

Development of a method for the simultaneous detection of Cr(iii) and Cr(vi) in exhaled breath condensate samples using μ LC-ICP-MS

LEESE, Elizabeth and MORTON, Jackie and GARDINER, Philip
<<http://orcid.org/0000-0002-2687-0106>> and CAROLAN, Vikki
<<http://orcid.org/0000-0001-7384-4018>>

Available from Sheffield Hallam University Research Archive (SHURA) at:

<https://shura.shu.ac.uk/12281/>

This document is the author deposited version. You are advised to consult the publisher's version if you wish to cite from it.

Published version

LEESE, Elizabeth and MORTON, Jackie and GARDINER, Philip and CAROLAN, Vikki (2016). Development of a method for the simultaneous detection of Cr(iii) and Cr(vi) in exhaled breath condensate samples using μ LC-ICP-MS. *Journal of analytical atomic spectrometry*, 31 (4), 924-933.

Copyright and re-use policy

See <http://shura.shu.ac.uk/information.html>

CrossMark
click for updatesCite this: *J. Anal. At. Spectrom.*, 2016, **31**, 924

Development of a method for the simultaneous detection of Cr(III) and Cr(VI) in exhaled breath condensate samples using μ LC-ICP-MS

Elizabeth Leese,^{*ab} Jackie Morton,^a Philip H. E. Gardiner^b and Vikki A. Carolan^b

The method described in this study is a robust and sensitive method for the simultaneous determination of trivalent and hexavalent chromium (Cr(III) and Cr(VI)) in an exhaled breath condensate (EBC) sample and drinking water proficiency testing material sample. The method uses a hyphenated micro liquid chromatography (μ LC) system coupled to inductively coupled plasma mass spectrometry (ICP-MS). The optimised method incorporates a pH adjusted EDTA complexation step to stabilise Cr(VI) and Cr(III). Separation was achieved using an anion exchange micro-sized column. The limits of quantification were 0.040 $\mu\text{g L}^{-1}$ and 0.013 $\mu\text{g L}^{-1}$ for Cr(III) and Cr(VI) respectively. EBC is the non-invasive collection of condensate from cooled exhaled breath during regular tidal breathing. We propose that EBC may be a suitable alternative biological matrix to give a more comprehensive picture of exposure to Cr(VI) following an inhalation exposure. This study shows that water samples and EBC samples containing Cr(III) and Cr(VI) can be stored in the fridge ($\sim 4^\circ\text{C}$) for up to 6 weeks when they have been diluted 10 fold with an EDTA solution, and that EBC samples must not be frozen after collection if the integrity of Cr(VI) is to be maintained. This study has demonstrated a practical approach to collecting EBC samples from workers potentially occupationally exposed to hexavalent chromium and that the integrity of the chromium species will be maintained prior to analysis.

Received 30th October 2015
Accepted 3rd February 2016

DOI: 10.1039/c5ja00436e

www.rsc.org/jaas

Introduction

Trivalent chromium is biologically important as an essential element for humans, needed for glucose and lipid metabolism.¹ Hexavalent chromium however is considered toxic and IARC has classified it as a group 1 agent, defined as carcinogenic to humans.² Hexavalent chromium is known to cause lung and nasal cancer as well as other health conditions such as skin allergies, skin ulcerations, nasal perforations, respiratory problems, kidney and liver damage.³ Trivalent chromium [+3, Cr(III)] is the most common form of chromium, whilst hexavalent chromium [+6, Cr(VI)] is rarely found naturally in the environment, therefore the majority of hexavalent chromium is a product of industry and manufacturing.⁴

Chromium compounds are occupationally important in many industries such as the production and welding of stainless steel including cutting and finishing, electroplating, smelting, pigments and paints used in aerospace and maritime industries, construction and those using pesticides and tanning agents.⁴ Exposure to Cr(VI) and its compounds can occur by ingestion or dermal exposure but in some industries the

inhalation of fumes and mists is most likely. Upon exposure to Cr(VI), the body naturally reduces Cr(VI) to Cr(III)^{5,6} before it is excreted in the urine, with studies suggesting occupational exposure to chromium mainly *via* inhalation results in elimination half-lives of 7 hours and 15–30 days.⁷ Apart from in bacterial cells, there is no evidence of humans or other biological systems having the ability to re-oxidise Cr(III) back to Cr(VI).⁸ One approach to assessing occupational exposure uses biological monitoring, involving the measurement of total chromium (without differentiation of Cr(III) and Cr(VI)) in a urine sample. Background unexposed reference ranges and occupational guidance values for total chromium in urine have been established in the UK,⁹ Germany¹⁰ and the US.¹¹ Cr(VI) has the ability to permeate cell membranes meaning Cr(VI) can also be measured in red blood cells (RBC),^{12,13} determined as Cr(III). Once inside RBCs Cr(VI) is rapidly reduced to Cr(III) as it binds with haemoglobin and other proteins.^{6,12} Methods for the separate measurements of Cr(III) and Cr(VI) have focused on the assessment of environmental sources of exposure such as natural water,^{14,15} drinking water,^{14,15} waste water¹⁶ and industrial waste.¹⁷

Whilst the measurement of total chromium in urine is a practical bio-monitoring approach, it does not give a comprehensive picture of exposure. It is possible that an alternative biological matrix to measure exposure to Cr(VI) for those exposed through inhalation might be exhaled breath

^aHealth and Safety Executive, Harpur Hill, Buxton, Derbyshire, SK17 9JN, UK. E-mail: Liz.leese@hsl.gsi.gov.uk; Tel: +44 (0)1298218434

^bBiomolecular Sciences Research Centre, Sheffield Hallam University, Howard Street, Sheffield S1 1WB, UK



condensate (EBC). EBC is the non-invasive collection of condensate from cooled exhaled breath during regular tidal breathing. The collection of exhaled air will contain mostly water vapour but also droplets of fluid from the respiratory tract,¹⁸ meaning EBC measurements will reflect markers and molecules from the mouth, tracheobronchial system and the alveoli.¹⁹ EBC is reported to contain both volatile substances in the gaseous phase along with low volatile and non-volatile substances in droplets.²⁰ The non-volatile substances will include salts, proteins, lipids, and environmental contaminants.^{18–20} Because EBC is relatively free of the interfering solutes found in other biological matrices it represents a suitable biological matrix for elemental analysis to assess inhalation exposure to trace elements. Studies have already shown that occupationally inhaled elements have been detected in EBC for cobalt and tungsten,^{21,22} iron, nickel and total chromium,^{23,24} manganese²⁵ and hexavalent chromium (using liquid/liquid solvent extraction and analysis by electrothermal atomic absorption spectrometry).²⁶ There are currently no publications for the simultaneous detection and determination of Cr(III) and Cr(VI) in an EBC sample. Due to the difference in toxicities of Cr(III) and Cr(VI) and the conversion from Cr(VI) to Cr(III) in the human body after exposure, it is important to measure both species of chromium to enable a more comprehensive picture of exposure assessment.

