sci. Technol. 25, 1414-1419.
- 493 Meharg, A.A., 2004. Arsenic in rice-understanding a new disaster for South-East Asia. Trends Plant
- 494 Sci. 9, 415-417.
- 495 Mitsunobu, S., Toda, M., Hamamura, N., Shiraishi, F., Tominaga, Y., Sakata, M., 2020.
- 496 Millimeter-scale topsoil layer blocks arsenic migration in flooded paddy soil. Geochim. Cosmochim.
- 497 Ac. 274, 211-227.
- Mucci, A., Richard, L.F., Lucotte, M., Guignard, C., 2000. The differential geochemical behavior of

- arsenic and phosphorus in the water column and sediments of the Saguenay Fjord estuary, Canada.
- 500 Aquat. Geochem. 6, 293-324.
- Myers, C.R., Nealson, K.H., 1988. Microbial reduction of manganese oxides: Interactions with iron
- and sulfur. Geochim. Cosmochim. Ac. 52, 2727-2732.
- Pester, M., Knorr, K.H., Friedrich, M.W., Wagner, M., Loy, A., 2012. Sulfate-reducing microorganisms
- in wetlands–fameless actors in carbon cycling and climate change. Front. Microbiol. 3, 72.
- 505 Polizzotto, M.L., Kocar, B.D., Benner, S.G., Sampson, M., Fendorf, S., 2008. Near-surface wetland
- sediments as a source of arsenic release to ground water in Asia. Nature 454, 505-508.
- Roberts, L.C., Hug, S.J., Voegelin, A., Dittmar, J., Kretzschmar, R., Wehrli, B., Saha, G.C.,
- 508 Badruzzaman, A.B.M., Ali, M.A., 2010. Arsenic dynamics in porewater of an intermittently irrigated
- paddy field in Bangladesh. Environ. Sci. Technol. 45, 971-976.
- 510 Shakoor, M., Niazi, N., Bibi, I., Rahman, M., Naidu, R., Dong, Z., Shahid, M., Arshad, M., 2015.
- 511 Unraveling health risk and speciation of arsenic from groundwater in rural areas of Punjab, Pakistan.
- 512 Int. J. Environ. Res. Pub. He. 12, 12371-12390.
- 513 Somenahally, A.C., Hollister, E.B., Yan, W., Gentry, T.J., Loeppert, R.H., 2011. Water management
- 514 impacts on arsenic speciation and iron-reducing bacteria in contrasting rice-rhizosphere compartments.
- 515 Environ. Sci. Technol. 45, 8328-8335.
- 516 Suda, A., Makino, T., 2016. Functional effects of manganese and iron oxides on the dynamics of trace
- elements in soils with a special focus on arsenic and cadmium: a review. Geoderma 270, 68-75.
- 518 Suzuki, Y., Shimoda, Y., Endo, Y., Hata, A., Yamanaka, K., Endo, G., 2009. Rapid and effective
- 519 speciation analysis of arsenic compounds in human urine using anion-exchange columns in

- 520 HPLC-ICP-MS. J. Occup. Health 51, 380-385.
- Takahashi, Y., Minamikawa, R., Hattori, K.H., Kurishima, K., Kihou, N., Yuita, K., 2004. Arsenic
- behavior in paddy fields during the cycle of flooded and non-flooded periods. Environ. Sci. Technol. 38,
- 523 1038-1044.
- 524 Tong, H., Liu, C., Hao, L., Swanner, E.D., Chen, M., Li, F., Xia, Y., Liu, Y., Liu, Y., 2019. Biological
- Fe(II) and As(III) oxidation immobilizes arsenic in micro-oxic environments. Geochim. Cosmochim.
- 526 Ac. 265, 96-108.
- 527 Tufano, K.J., Reyes, C., Saltikov, C.W., Fendorf, S., 2008. Reductive processes controlling arsenic
- retention: revealing the relative importance of iron and arsenic reduction. Environ. Sci. Technol. 42,
- 529 8283-8289.
- Tufano, K.J., Scott, F., 2008. Confounding impacts of iron reduction on arsenic retention. Environ. Sci.
- 531 Technol. 42, 4777-4783.
- Violante, A., Pigna, M., 2002. Competitive sorption of arsenate and phosphate on different clay
- 533 minerals and soils. Soil Sci. Soc. Am. J. 66, 1788-1796.
- Wang, H.Y., Byme, J.M., Perez, J.P.H., Thomas, A.N., Göttlicher, J., Höfer, H.E., Mayanna, S., Kontny,
- 535 A., Kappler, A., Guo, H.M., Benning, L.G., Norra, S., 2020a. Arsenic sequestration in pyrite and
- greigite in the buried peat of As-contaminated aquifers. Geochim. Cosmochim. Ac. 284, 107-119.
- Wang, J., Kerl, C.F., Hu, P., Martin, M., Mu, T., Brüggenwirth, L., Wu, G., Said-Pullicino, D., Romani,
- 538 M., Wu, L., 2020b. Thiolated arsenic species observed in rice paddy pore waters. Nature Geosci. 13,
- 539 282-287.
- 540 Wang, M., Tang, Z., Chen, X.P., Wang, X., Zhou, W.X., Tang, Z., Zhang, J., Zhao, F.J., 2019. Water