In order to measure Cr(III) and Cr(VI) it is necessary to maintain the integrity of each species in the sample. The stability of both trivalent and hexavalent chromium depends on pH. In acidic solutions, trivalent chromium compounds mainly exist as $\text{Cr}(\text{H}_2\text{O})_6^{3+}$ and hydrolyse to a hydroxide in less acidic and alkaline conditions. Hexavalent chromium compounds exist either as a chromate (CrO_4^{2-} , HCrO_4^- , H_2CrO_4) or dichromate ($\text{Cr}_2\text{O}_7^{2-}$).^{3,27} In extremely acidic conditions only, hexavalent chromium exists as H_2CrO_4 ; in these acidic conditions hexavalent chromium is a potent oxidant, which after oxidising can also begin to reduce to trivalent chromium compounds. In the pH range of 2–6 hexavalent chromium compounds exist as HCrO_4^- and HCr_2O_7^- , with HCr_2O_7^- reducing to $\text{Cr}_2\text{O}_7^{2-}$ at concentrations higher than 10^{-2} M. Above pH 6, hexavalent chromium only exists as CrO_4^{2-} .^{3,8,27,28} Due to this pH dependence and the fact that trivalent chromium compounds are typically cationic in solution and hexavalent chromium compounds are typically anionic in solution, speciation using ion exchange chromatography methodologies is an analytical challenge. These challenges have resulted in analysis either only determining Cr(VI) using colorimetric analysis with diphenylcarbohydrazide (DPC)¹⁵ or by determining both Cr(VI) by anion chromatography and total chromium to calculate Cr(III) by subtraction.¹⁴ Cr(III) and Cr(VI) have been simultaneously determined by ICP-MS using reverse phase chromatography combined with ion-pairing reagents.^{27,29} The use of non-traditional HPLC chromatographic methods such as solid phase extraction based fractionation,^{14,30,31} micro columns (PTFE tubes filled with activated alumina)^{32–35} and short column capillary electrophoresis³⁶ with subsequent detection by AAS, ICP-AES or ICP-MS have all been employed as simple techniques to separate and determine Cr(III) and Cr(VI). Whilst all

these methods have their own advantages and disadvantages (such as interferences, contamination, raised backgrounds, unsuitable limits of detection and time consuming procedures), it has meant that other research has focused on anion exchange chromatography coupled to ICP-MS. Both Chen *et al.*³⁷ and Pansar-Kallio and Manninen³⁸ employed a dual column approach to overcome the issue of the opposite charges of the chromium species. Other authors overcame this ionic challenge by using derivatisation, where Cr(III) is complexed with EDTA to change the ionic charge from a cation to an anion complex, enabling separation of the two chromium species using the same sample injection on an anion exchange column.^{39,40} Suzuki and Serita⁴¹ were the first to report this technique using HPLC-AAS, with Chen *et al.*³⁷ reporting that alternative complexing agents such as diethylenetriaminepentaacetic acid (DTPA) and 2,6-pyridinedicarboxylic acid (PDCA) were not as effective as EDTA. Other publications have reported the successful speciation of Cr(III) and Cr(VI) using newly introduced speciality anion columns with sulfonic and alkyl quaternary ammonium functional groups which boast both anionic and cationic properties, and have been employed by authors concerned with species' integrity related to sample pre-treatment.^{42–44}

The initial aim of this study was to develop a speciation method to simultaneously determine both Cr(III) and Cr(VI) using a small anion exchange speciality column with cationic capabilities using a OneFAST system coupled to an ICP-MS. The method developed was then used to investigate the stability of both chromium species in an EBC sample with the view to establishing a viable technique to employ for workplace monitoring. The objective, being to establish a method to allow the collection and analysis of EBC samples from workers who may be occupationally exposed to hexavalent chromium.

Experimental

Reagents

Chromium speciation compounds potassium dichromate ($\text{K}_2\text{Cr}_2\text{O}_7$) and chromium(III) chloride (CrCl_3) were obtained from Fisher Scientific (Loughborough, UK). Ammonia solution (NH_3), UpA nitric acid and UpA hydrochloric acid was supplied by Romil (UK). Butan-1-ol was from Fisher Scientific (Loughborough, UK).

All solutions were made using ultrapure deionised water (18.2 M Ω cm) from a Millipore system (Merck Millipore, Billerica, MA, USA). Single standard stock solutions of 1000 mg L⁻¹ of Cr(III) and Cr(VI) were prepared fresh in water for each day of analysis.

A commercially available proficiency testing material (PTM) from Sigma Aldrich (UK) for Cr(VI) in drinking water (Sample WS13-4 Lot LRAA1427) was used to validate the method. Manufacturer instructions suggest the PTM is diluted 100 fold in water. However for this study the PTM was diluted 1000 fold in water to more accurately reflect the levels of Cr(VI) expected in EBC samples. Previous studies that measured total chromium in EBC²⁴ or Cr(VI) only in EBC⁴⁵ reported levels less than 10 $\mu\text{g L}^{-1}$ and 5 $\mu\text{g L}^{-1}$ of chromium in EBC respectively.



Instrumentation

Separation was achieved using a hyphenated μ LC system with an ICP-MS (XSERIES 2, Thermo Fisher Scientific, Hemel Hempstead, UK). The separation of Cr(III) and Cr(VI) was achieved using a 5 cm anion exchange guard column, (Dionex IONPAC AG7 4 mm \times 50 mm i.d., 10 μ m, Thermo Fisher Scientific). The Dionex AG7 anion exchange column has cationic exchange capability due to the presence of sulfonic and alkyl quaternary ammonium functional groups, and is suitable for a full pH range of 0–14. The micro-flow delivery of both sample and mobile phase was accomplished using an ESI OneFAST system (Elemental Scientific, Warrington, UK), with a six port switching valve and 500 μ L sample loop. The ICP-MS was operated in collision cell (CCT) mode using 7% hydrogen in helium (approximately 3.5 mL min⁻¹). The ICP-MS conditions were optimised daily using a 10 μ g L⁻¹ tuning solution containing arsenic, chromium, cobalt, indium and uranium (made in 1% (v/v) nitric acid from 1000 mg L⁻¹ stock standards (ICP-MS standards, BDH, Poole, UK)). Table 1 summarises the ICP-MS and OneFAST operating conditions.

Exhaled breath condensate collection

EBC was collected using a portable TURBO-DECCS condenser (Transportable Unit for Research on Biomarkers Obtained from Disposable Exhaled Condensate Collection Systems) from Ital-Chill (Parma, Italy). This technique consists of a disposable respiratory system comprising of a mouthpiece connected to a one-way aspiration valve and saliva trap with an EBC sample collection tube at the end inserted into a temperature controlled chilling unit set at -5 $^{\circ}$ C (see Fig. 1). The collection of exhaled



Fig. 1 TURBO-DECCS condenser consisting of a disposable respiratory system comprising of a mouthpiece connected to a one-way aspiration valve and saliva trap with an EBC sample collection tube at the end inserted into a temperature controlled chilling unit set at -5 $^{\circ}$ C.

air results in low sample volumes of EBC; 15 minutes of regular tidal breathing will typically provide less than 2 mL of EBC sample. It was necessary to collect large volumes of EBC from a volunteer not occupationally exposed to hexavalent chromium for use in method development as a blank sample and for recovery. In addition to the volunteer supplying the bulk EBC, five further volunteers supplied EBC samples for the optimised method applications. Blank EBC was collected from the volunteer by regular tidal breathing through the mouth piece of the TURBO-DECCS. For the most part a complete seal around the mouthpiece with the mouth is maintained allowing periodic swallowing of any accumulated saliva.

Method development

See Fig. 2 for a flow chart outlining the six stages of the method development.

Sample storage experiment

1. Standards made in water. Individual chromium species standards of Cr(III) and Cr(VI), along with a mixed species standard, at a concentration of 5 μ g L⁻¹ were prepared in water. Four aliquots of each of the three standards were pipetted into individual sample tubes. The first set of aliquots was analysed by μ LC-ICP-MS immediately after preparation; the remaining sets were capped and one of each standard was stored in the freezer (~ -20 $^{\circ}$ C), the fridge (~ 4 $^{\circ}$ C) and at laboratory room temperature (~ 22 $^{\circ}$ C) for 24 hours. After 24 hours storage each aliquot was removed from their respective storage conditions, placed on a roller mixer and left to reach room temperature. After reaching room temperature each aliquot was analysed by μ LC-ICP-MS.

2. Derivatisation by complexation of Cr(III) with EDTA. A solution of 0.5 mM EDTA was made in water and used to prepare a 5 μ g L⁻¹ Cr(III) standard. The standard was analysed immediately and again in 1 hour after being left to stand at room temperature.

Table 1 Operating conditions of the ICP-MS and OneFAST system

XSERIES2 ICP-MS Settings	Parameters
RF power	1400 W
Nebuliser gas flow	0.89–0.90 L min ⁻¹
Sample/skimmer cones	Ni/Ni Xt cones
Collision gas 7% He ⁺ H ₂ flow rate	3.5 mL min ⁻¹
Isotope monitored	⁵² Cr
Dwell time	100 ms
μ LC conditions	Parameters
Analytical column	Dionex IONPAC AG7 (4 mm \times 50 mm, 10 μ m)
Mobile phase	4% v/v ammonia solution & 3.2% v/v nitric acid, pH 1.8–2
Sample injection time	30 s
Flow rate	1 mL min ⁻¹
Total acquisition time	230 s
Sample loop	500 μ L
Peristaltic pump tubing	Green/orange 0.38 mm
ESI switching valve	P6 – 6 port
Valve to column connector	0.25 ID (green)
Column to ICP-MS nebuliser	0.5 ID (orange)
Nebuliser	PFA-ST



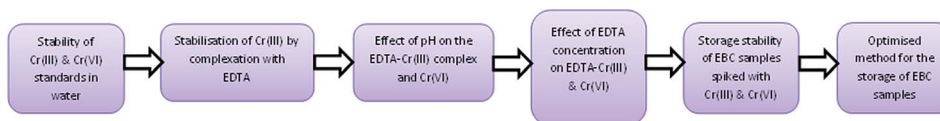


Fig. 2 Flow chart showing optimisation stages of the method development.

3. Effect of pH on the EDTA–Cr(III) complex and Cr(VI). In a preparation of 0.5 mM EDTA in water, the pH is 5. Six solutions of 0.5 mM EDTA were prepared; five of those solutions were adjusted to achieve six different pH readings; pH 4, 5, 7, 7.6, 8 and 9. 10% v/v nitric acid was used to adjust to pH 4 and 10% v/v ammonia solution was used to adjust to pH 7, 7.6, 8 and 9.

4. Concentration of EDTA. To investigate the influence of the concentration of the EDTA solution on either retention time of the chromium species or peak shape a $5 \mu\text{g L}^{-1}$ mixed species standard was prepared in 0.5 mM, 0.6 mM and 0.7 mM EDTA solutions. Each solution was adjusted to pH 8 using 10% v/v ammonia solution, and left to stand at room temperature for 1 hour prior to analysis.

5. Storage stability of EBC samples. Two solutions of 0.5 mM EDTA were prepared and adjusted to pH 7.6 and pH 8 using 10% v/v ammonia solution. Two equal volumes of “blank” EBC were spiked with $5 \mu\text{g L}^{-1}$ Cr(III) and $5 \mu\text{g L}^{-1}$ Cr(VI). Each of

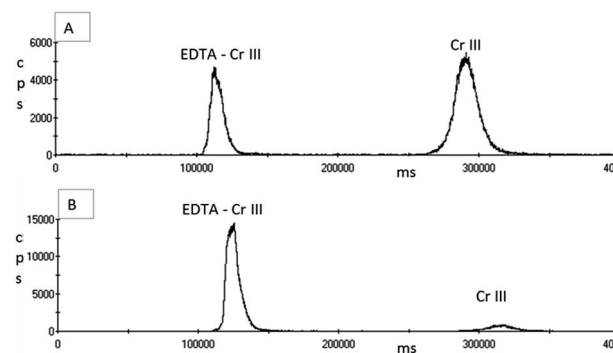


Fig. 4 Chromatograms of a $5 \mu\text{g L}^{-1}$ Cr(III) standard prepared in 0.5 mM EDTA analysed using an ESI OneFAST system coupled to a Dionex AG7 anion exchange column and ICP-MS using a nitric acid and ammonia mixed mobile phase. (A) Analysed immediately after preparation. (B) Analysed after being left to stand for 1 hour.

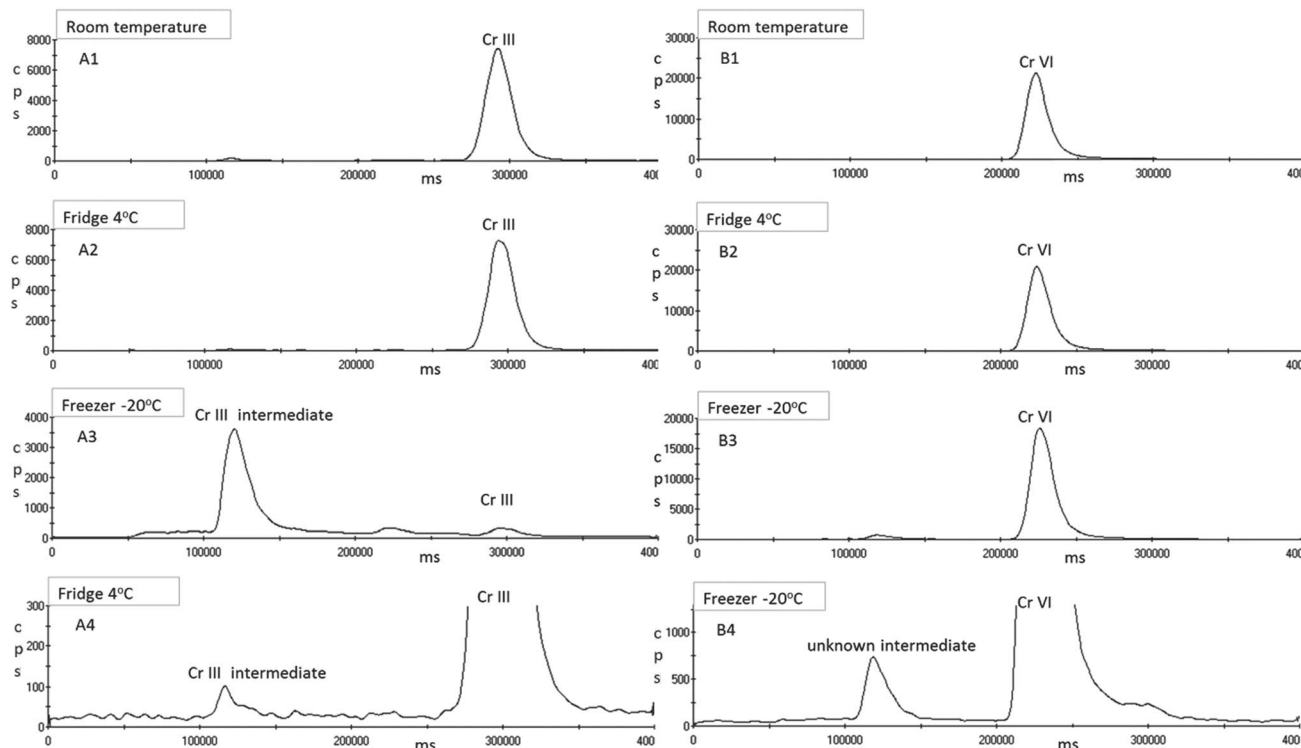


Fig. 3 Chromatograms of $5 \mu\text{g L}^{-1}$ single standards of Cr(VI) and Cr(III) analysed using an ESI OneFAST system coupled to a Dionex AG7 anion exchange column and ICP-MS using a nitric acid and ammonia mixed mobile phase. (A1–A4) Chromatograms of Cr(III). (B1–B4) Chromatograms of Cr(VI). (A1 & B1) Chromium species stored at laboratory room temperature ($\sim 22^\circ\text{C}$). (A2 & B2) Chromium species stored at fridge temperature (4°C). (A3 & B3) Chromium species stored at freezer temperature (-20°C) for 24 hour. (A4) Baseline of chromatogram (A2) showing Cr(III) intermediate compound. (B4) Baseline of chromatogram (B3) showing the unknown chromium intermediate compound.