- 541 management impacts the soil microbial communities and total arsenic and methylated arsenicals in rice
- 542 grains. Environ. Pollut. 247, 736-744.
- Weber, F.A., Hofacker, A.F., Voegelin, A., Kretzschmar, R., 2010. Temperature dependence and
- 544 coupling of iron and arsenic reduction and release during flooding of a contaminated soil. Environ. Sci.
- 545 Technol. 44, 116-122.
- White, B.P., Mulligan, S., Merrill, K., Wright, J., 2007. An examination of the relationships between
- motor and process skills and scores on thes sensory profile. Appl. Environ. Microb. 61, 154-160.
- Widerlund, A., Davison, W., 2007. Size and density distribution of sulfide-producing microniches in
- lake sediments. Environ. Sci. Technol. 41, 8044-8049.
- Williams, P.N., Zhang, H., Davison, W., Meharg, A.A., Hossain, M., Norton, G.J., Brammer, H., Islam,
- 551 M.R., 2011. Organic matter-solid phase interactions are critical for predicting arsenic release and plant
- uptake in Bangladesh paddy soils. Environ. Sci. Technol. 45, 6080-6087.
- Wu, Z., Jiao, L., Wang, S., Xu, Y., 2016a. Multi-metals measured at sediment-water interface (SWI) by
- diffusive gradients in thin films (DGT) technique for geochemical research. Arch. Environ. Con. Tox.
- 555 70,429-437.
- Wu, Z., Ren, D., Zhou, H., Hang, G., Li, J., 2016b. Sulfate reduction and formation of iron sulfide
- minerals in nearshore sediments from Qi'ao Island, Pearl River Estuary, Southern China. Quatem. Int.
- 558 452, 137-147.
- 559 Xiu, W., Guo, H., Shen, J., Liu, S., Ding, S., Hou, W., Ma, J., Dong, H., 2016. Stimulation of Fe (II)
- oxidation, biogenic lepidocrocite formation, and arsenic immobilization by Pseudogulbenkiania sp.
- strain 2002. Environ. Sci. Technol. 50, 6449-6458.

- Xu, W., Wang, H., Liu, R., Zhao, X., Qu, J., 2011. Arsenic release from arsenic-bearing Fe-Mn binary
- oxide: Effects of Eh condition. Chemosphere 83, 1020-1027.
- Xu, X., Chen, C., Wang, P., Kretzschmar, R., Zhao, F.J., 2017. Control of arsenic mobilization in paddy
- soils by manganese and iron oxides. Environ. Pollut. 231, 37-47.
- 566 Yuan, Z.-F., Gustave, W., Sekar, R., Bridge, J., Wang, J.-Y., Feng, W.-J., Guo, B., Chen, Z., 2021.
- 567 Simultaneous measurement of aqueous redox-sensitive elements and their species across the soil-water
- 568 interface. J. Environ. Sci. 102, 1-10.
- Yuan, Z.F., Ata-Ul-Karim, S.T., Cao, Q., Lu, Z., Cao, W., Zhu, Y., Liu, X., 2016. Indicators for
- diagnosing nitrogen status of rice based on chlorophyll meter readings. Field Crops Res. 185, 12-20.
- 571 Yuan, Z.F., Gustave, W., Bridge, J., Liang, Y., Sekar, R., Boyle, J., Jin, C.Y., Pu, T.Y., Ren, Y.X., Chen,
- 572 Z., 2019. Tracing the dynamic changes of element profiles by novel soil porewater samplers with
- ultralow disturbance to soil—water interface. Environ. Sci. Technol. 53, 5124-5132.
- Zachara, J.M., Kukkadapu, R.K., Fredrickson, J.K., Gorby, Y.A., Smith, S.C., 2002. Biomineralization
- of poorly crystalline Fe(III) oxides by dissimilatory metal reducing bacteria (DMRB). Geomicrobiol. J.
- 576 19,179-207.
- 577 Zhang, H., Selim, H.M., 2008. Competitive sorption-desorption kinetics of arsenate and phosphate in
- 578 soils. Soil Sci. 173, 3-12.
- 579 Zhang, J., Ma, T., Yan, Y., Xie, X., Abass, O.K., Liu, C., Zhao, Z., Wang, Z., 2018. Effects of Fe-S-As
- coupled redox processes on arsenic mobilization in shallow aquifers of Datong Basin, northern China.
- 581 Environ. Pollut. 237, 28-38.
- Zhao, Y., Su, J., Ye, J., Rensing, C., Tardif, S., Zhu, Y., Brandt, K.K., 2019. AsChip: a high-throughput

- 583 qPCR chip for comprehensive profiling of genes linked to microbial cycling of arsenic. Environ. Sci.
- 584 Technol. 53, 798-807.
- Zheng, J., Hintelmann, H., Dimock, B., Dzurko, M.S., 2003. Speciation of arsenic in water, sediment,
- and plants of the Moira watershed, Canada, using HPLC coupled to high resolution ICP-MS. Anal.
- 587 Bioanal. Chem. 377, 14-24.
- Zobrist, J., Dowdle, P.R., Davis, J.A., Oremland, R.S., 2000. Mobilization of arsenite by dissimilatory
- reduction of adsorbed arsenate. Environ. Sci. Technol. 34, 4747-4753.