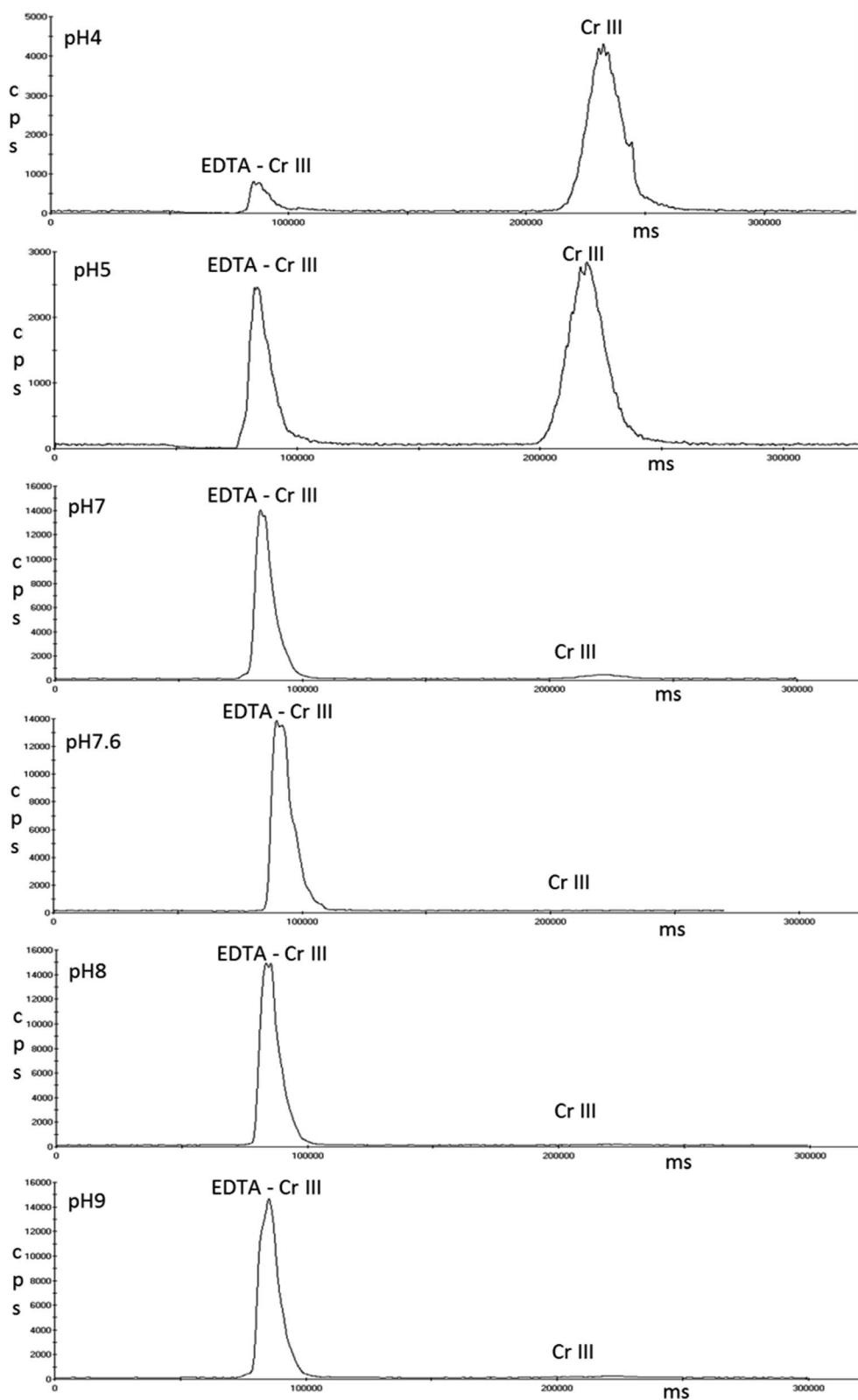


Fig. 5 Chromatograms showing the effect of pH of the EDTA solution to successfully complex and stabilise Cr(III), analysed using an ESI OneFAST system coupled to a Dionex AG7 anion exchange column and ICP-MS using a nitric acid and ammonia mixed mobile phase.



the two spiked concentrations of EBC and a “blank” unspiked EBC sample were aliquoted in triplicate for four sets of samples, and then each diluted 10 fold with the pH 7.6 solution of 0.5 mM EDTA. Four further sets of these three samples were prepared and diluted 10 fold with the pH 8 solution of 0.5 mM EDTA. All samples were capped and left to stand at room temperature for 1 hour. One of the four sets of samples of each pH were analysed by μ LC-ICP-MS. The remaining sets were stored in the fridge ($\sim 4^\circ\text{C}$), and analysed at 1 week, 4 weeks and 6 weeks.

6. Optimised method. Separation of Cr(III) and Cr(VI) was achieved using a Dionex AG7 column and an isocratic flow of ammonia solution and nitric acid mobile phase. The mobile phase was prepared daily in ultrapure deionised water, by adding 4% v/v ammonia solution and 3.2% v/v ultrapure nitric acid. The pH was adjusted to between pH 1.8–2 using the ammonia solution. The flow rate of both the mobile phase and sample injection onto the column is controlled by the ICP-MS peristaltic pump at a constant 1 mL min^{-1} . The sample is injected onto the column for 30 seconds and overall sample acquisition time is 230 seconds.

The optimised method incorporates a derivatisation step, with a 10 fold dilution of the EDTA solution (at a concentration of 0.5 mM adjusted to pH 8 with 10% v/v ammonia solution)

with the EBC sample. From the 1000 mg L^{-1} single standard stock solutions of Cr(III) and Cr(VI) $100\text{ }\mu\text{g L}^{-1}$ single and mixed species working solutions were prepared daily. Using the $100\text{ }\mu\text{g L}^{-1}$ mixed species working solution, calibration standards of 0.01, 0.02, 0.05, 0.1, 0.5, 1, 2 and $5\text{ }\mu\text{g L}^{-1}$ were prepared in the 0.5 mM EDTA solution using a 10 fold dilution to 50 mL. Blank and spiked EBC samples were diluted 10 fold with the EDTA solution to 1.5 mL. The original 1 in 1000 preparation of the proficiency testing material to validate the method was subsequently diluted 10 fold to 50 mL with the EDTA solution. All samples including standards were left to stand at room temperature for a minimum of 1 hour prior to analysis by μ LC-ICP-MS.

Results & discussion

Method development

Following initial work with the AG7 ion exchange column, the simultaneous separation of Cr(III) and Cr(VI) was achieved in spiked aqueous samples using an isocratic flow of 4% v/v ammonia solution and 3.2% v/v ultrapure nitric acid, with pH adjustment to between pH 1.8–2 using the ammonia solution.

1. Standards made in water. A spiked aqueous standard was stored at various temperatures for 24 hours prior to analysis

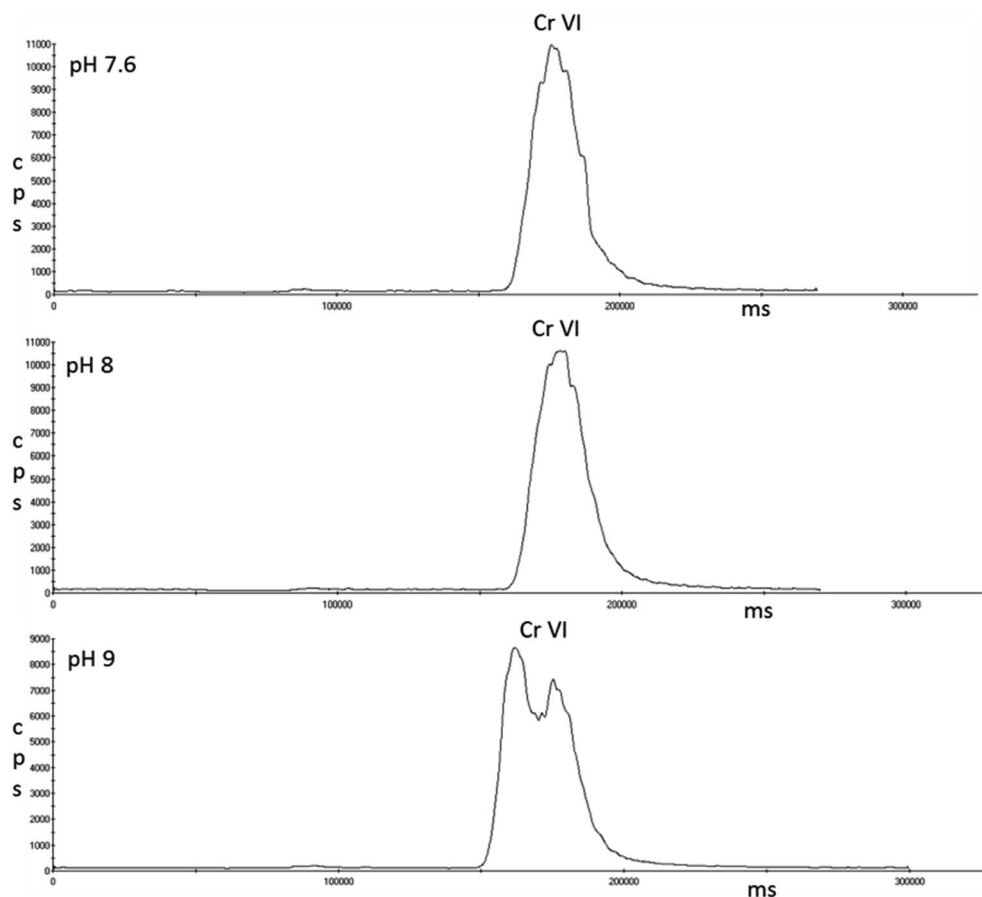


Fig. 6 Chromatograms showing the effect of pH of the EDTA solution on Cr(VI) analysed using an ESI OneFAST system coupled to a Dionex AG7 anion exchange column and ICP-MS using a nitric acid and ammonia mixed mobile phase.



to determine stability. The results (see Fig. 3) show that Cr(vi) was stable when stored at both room temperature and refrigerated, but species instability and conversion of Cr(vi) to an unknown intermediate chromium species (chromatograms B3 and B4 of Fig. 3) with the same retention time as the Cr(III) intermediate was observed when stored frozen at $-20\text{ }^{\circ}\text{C}$. Instability of Cr(III) was observed in all storage conditions (see Fig. 3). The pH of water standards is neutral, so it is possible that the unknown Cr(III) intermediate observed in chromatograms (A3) and (A4) of Fig. 3 is a result of the incomplete hydrolysis or dissociation of the trivalent chromium compound $\text{Cr}(\text{H}_2\text{O})_6^{3+}$ to $\text{Cr}(\text{OH})_2^+$, $\text{Cr}(\text{OH})_2^+$ or $\text{Cr}(\text{OH})_3$. Publications concerned with the determination of trace elements in EBC, specify that samples of EBC were frozen at $-80\text{ }^{\circ}\text{C}$ after collection.^{20,21,24,25,46} Initial observations of freezing a spiked aqueous standard of chromium Cr(III) and Cr(vi) at $-80\text{ }^{\circ}\text{C}$ showed the same pattern of conversion to intermediate species as the aqueous standard stored at $-20\text{ }^{\circ}\text{C}$ for 24 hours. If Cr(vi) cannot be successfully stored frozen, chromium speciation samples would need to be stored under refrigeration. Cr(III) cannot be successfully stored at any temperature so the complexation with EDTA is needed for stability.

2. Derivatisation by complexation of Cr(III) with EDTA. The first thing to note is that by complexing Cr(III) with EDTA the elution order of the two chromium species changes. EDTA does not form a complex with Cr(vi). Without complexation, Cr(III) elutes after Cr(vi) with a retention time of approximately 4.5 minutes. Cr(III) in solution exists in the form of a cationic hydroxy complex and is retained on the sulfonated region of the column whilst the Cr(vi) ions are retained on the alkyl quaternary ammonium region. However after complexation the

EDTA–Cr(III) complex elutes first with a retention time of just over 1 minute in the final optimised method, indicating the anionic EDTA–Cr(III) ions are no longer retained by the column.

The chromatograms in Fig. 4 show that the complexation reaction of Cr(III) with the EDTA solution was not instantaneous but achieved near full complexation after one hour at room temperature prior to analysis. In many of the studies published where EDTA complexation was used to stabilise or change the charge of Cr(III), concentrations lower than 0.5 mM EDTA were used and heat was also used for the derivatisation reaction to occur. Both Suzuki & Serita⁴¹ and Byrdy *et al.*³⁹ heated the samples in a heat block for 1 hour at $50\text{ }^{\circ}\text{C}$; Gurleyuk & Wall-schlager⁴⁰ heated their samples for 1 hour at $70\text{ }^{\circ}\text{C}$ and Leist *et al.*⁴⁷ heated for 1.5 hours at $70\text{ }^{\circ}\text{C}$. This approach would not be practical after the collection of EBC samples from workers at a place of work, and samples may not be delivered to the laboratory for 24 hours, meaning immediate heating of samples after collection in a heat block is not a viable option. However, Wolf *et al.*²⁷ found that the complexation reaction was fully achieved with 0.5 mM of EDTA when the samples were left to stand at room temperature for 30 minutes prior to analysis; having adjusted the pH of the EDTA solution to pH 7.6. The pH of the EDTA solution used in this study without any adjustment was pH 5, which may explain why the complexation reaction shown in chromatogram B of Fig. 4 was not 100% effective.

3. Effect of pH on the EDTA–Cr(III) complex and Cr(vi). Having undertaken the experiment outlined in Section 2 of the method development – Derivatisation by complexation of Cr(III) with EDTA, it was necessary to determine the effect of pH on the ability of EDTA to successfully complex Cr(III). It can be seen in Fig. 5 that EDTA at pH 4, 5 and 7 did not fully complex and

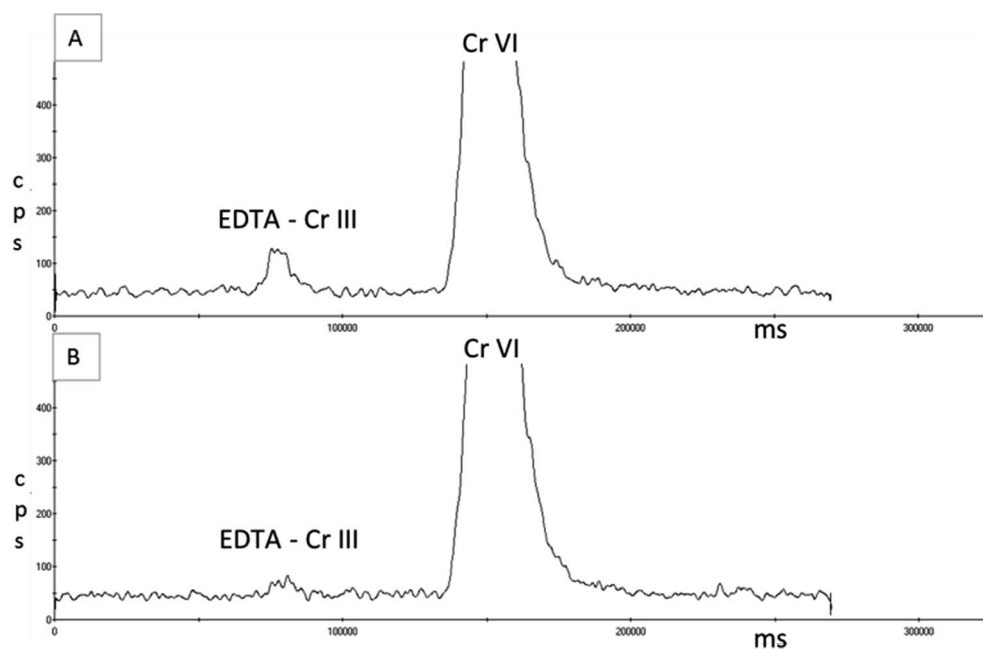


Fig. 7 Chromatograms of two Cr(vi) spiked EBC samples, stored at fridge temperature ($\sim 4\text{ }^{\circ}\text{C}$) for 6 weeks prior to analysis using an ESI OneFAST system coupled to a Dionex AG7 anion exchange column and ICP-MS using a nitric acid and ammonia mixed mobile phase. (A) EDTA pH 7.6. (B) EDTA pH 8.



stabilise Cr(III), meaning the elution of Cr(III) resulted in two peaks, whereas EDTA at pH 7.6, 8 and 9 show full complexation of Cr(III). The retention time of the EDTA–Cr(III) complex is the same throughout the pH range, however pH does affect the efficiency of complexation. The pH of the EDTA solution in the study reported by Wolf *et al.*²⁷ is within a range of pH 7.2–7.6.

Single standards of $5 \mu\text{g L}^{-1}$ Cr(VI) were prepared in the three pH solutions of EDTA that achieved full complexation of Cr(III); pH 7.6, 8 and 9. Fig. 6 shows that pH 7.6 and 8 had no effect on Cr(VI); however peak splitting was observed at pH 9.

4. Concentration of EDTA. As pH is an important factor in the complexation of EDTA with Cr(III), it was necessary to determine if the increased concentration of EDTA was also an important factor. As both pH 7.6 and pH 8 have both shown to

be a suitable pH for the EDTA solution, pH 8 was selected for the pH of the three different concentrations of EDTA solutions. The varying concentrations of 0.5, 0.6 and 0.7 mM EDTA solution at pH 8 made no difference to the retention, sensitivity or peak shape of either chromium species (data not shown).

5. Storage stability of EBC samples. Having developed the chromium speciation method, storage stability studies of Cr(III) and Cr(VI) in EBC samples were undertaken. Analysis of EBC spiked with single standards of Cr(III) and Cr(VI) at a concentration of $5 \mu\text{g L}^{-1}$ diluted 1 in 10 with a 0.5 mM EDTA solution of either pH 7.6 or pH 8 showed that the EBC EDTA–Cr(III) complex was stable with no sign of deterioration or species conversion for the entire 6 week period at both pH values. The Cr(VI) sample was also stable for the 6 week storage period when the Cr(VI)

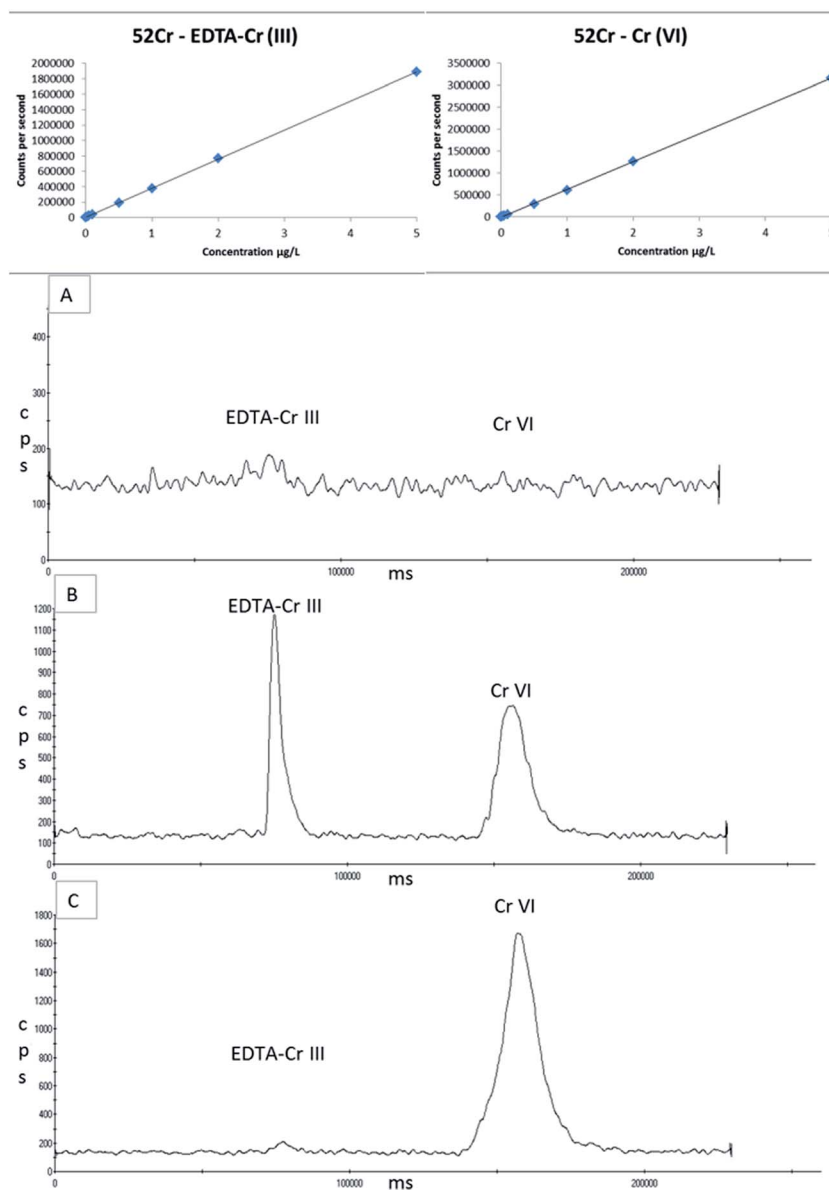


Fig. 8 Linear calibration curves of 0–5 $\mu\text{g L}^{-1}$ for both EDTA–Cr(III) and Cr(VI) using an ESI OneFAST system coupled to a Dionex AG7 anion exchange column and ICP-MS using a nitric acid and ammonia mixed mobile phase. (A) Blank EBC sample. (B) A 2 $\mu\text{g L}^{-1}$ spiked blank EBC sample with EDTA–Cr(III) and Cr(VI). (C) Commercially available proficiency testing material for Cr(VI) in drinking water.



spiked EBC samples were diluted with the pH 8 solution of EDTA. However the Cr(VI) spiked EBC samples that had been diluted with the pH 7.6 EDTA solution showed signs of instability after only 1 week of storage. At pH 7.6 the Cr(VI) spiked EBC sample began to show a peak at the retention time of EDTA–Cr(III) and this had increased in size when analysed at 4 weeks and again at 6 weeks showing a 6% recovery of EDTA–Cr(III) in an EBC Cr(VI) only sample whereas the pH 8 solution showed no EDTA–Cr(III) at 6 weeks (see Fig. 7). The pH of undiluted EBC was seen to range from pH 6–6.5 and, although the prepared EBC was diluted 1 in 10 with the EDTA solution adjusted to pH 7.6, the final pH of the dilution would be less than pH 7.6. Wolf *et al.*²⁷ determined that when the pH of their EDTA/TBAOH solution was less than pH 7.4 more than 10% of their Cr(VI) samples were present as HCrO_4^- and when it dropped to between pH 6 and pH 7 their Cr(VI) sample existed as HCrO_4^{2-} . Either the EBC Cr(VI) sample here is slowly reducing to Cr(III) in the neutral to acidic EDTA solution during storage or it is slowly converting to a different compound of Cr(VI) which has the same retention time as EDTA–Cr(III). It is neither appropriate nor practical to measure the pH of each person's EBC sample and adjust accordingly when work place monitoring, so an EDTA solution at pH 8 was chosen to compensate for any reduction in pH as a result of the EBC.

6. Optimised method. The chromatogram in Fig. 8 shows that this optimised method achieves full separation of EDTA–Cr(III) and Cr(VI) in 3 minutes (180 000 ms). The calibration range used in this study was 0–5 $\mu\text{g L}^{-1}$ for both species of chromium. The limit of detection (LOD) was calculated as three times the standard deviation of the blank; this was 0.006 $\mu\text{g L}^{-1}$ and 0.002 $\mu\text{g L}^{-1}$ for Cr(III) and Cr(VI) respectively. The limit of quantification (LOQ) was calculated as 10 times the mean background equivalent concentration (BEC); this was 0.040 $\mu\text{g L}^{-1}$ and 0.013 $\mu\text{g L}^{-1}$ for Cr(III) and Cr(VI) respectively.

The “blank” EBC spiked at a concentration of 5 $\mu\text{g L}^{-1}$, analysed on four separate occasions over the 6 week storage period, gave an average recovery of 91% \pm 0.13 for Cr(III) and 95% \pm 0.02 for Cr(VI). The proficiency testing material when prepared 1 in 1000 for this study has a target value of 2.81 $\mu\text{g L}^{-1}$ and an acceptance range of 2.28–3.28 $\mu\text{g L}^{-1}$. The PTM sample was analysed three times and gave an average within range result of 2.65 \pm 0.03 $\mu\text{g L}^{-1}$ for Cr(VI).

Calibration standards, samples and quality control material are diluted 10 fold with 0.5 mM EDTA solution adjusted to pH 8 with 10% v/v ammonia solution. These samples are left to stand a minimum of 1 hour prior to analysis $\mu\text{LC-ICP-MS}$, and can be stored refrigerated for up to 6 weeks.

The use of ICP-MS can result in spectroscopic interferences, *e.g.* with polyatomic interferences $^{40}\text{Ar}^{12}\text{C}^+$ and $^{36}\text{Ar}^{16}\text{O}^+$ on ^{52}Cr . However the optimised use of the collision cell will avoid most of these interferences. It is also possible that the sample matrix could create additional interferences given that element such as C, O and Cl are abundant in biological samples. The main contributor to a potential high background signal intensity on ^{52}Cr due to $^{40}\text{Ar}^{12}\text{C}^+$. Having injected a weak butanol solution with and without the EBC matrix, no peak or increase in background signal intensity was observed. The main chlorine based

interference on ^{52}Cr would be the $^{35}\text{Cl}^{16}\text{O}^+\text{H}^+$. To investigate the impact of a chloride interference on the chromium species a weak solution of HCl was injected onto the column. Prior to complexing Cr(III) with EDTA, Cl eluted before Cr(VI) and Cr(III). However, after complexing Cr(III) with EDTA and altering the retention time, when a weak solution of HCl is injected onto the column or added to ‘blank’ EBC, Cl and EDTA–Cr(III) elute at the same time. However, unlike a urine sample, the chloride content in EBC is negligible, as can be seen in the blank EBC in Fig. 8a. Furthermore the recovery of the spiked sample for Cr(III) in the stability study was 91% so no over recovery from an interference is evident. To confirm the absence of a chloride interference, two EBC samples were collected from five volunteers who were not occupationally exposed to chromium. There was no significant peaks obtained at Cr(III) in fact both EDTA–Cr(III) and Cr(VI) were less than the LOQ for all 10 EBC samples analysed.

Conclusion

The method described in this study is a robust and sensitive method for the simultaneous determination of Cr(III) and Cr(VI) in an EBC sample. This speciation method has been validated with spiked samples of blank EBC and a commercially available drinking water proficiency testing material sample for Cr(VI). It has been demonstrated that water samples and EBC samples containing Cr(III) and Cr(VI) can be stored in the fridge ($\sim 4^\circ\text{C}$) for up to 6 weeks when they have been diluted 10 fold with the EDTA solution, and that EBC samples must not be frozen after collection if the integrity of Cr(VI) is to be maintained. This study has developed a practical approach to collecting EBC samples in the workplace from workers who are potentially occupationally exposed to hexavalent chromium and that the integrity of the chromium species will be intact prior to analysis by $\mu\text{LC-ICP-MS}$. The next stage of this study will be to undertake site visits to collect EBC samples from chromium workers and apply the methods developed here to these ‘real’ samples.

Acknowledgements

This publication and the work it describes were funded by the Health and Safety Executive (HSE). Its contents, including any opinions and/or conclusions expressed, are those of the authors alone and do not necessarily reflect HSE policy.

References

- 1 G. F. Nordberg, B. A. Fowler, M. Nordberg and L. T. Friberg, *Handbook on the toxicology of metals*, Elsevier Academic Press, London, 3rd edn, 2007.
- 2 Agents classified by the IARC monographs, volumes 1–113, http://monographs.iarc.fr/ENG/Classification/latest_classif.php (Accessed September 2015).
- 3 R. Rakhunde, L. Deshpande and H. D. Juneja, *Crit. Rev. Environ. Sci. Technol.*, 2012, 42(7), 776–810.
- 4 IARC monographs on the evaluation of carcinogenic risks to humans, Volume 100 C. A review of human carcinogens:



- arsenic, metals, fibres and dusts. <http://monographs.iarc.fr/ENG/Monographs/vol100C/index.php> (accessed September 2015).
- 5 D. J. Paustenbach, B. L. Finley, F. S. Mowat and B. D. Kerger, *J. Toxicol. Environ. Health, Part A*, 2003, **66**(14), 1295–1339.
 - 6 F. S. De, D. Serra, A. Camoirano and P. Zancchi, *Biol. Trace Elem. Res.*, 1989, **21**, 179–187.
 - 7 P. Hoet, in *Handbook of elemental speciation II: Species in the environment, food, medicine and occupational health*, ed. R. Cornelis, J. Caruso, H. Crews and K. Heumann, Wiley-Blackwell, 2005, ch. 2.6.2, pp. 136–157.
 - 8 A. Zhitkovich, *Chem. Res. Toxicol.*, 2011, **24**(10), 1617–1629.
 - 9 HSL, *Guidance on laboratory techniques in occupational medicine*, HSE, 12th edn, 2013.
 - 10 DFG German DRF Deutsche Forschungsgemeinschaft, *List of MAK and BAT values 2009*, Wiley-VCH, Germany, 2012.
 - 11 American Conference of Governmental Industrial Hygienists, *TLVs and BEIs: Threshold limit values for chemical substances and physical agents*, Signature Publications, USA, 2015.
 - 12 G. E. Corbett, D. G. Dodge, E. O'Flaherty, J. Liang, L. Throop, B. L. Finley and B. D. Kerger, *Environ. Res.*, 1998, **78**(1), 7–11.
 - 13 J. P. Goulle, E. Saussereau, J. Grosjean, C. Doche, L. Mahieu, J. M. Thouret, M. Guerbet and C. Lacroix, *Forensic Sci. Int.*, 2012, **217**(1–3), 8–12.
 - 14 M. L. Chen, Y. N. Zhao, D. W. Zhang, Y. Tian and J. H. Wang, *J. Anal. At. Spectrom.*, 2010, **25**(11), 1688–1694.
 - 15 Y. M. Scindia, A. K. Pandey, A. R. Reddy and S. B. Manohar, *Anal. Chim. Acta*, 2004, **515**(2), 311–321.
 - 16 A. S. Stasinakis, N. S. Thomaidis and T. D. Lekkas, *Anal. Chim. Acta*, 2003, **478**(1), 119–127.
 - 17 L. Girard and J. Hubert, *Talanta*, 1996, **43**(11), 1965–1974.
 - 18 M. Corradi and A. Mutti, *Acta Biol. Med.*, 2005, **76**(2), 20–29.
 - 19 S. A. Kharitonov and P. J. Barnes, *Am. J. Respir. Crit. Care Med.*, 2001, **163**(7), 1693–1722.
 - 20 M. Goldoni, M. Corradi, P. Mozzoni, G. Folesani, R. Alinovi, S. Pinelli, R. Andreoli, D. Pignini, R. Tillo, A. Filetti, C. Garavelli and A. Mutti, *J. Breath Res.*, 2013, **7**(1), 017101.
 - 21 M. Goldoni, S. Catalani, G. De Palma, P. Manini, O. Acampa, M. Corradi, R. Bergonzi, P. Apostoli and A. Mutti, *Environ. Health Perspect.*, 2004, **112**(13), 1293–1298.
 - 22 H. C. Broding, B. Michalke, T. Goen and H. Drexler, *Int. Arch. Occup. Environ. Health*, 2009, **82**(5), 565–573.
 - 23 F. Hoffmeyer, T. Weiss, M. Lehnert, B. Pesch, H. Berresheim, J. Henry, M. Raulf-Heimsoth, C. H. Broding, J. Buenger, V. Harth and T. Bruening, *J. Environ. Monit.*, 2011, **13**(1), 212–218.
 - 24 A. Caglieri, M. Goldoni, O. Acampa, R. Andreoli, M. V. Vettori, M. Corradi, P. Apostoli and A. Mutti, *Environ. Health Perspect.*, 2006, **114**(4), 542–546.
 - 25 S. Hulo, N. Cherot-Kornobis, M. Howsam, S. Crucq, V. de Broucker, A. Sobaszek and J. L. Edme, *Toxicol. Lett.*, 2014, **226**(1), 63–69.
 - 26 M. Goldoni, A. Caglieri, D. Poli, M. V. Vettori, M. Corradi, P. Apostoli and A. Mutti, *Anal. Chim. Acta*, 2006, **562**(2), 229–235.
 - 27 R. E. Wolf, J. M. Morrison and M. B. Goldhaber, *J. Anal. At. Spectrom.*, 2007, **22**(9), 1051–1060.
 - 28 J. Scancar and R. Milacic, *J. Anal. At. Spectrom.*, 2014, **29**(3), 427–443.
 - 29 K. Neubauer, W. Reuter and P. Perrone, *Application Note*, Perkin Elmer Life and Analytical Sciences, USA, 2003.
 - 30 C. Puls and A. Limbeck, *J. Anal. At. Spectrom.*, 2009, **24**(10), 1434–1440.
 - 31 M. M. L. Guerrero, E. V. Alonso, J. M. C. Pavon, M. T. S. Cordero and A. G. de Torres, *J. Anal. At. Spectrom.*, 2012, **27**(4), 682–688.
 - 32 M. J. Marques, A. Morales-Rubio, A. Salvado and M. de la Guardia, *Talanta*, 2001, **53**(6), 1229–1239.
 - 33 J. Prokisch, S. A. Katz, B. Kovacs and Z. Gyori, *J. Chromatogr. A*, 1997, **774**(1–2), 363–371.
 - 34 R. P. Monasterio, G. E. Lascalea, L. D. Martinez and R. G. Wuilloud, *J. Trace Elem. Med. Biol.*, 2009, **23**(3), 157–166.
 - 35 M. Sperling, S. K. Xu and B. Welz, *Anal. Chem.*, 1992, **64**(24), 3101–3108.
 - 36 B. H. Li and X. P. Yan, *Electrophoresis*, 2007, **28**(9), 1393–1398.
 - 37 Z. Chen, M. Megharaj and R. Naidu, *Talanta*, 2007, **72**(2), 394–400.
 - 38 M. Panssar-Kallio and P. Manninen, *Anal. Chim. Acta*, 1996, **318**(3), 335–343.
 - 39 F. A. Byrdy, L. K. Olson, N. P. Vela and J. A. Caruso, *J. Chromatogr. A*, 1995, **712**(2), 311–320.
 - 40 H. Gurleyuk and D. Wallschlagel, *J. Anal. At. Spectrom.*, 2001, **16**(9), 926–930.
 - 41 Y. Suzuki and F. Serita, *Ind. Health.*, 1985, **23**(3), 207–220.
 - 42 F. Seby, S. Charles, M. Gagean, H. Garraud and O. F. X. Donard, *J. Anal. At. Spectrom.*, 2003, **18**(11), 1386–1390.
 - 43 C. Barnowski, N. Jakubowski, D. Stuewer and J. A. C. Broekaert, *J. Anal. At. Spectrom.*, 1997, **12**(10), 1155–1161.
 - 44 L. Xing and D. Beauchemin, *J. Anal. At. Spectrom.*, 2010, **25**(7), 1046–1055.
 - 45 M. Goldoni, A. Caglieri, G. De Palma, O. Acampa, P. Gergelova, M. Corradi, P. Apostoli and A. Mutti, *J. Environ. Monit.*, 2010, **12**(2), 442–447.
 - 46 M. Gube, J. Ebel, P. Brand, T. Goen, K. Holzinger, U. Reisgen and T. Kraus, *Int. Arch. Occup. Environ. Health*, 2010, **83**(7), 803–811.
 - 47 M. Leist, R. Leiser and A. Toms, *The Application Notebook-Advertising Supplement Mass Spectrometry*, Varian Inc., 2006, pp. 29–31.

